Package: omicsViewer (via r-universe)

November 1, 2026

Title Interactive and explorative visualization of SummarizedExperssionSet or ExpressionSet using omicsViewer

Version 1.17.0

Description omicsViewer visualizes ExpressionSet (or SummarizedExperiment) in an interactive way. The omicsViewer has a separate back- and front-end. In the back-end, users need to prepare an ExpressionSet that contains all the necessary information for the downstream data interpretation. Some extra requirements on the headers of phenotype data or feature data are imposed so that the provided information can be clearly recognized by the front-end, at the same time, keep a minimum modification on the existing ExpressionSet object. The pure dependency on R/Bioconductor guarantees maximum flexibility in the statistical analysis in the back-end. Once the ExpressionSet is prepared, it can be visualized using the front-end, implemented by shiny and plotly. Both features and samples could be selected from (data) tables or graphs (scatter plot/heatmap). Different types of analyses, such as enrichment analysis (using Bioconductor package fgsea or fisher's exact test) and STRING network analysis, will be performed on the fly and the results are visualized simultaneously. When a subset of samples and a phenotype variable is selected, a significance test on means (t-test or ranked based test; when phenotype variable is quantitative) or test of independence (chi-square or fisher's exact test; when phenotype data is categorical) will be performed to test the association between the phenotype of interest with the selected samples. Additionally, other analyses can be easily added as extra shiny modules. Therefore, omicsViewer will greatly facilitate data exploration, many different hypotheses can be explored in a short time without the need for knowledge of R. In addition, the resulting data could be easily shared using a shiny server. Otherwise, a standalone version of omicsViewer together with designated omics data could be easily created by integrating it with portable R, which can be shared with collaborators or submitted as supplementary data together with a manuscript.

Depends R (>= 4.2)

License GPL-2

Imports htmltools, shinydashboard, survminer, survival, fastmatch, reshape2, stringr, beeswarm, grDevices, DT, shiny, shinythemes, shinyWidgets, plotly, networkD3, httr, matrixStats, RColorBrewer, Biobase, fgsea, openxlsx, psych, shinybusy, ggseqlogo, htmlwidgets, graphics, grid, stats, utils, methods, shinyjs, curl, flatxml, ggplot2, S4Vectors, SummarizedExperiment, RSQLite, Matrix, shinycssloaders, ROCR, drc

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URL https://github.com/mengchen18/omicsViewer

Video https://www.youtube.com/watch?v=0nirB-exquY&list=PLo2m88lJf-RRoLKMY8UEGqCpraKYrX5lk

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`.e2EC50` *convert e (inflection point) to EC50*

Description

convert e (inflection point) to EC50

Usage

```
.e2EC50(b, d, e, f)
```

Arguments

`b`	Hill's slope. The Hill's slope refers to the steepness of the curve. It could either be positive or negative.
`d`	Hihgest response value.
`e`	Inflection point. The inflection point is defined as the point on the curve where the curvature changes direction or signs. In models where f = 1 (2-4 parameter models), e is EC50.
`f`	Asymmetry factor. When f=1 we have a symmetrical curve around inflection point and so we have a four-parameters logistic equation.

Note

Only has an effect when using LL.5 and LL2.5 model

`.modelFormula` *model fitted by drc*

Description

model fitted by drc

Usage

```
.modelFormula(x, b, c = 0, d = 1, e, f = 1)
```

Arguments

x	numerical vector of doses/time points/concentrations
b	Hill's slope. The Hill's slope refers to the steepness of the curve. It could either be positive or negative.
c	Lowest response value.
d	Hihgest response value.
e	Inflection point. The inflection point is defined as the point on the curve where the curvature changes direction or signs. In models where f = 1 (2-4 parameter models), e is EC50.
f	Asymmetry factor. When f=1 we have a symmetrical curve around inflection point and so we have a four-parameters logistic equation.

Details

func(x) = c + (d - c) / (1 + (x/e)^b)^f

---

asEsetWithAttr *Convert SummarizedExperiment to ExpressionSet retaining all attributes*

Description

Convert SummarizedExperiment to ExpressionSet retaining all attributes

Usage

```
asEsetWithAttr(x)
```

Arguments

x	an object of class SummarizedExperiment

Value

an object of class ExpressionSet

`correlationAnalysis` *Correlating a expression matrix with phenotypical variables*

Description

This is a convenience function to perform correlation analysis, the output is in a format ready to be incorporated into object to be visualized by `omicsViewer`.

Usage

```
correlationAnalysis(
  x,
  pheno,
  min.value = MIN_SAMPLES_CORRELATION,
  prefix = "Cor"
)
```

Arguments

`x`	an expression matrix, rows are the features (e.g. proteins), columns are the samples
`pheno`	a `data.frame` storing the numerical phenotypical variable to be correlated with the rows (features) in expression matrix.
`min.value`	the minimum number of samples required in the correlation analysis, if lower than this number, NA will be returned.
`prefix`	prefix of the names. Usually don't need to be changed by the user. When changes are needed, the prefix should be in a format like [analysis name]|[subset] so the "analysis name" and "subset" can be selected in the `omicsViewer`.

Value

Every correlation analysis returns a `data.frame` with five columns: `R` - pearson correlation coefficient `N` - number of values used in the analysis `P` - p-values returned by pearson correlation analysis `logP` - log transformed p-values `range` - the range of values in expression matrix used in the analysis

Examples

```
e1 <- matrix(rnorm(500), 50, 10)
rownames(e1) <- paste0("FT", 1:50)
p1 <- matrix(rnorm(50), 10, 5)
colnames(p1) <- paste0("PH", 1:5)
colnames(e1) <- rownames(p1) <- paste0("S", 1:10)
correlationAnalysis(x = e1, pheno = p1, min.value = 8)
```

csc2list *convert a column compressed sparse matrix to a list*

Description

convert a column compressed sparse matrix to a list

Usage

```
csc2list(x)
```

Arguments

x	a matrix or CsparseMatrix object

Value

a sparse frame in data.frame

draw_roc_pr *Drawing ROC and PR curve*

Description

Drawing ROC and PR curve

Usage

```
draw_roc_pr(value, label)
```

Arguments

value	a numerical vector indicates the predictions
label	true class labels, could be two or more unique values

Examples

```
v <- sort(rnorm(100))
l <- sample(1:2, size = 100, replace = TRUE)
draw_roc_pr(v, l)
l <- rep(c("b", "c", "a", "d"), each = 25)
draw_roc_pr(v, l)
draw_roc_pr(v, sample(l))
```

drmMat *Fitting dose-response models for omics data matrix*

Description

A convenient function to fit dose response models for every row in an omics matrix using `drm` function in the `drc` package.

Usage

```
drmMat(
  x,
  fitvar,
  fitvar.name = c("Dose", "Time", "Concentration")[1],
  curveid = NA,
  fct.name = c("LL.4()", "LL.3()", "LL.2()", "LL.5()")[1]
)
```

Arguments

`x`	a numerical matrix where the rows are features and columns are samples.
`fitvar`	a numerical variable has the same length as `ncol(x)` to indicate the dose/time/concentration conditions.
`fitvar.name`	the name of the `fitvar`, a length one character. Will be used as the label for x-axis when drawing the dose curve.
`curveid`	a numeric vector or factor containing the grouping of the columns in `x`.
`fct.name`	the function name, e.g. "LL.4()", "LL.3()", "LL.2()" and "LL.5()", which are defined in the `drc` package.

Value

a list of `drc` object

exprspca *Perform PCA and prepare results for* `omicsViewer`

Description

This is a convenience function to perform PCA on expression matrix, the output of PCA will be in a format ready to be incorporated into object to be visualized by `omicsViewer`.

Usage

```
exprspca(x, n = min(8, ncol(x) - 1), prefix = "PCA|All", fillNA = FALSE, ...)
```

Arguments

x	an expression matrix, where rows are features and samples are on columns.
n	number of components to keep
prefix	prefix of the names. Usually don't need to be changed by the user. When changes are needed, the prefix should be in a format like [analysis name]|[subset] so the "analysis name" and "subset" can be selected in the omicsViewer.
fillNA	logical; whether NA should be filled? If FALSE (default), na.omit will be called before PCA. If TRUE, the missing value will be replaced using fillNA.
...	other parameters passed to prcomp

Value

a data.frame storing the PCA results

Examples

```
# reading expression
packdir <- system.file("extdata", package = "omicsViewer")
expr <- read.delim(file.path(packdir, "expressionMatrix.tsv"), stringsAsFactors = FALSE)
# call PCA
pc <- exprspca(expr)
head(pc$samples)
head(pc$features)
```

---

extendMetaData *Add extra columns to the phenoData/colData or featureData/rowData in ExpressionSet/SummarizedExperiment*

Description

Add extra columns to the phenoData/colData or featureData/rowData in ExpressionSet/SummarizedExperiment

Add extra columns to the phenoData/colData or featureData/rowData in ExpressionSet/SummarizedExperiment

Add extra columns to the phenoData/colData or featureData/rowData in ExpressionSet/SummarizedExperiment

Usage

```
extendMetaData(object, newData, where)

## S4 method for signature 'ExpressionSet,data.frame'
extendMetaData(
  object,
  newData,
  where = c("pData", "fData", "colData", "rowData")[1]
)
```

```
## S4 method for signature 'SummarizedExperiment,data.frame'
extendMetaData(
  object,
  newData,
  where = c("pData", "fData", "colData", "rowData")[1]
)

## S4 method for signature 'SummarizedExperiment,DFrame'
extendMetaData(
  object,
  newData,
  where = c("pData", "fData", "colData", "rowData")[1]
)
```

Arguments

object	an object of ExpressionSet-class
newData	a data.frame containing the data to be added
where	where to add the extra columns, should be one of "pData", "fData", "rowData" and "colData".

Value

an object of ExpressionSet-class

Note

The attributes in the pheno data and feature data will be preserved

Examples

```
est <- Biobase::ExpressionSet(assayData=matrix(runif(1000), nrow=100, ncol=10))
Biobase::pData(est)
est <- extendMetaData(est, data.frame(letter = letters[1:10]), where = "pData")
Biobase::pData(est)
```

---

extractParamDC *Extracting parameters from drc models*

Description

Extracting parameters from drc models

Usage

```
extractParamDC(mod, prefix = "ResponseCurve")
```

Arguments

mod	a drc object
prefix	for column header, the column will be named as prefix|curveid|curveparameter

Note

when LL2.X is used, e is estimated as log(e), this function will return e in linear scale instead.

---

extractParamDCList *Extracting parameter from a list of drc object*

Description

Extracting parameter from a list of drc object and return a data.frame, which can be incorporated into the object visualized by omicsViewer

Usage

```
extractParamDCList(x, prefix = "ResponseCurve")
```

Arguments

x	a list of drc object
prefix	for column header

Value

a data.frame

---

fgsea1 *Wrapper of fgseaMultilevel function to take binary gene set matrix as input*

Description

Wrapper of fgseaMultilevel function to take binary gene set matrix as input

Usage

```
fgsea1(gs, stats, gs_desc = NULL, ...)
```

Arguments

gs	either a data.frame or a (sparse) matrix input. If a data.frame object is given, it should have at least three columns named as "featureId", "gsId" and "weight". If a matrix is given, the matrix is binary matrix where rows are features and columns are gene sets. The values in the matrix should be either 1 or 0 representing the presence and absence of a feature in the genesets, repectively.
stats	ranking stats
gs_desc	description of gene sets, it should be a named vector and the names should be the same as colnames(gs)
...	other parameters passed to fgseaMultilevel

Value

a `data.frame` of fgsea results

Examples

```
## not for users
# library(fgsea)
# library(Biobase)
# dat <- readRDS(system.file(package = "omicsViewer", "extdata/demo.RDS"))
# fd <- fData(dat)
# fdgs <- fd[, grep("^GS\|", colnames(fd))]
# res <- fgsea1(fdgs, stats = fd$`t-test|OV_BR|md`, minSize = 5, maxSize = 500)
# res <- fgsea1(
#   fdgs, stats = fd$`t-test|OV_BR|md`,
#   minSize = 5, maxSize = 500, gs_desc = colnames(fdgs))
```

---

fillNA *Filling NAs in a matrix using constants calculated from user the defined function*

Description

This function is usually use to impute missing values in expression matrix, where the rows are feature and columns are samples. This function impute the missing values on the row-wise, that is, every row will be imputed using different constant.

Usage

```
fillNA(
  x,
  maxfill = quantile(x, probs = 0.15, na.rm = TRUE),
  fillingFun = function(x) min(x, na.rm = TRUE) - log10(2)
)
```

Arguments

x	a matrix with NA values
maxfill	the maximum filled value, if the value calculated by fillingFun is greater than maxfill, then maxfill will the used to replace NAs.
fillingFun	function to calculate the filling values. It should be a function accept at least one argument "x", which is a row of input expression matrix. The default is function(x) min(x, na.rm = TRUE) - log10(2) corresponds to the "half of lowest detected values" if the expression matrix is log10 transformed. More examples:#' function(x) min(x, na.rm = TRUE) - 1 # half of lowest detected value when expression matrix is in log2 scale function(x) 0 # replace NA by 0

Value

a matrix without NAs

Note

The returned matrix may have -Inf, which may need to be filtered/replaced additionally

Examples

```
m <- matrix(rnorm(200), 20, 10)
m[sample(1:200, size = 20)] <- NA
mf <- fillNA(m)
```

---

filterRow *Filter out rows of expression matrix*

Description

The function is used to filter rows with values of low intensities or do not reproducible presented in replicates.

Usage

```
filterRow(x, max.quantile = NULL, max.value = NULL, var = NULL, min.rep = 2)
```

Arguments

x	an expression matrix
max.quantile	a single numerical value between (0, 1), if the row maximum is smaller than this quantile (calculated from the whole matrix), the row will be removed.
max.value	a single numerical value, if the the maximum value of a rwo is smaller than this value, the row will be removed. Only used if max.quantile is set to "NULL".

var	variables has the same length as the column number in x to indicate which sample is from which group
min.rep	the minimum number of replicate in at least one of the groups, if less than this value, the row will be removed.

Value

a logical vector where the TRUE means row to keep

Examples

```
e1 <- matrix(rnorm(5000, sd = 0.3), 500, 10) + rnorm(500)
f <- filterRow(x = e1, max.quantile = 0.25)
table(f)
```

---

getAutoRIF *Get genes associated with search terms and AutoRIF annotations*

Description

Get genes associated with search terms and AutoRIF annotations

Usage

```
getAutoRIF(term, rif = c("generif", "autorif")[1], filter = TRUE)
```

Arguments

term	a character vector of terms want to search
rif	either autorif or generif, see "https://maayanlab.cloud/geneshot/"
filter	whether the result should be filtered. The least frequently mentioned genes (most like 1 or 2 times) will be removed.

Value

a data.frame of 4 columns: gene, n, perc, rank.

Note

https://amp.pharm.mssm.edu/geneshot/

References

Alexander Lachmann, Brian M Schilder, Megan L Wojciechowicz, Denis Torre, Maxim V Kuleshov, Alexandra B Keenan, Avi Ma'ayan, Geneshot: search engine for ranking genes from arbitrary text queries, Nucleic Acids Research, Volume 47, Issue W1, 02 July 2019, Pages W571–W577, https://doi.org/10.1093/nar/gkz393

Alexander Lachmann, Brian M Schilder, Megan L Wojciechowicz, Denis Torre, Maxim V Kuleshov, Alexandra B Keenan, Avi Ma'ayan, Geneshot: search engine for ranking genes from arbitrary text queries, Nucleic Acids Research, Volume 47, Issue W1, 02 July 2019, Pages W571–W577, https://doi.org/10.1093/nar/gkz393

Examples

```
a <- getAutoRIF("mtor signaling")
```

`getMQParams` *Parse mqpar.xml file*

Description

Getting the experimental informatione (TMT or label free) from mqpar.xml file.

Usage

```
getMQParams(x)
```

Arguments

`x`	the path to mqpar.xml file

Value

a list of MQ paramters

`getStringId` *Mapping ids to string ids*

Description

the string ids can be used as background in the string enrichment analysis

Usage

```
getStringId(genes, taxid = 9606, caller = "omicsViewer")
```

Arguments

genes	a character vector of gene ids
taxid	taxonomy ids
caller	your identifier for string-db.org

Note

https://string-db.org/cgi/help.pl?subpage=api

Examples

```
1
# gg = c('P04637', 'P00533', 'P04626', "Q8IYB3", "O75494", "Q9Y696")
# getStringId(gg)
```

---

getUPRefProteomeID *get uniprot reference proteome IDs*

Description

get uniprot reference proteome IDs

get uniprot reference proteome IDs

Usage

```
getUPRefProteomeID(
  domain = c("Eukaryota", "Archaea", "Bacteria", "Viruses")[1]
)

downloadUPRefProteome(
  id,
  domain = c("Eukaryota", "Archaea", "Bacteria", "Viruses")[1],
  destdir = "./"
)
```

Arguments

domain	the domain, one of "Eukaryota", "Archaea", "Bacteria" or "Viruses"
id	the UP id to download
destdir	destination directory

Value

a character vector of UP ids

a character vector of UP ids

Functions

- `getUPRefProteomeID()`: get uniprot reference protein IDs

`gsAnnotIdList` *Annotation of gene/protein function using multiple IDs.*

Description

Annotation of gene/protein function using multiple IDs.

Usage

```
gsAnnotIdList(
  idList,
  gsIdMap,
  minSize = 5,
  maxSize = 500,
  data.frame = FALSE,
  sparse = TRUE
)
```

Arguments

`idList`	list of protein IDs, e.g. list(c("ID1", "ID2"), c("ID13"), c("ID4", "ID8", "ID10"))
`gsIdMap`	a data frame for geneset to id map, it has two columns - id: the ID column - term: annotation terms e.g. gsIdMap <- data.frame(id = c("ID1", "ID2", "ID1", "ID2", "ID8", "ID10"), term = c("T1", "T1", "T2", "T2", "T2", "T2"), stringsAsFactors = FALSE)
`minSize`	minimum size of gene sets
`maxSize`	maximum size of gene sets
`data.frame`	logical; whether to organize the result into `data.frame` format, see "Value" section.
`sparse`	logical; whether to return a sparse matrix, only used when data.frame=FALSE

Value

A binary matrix (if `data.frame` = FALSE), the number of rows is the same with length of idList, the columns are the annotated gene set; or a `data.frame` (if `data.frame` = TRUE) with three columns: featureId, gsId, weight.

Examples

```
terms <- data.frame(
  id = c("ID1", "ID2", "ID1", "ID2", "ID8", "ID10"),
 term = c("T1", "T1", "T2", "T2", "T2", "T2"),
  stringsAsFactors = FALSE
)
features <- list(c("ID1", "ID2"), c("ID13"), c("ID4", "ID8", "ID10"))
gsAnnotIdList(idList = features, gsIdMap = terms, minSize = 1, maxSize = 500)

terms <- data.frame(
id = c("ID1", "ID2", "ID1", "ID2", "ID8", "ID10", "ID4", "ID4"),
term = c("T1", "T1", "T2", "T2", "T2", "T2", "T1", "T2"),
stringsAsFactors = FALSE
)
features <- list(F1 = c("ID1", "ID2", "ID4"), F2 = c("ID13"), F3 = c("ID4", "ID8", "ID10"))
gsAnnotIdList(features, gsIdMap = terms, data.frame = TRUE, minSize = 1)
gsAnnotIdList(features, gsIdMap = terms, data.frame = FALSE, minSize = 1)
```

---

hasAttr *Check whether an object has an attribute*

Description

Check whether an object has an attribute

Usage

```
hasAttr(x, attr.name)
```

Arguments

x	the object
attr.name	a character vector containing the name of attributes to be checked

Value

a logical value/vector has the same length as attr.name

`hclust2str` *Convert hclust object to/from single character*

Description

Convert hclust object to/from single character

Usage

```
hclust2str(x)

str2hclust(x)
```

Arguments

`x`	a character of length one or an hclust object

Value

a character stores the hclust object

a hclust object

Note

The $call element in hclust will not retained in the conversion. The conversion decrease the precision in $height element.

Examples

```
# not for end users
# m <- matrix(rnorm(50), 25)
# hc <- hclust(dist(m))
# plot(hc)
# te <- hclust2str(hc)
# hc2 <- str2hclust(te)
# plot(hc2)
```

jaccardList *Calculate Jaccard distance from a list*

Description

Calculate Jaccard distance from a list

Usage

```
jaccardList(x)
```

Arguments

x	a list

Value

an dist object

L1_data_space_module *Application level 1 module - data space*

Description

Application level 1 module - data space

Usage

```
L1_data_space_module(
  id,
  expr,
  pdata,
  fdata,
  reactive_x_s = reactive(NULL),
  reactive_y_s = reactive(NULL),
  reactive_x_f = reactive(NULL),
  reactive_y_f = reactive(NULL),
  cormat = reactive(NULL),
  status = reactive(NULL)
)
```

Arguments

id	module id
expr	reactive value; expression matrix
pdata	reactive value; phenotype data
fdata	reactive value; feature data
reactive_x_s	pre-selected x axis for sample space
reactive_y_s	pre-selected y axis for sample space
reactive_x_f	pre-selected x axis for feature space
reactive_y_f	pre-selected y axis for feature space
cormat	reactive value; correlation matrix. if not given, calculated on the fly.
status	intial status

L1_data_space_ui *Application level 1 ui - data space*

Description

Application level 1 ui - data space

Usage

```
L1_data_space_ui(id, activeTab = "Feature")
```

Arguments

id	id
activeTab	one of "Feature", "Feature table", "Sample", "Sample table", "Heatmap"

L1_result_space_module *Utility L1 result space ui*

Description

Utility L1 result space ui

Usage

```
L1_result_space_module(
  id,
  reactive_expr,
  reactive_phenoData,
  reactive_featureData,
  reactive_i = reactive(NULL),
  reactive_highlight = reactive(NULL),
  additionalTabs = NULL,
  object = NULL,
  status = reactive(NULL)
)
```

Arguments

id	module id
reactive_expr	expression matrix
reactive_phenoData	phentype data
reactive_featureData	feature data
reactive_i	row ID/name of rows selected
reactive_highlight	col ID/name of columns selected
additionalTabs	additional tabs added to "Analyst" panel
object	originally loaded object, mostly an ExpressionSet or SummarizedExperiment object
status	intial status

L1_result_space_ui *Utility L1 result space ui*

Description

Utility L1 result space ui

Usage

```
L1_result_space_ui(id)
```

Arguments

id	id

list2csc *convert a list to column compressed sparse matrix*

Description

convert a list to column compressed sparse matrix

Usage

```
list2csc(l, dimnames)
```

Arguments

l	a data.frame with at least two columns - featureId, gsId; optionally a "weight" column.
dimnames	a list of dimnames, should contain at least one element for the row names.

Value

a sparse matrix, CsparseMatrix, column compressed

meta_scatter_module *Utility - scatter plot for meta shiny module*

Description

Utility - scatter plot for meta shiny module

Usage

```
meta_scatter_module(
  id,
  reactive_meta = reactive(NULL),
  reactive_expr = reactive(NULL),
  combine = c("pheno", "feature"),
  source = "plotlyscattersource",
  reactive_x = reactive(NULL),
  reactive_y = reactive(NULL),
  reactive_status = reactive(NULL)
)
```

Arguments

id	Character. Namespace ID for the Shiny module.
reactive_meta	reactive meta data, phenotype data or feature data
reactive_expr	reactive expression data
combine	how to combine the expression and meta data, pheno or feature?
source	source id for plotly object
reactive_x	reactive value for pre-selected x-aixs
reactive_y	reactive value for pre-selected y-aixs
reactive_status	the status of scatter plot, e.g. x-, y-axis, color variable, shape variable, etc.

---

multi.t.test *Function to perform multiple t-tests on an expression matrix*

Description

This is a convenience function to perform multiple student's t-test. The output is in a format ready to be incorporated into object to be visualized by omicsViewer. This function use t.test.

Usage

```
multi.t.test(x, pheno, compare = NULL, fillNA = FALSE, ...)
```

Arguments

x	an expression matrix, usually log10 transformed.
pheno	phenotype data of x, the number of rows in pheno must equal the number of columns of x. Please refer to examples for more details.
compare	NULL or a matrix with three columns to define the comparisons to do. When a matrix is given, the first column should be one of the column headers in pheno; then the second and third columns should be two values presented (more than once) in the columns of pheno selected by the values in the first column. The samples mapped to the two values are compared. If paired comparisons to be done, the orders of samples should be mapped
fillNA	logical; whether NA should be filled? If FALSE (default), t test will be performed whenever possible. If not possible, then NA will be returned. If TRUE, the missing value will be replaced using fillNA.
...	other parameters passed to t.test

Value

a data.frame stores the t-test results with the follow columns: mean|[selected header in pheno]|[group 1 in test] - The mean value of group 1 n value|[selected header in pheno]|[group 1 in test] - The number of value used in the test for group 1 quantile|[selected header in pheno]|[group 1 in test] - The quantile of means values in group 1 mean|[selected header in pheno]|[group 2 in test] - The mean value of group 2 n value|[selected header in pheno]|[group 2 in test] - The number of value used in the test for group 2 quantile|[selected header in pheno]|[group 2 in test] - The quantile of means values in group 2 ttest|[group 1 in test]_vs_[group 2 in test]|pvalue - The p-value return by t.test ttest|[group 1 in test]_vs_[group 2 in test]|log.pvalue - The -log10 transformed p-value ttest|[group 1 in test]_vs_[group 2 in test]|fdr - The BH method corrected p-values, e.g. FDR ttest|[group 1 in test]_vs_[group 2 in test]|log.fdr - The -log10 transformed FDR ttest|[group 1 in test]_vs_[group 2 in test]|mean.diff - The difference between the means of the two groups, e.g. fold change

Examples

```
# reading expression
packdir <- system.file("extdata", package = "omicsViewer")
expr <- read.delim(file.path(packdir, "expressionMatrix.tsv"), stringsAsFactors = FALSE)
# reading phenotype data
pd <- read.delim(file.path(packdir, "sampleGeneral.tsv"), stringsAsFactors = FALSE)

## Single t-test
head(pd)
# define comparisons
tests <- c("Origin", "RE", "ME")
tres <- multi.t.test(x = expr, pheno = pd, compare = tests)

## multiple t-test
head(pd)
# define comparisons
tests <- rbind(
c("Origin", "RE", "ME"),
c("Origin", "RE", "LE"),
c('TP53.Status', "MT", "WT")
)
tres <- multi.t.test(x = expr, pheno = pd, compare = tests)
```

---

nColors *Generating k distinct colors*

Description

Mainly used in the shiny app to generate reproducible k distinct colors.

Usage

```
nColors(k, stop = FALSE)
```

Arguments

k	a number between 1 to 60 tells how many distinct colors to use
stop	logical; whether the function should return an error message if k is not in the range of 2 to 60. Default FALSE, the function will return NULL.

Value

a vector of hex code for k colors or NULL

Examples

```
nColors(5)
nColors(1, stop = FALSE)
```

---

normalize.nQuantiles *Normalization using n quantiles*

Description

Normalization using n quantiles

Usage

```
normalize.nQuantiles(x, probs = 0.5, shareFeature = FALSE, ref = 1)
```

Arguments

x	an expression matrix, usually log transformed
probs	the quantiles to be aligned across samples. If probs is a length 1 numerical vector, the quantiles will aligned. As a special case, probs = 0.5 equals the median centering. If probs' length is > 1, a shift and scaling factor of samples will be calculating by fitting linear models using quantiles of samples, the median and variance of samples will be corrected using the intersect and slope of the fitted model.
shareFeature	logocal; if TRUE, the normalization will be based on the shared features between samples
ref	the columns name or index to specify the reference sample, only used when shareFeature = TRUE

Value

a normalized matrix

Examples

```
e1 <- matrix(rnorm(5000), 500, 10)
e1[, 6:10] <- 0.3 *e1[, 6:10] + 3
boxplot(e1)
# median centering, no variance correction
e2 <- normalize.nQuantiles(x = e1, probs = 0.5)
boxplot(e2)
# median centering + variance stablization
e3 <- normalize.nQuantiles(x = e1, probs = seq(0.25, 0.75, by = 0.1))
boxplot(e3)
```

`normalize.totsum` *Normalize total sum*

Description

Normalize total sum

Usage

```
normalize.totsum(x)
```

Arguments

x	a log10 transformed expression matrix

Value

a normalized matrix

Examples

```
e1 <- matrix(rnorm(5000), 500, 10)
e1[, 6:10] <- e1[, 6:10]+3
boxplot(e1)
e2 <- normalize.totsum(x = e1)
boxplot(e2)
```

normalizeColWise *Column-wise normalization of expression matrix*

Description

A wrapper function of all column-wise normalization methods

Usage

```
normalizeColWise(
  x,
  method = c("Median centering", "Median centering (shared ID)", "Total sum",
    "median centering + variance stablization")[1]
)
```

Arguments

x	an expression matrix where rows are features and columns are samples, usually log transformed.
method	normalization method to use "Median centering" - median centering, see normalize.nQuantiles "Median centering (shared ID)" - median centering using shared features, see normalize.nQuantiles "Total sum" - total sum normalization "median centering + variance stablization" - 10 quantile normalization using 0.25, 0.3, ..., 0.75, see normalize.nQuantiles

Value

a normalized matrix

Examples

```
e1 <- matrix(rnorm(5000), 100, 50)+10
boxplot(e1)
e2 <- normalizeColWise(x = e1, method = "Median centering")
boxplot(e2)
```

normalizeData *Normalized expression matrix*

Description

A wrapper function of all normalization methods, including row-wise or column-wise normalization.

Usage

```
normalizeData(
  x,
 colWise = c("None", "Median centering", "Median centering (shared ID)", "Total sum",
    "median centering + variance stablization")[1],
  rowWise = c("None", "Reference", "Batch mean", "Batch reference")[1],
  ref = NULL,
  batch = NULL
)
```

Arguments

x	an expression matrix where rows are features and columns are samples, usually log transformed.
colWise	column-wise normalization method to use, see normalizeColWise
rowWise	row-wise normalization method to used Reference - using removeVarQC method Batch mean - using rowshift method without reference samples Batch reference - using rowshift method with reference samples
ref	index of reference samples
batch	batch factor

Value

a normalized matrix

Examples

```
e1 <- matrix(rnorm(5000), 100, 50)+10
boxplot(e1)
e2 <- normalizeData(x = e1, ref = seq(5, 45, by = 10), rowWise = "Reference")
boxplot(e2)
```

---

omicsViewer *Launch the omicsViewer Shiny Application*

Description

Starts an interactive Shiny application for exploring omics data, including visualization of expression matrices, feature and sample metadata, statistical analyses, and functional enrichment results. The viewer supports both ExpressionSet and SummarizedExperiment objects.

Usage

```
omicsViewer(
  dir,
  additionalTabs = NULL,
  filePattern = ".(RDS|DB|SQLITE|SQLITE3)$",
  ESVObj = NULL,
  esetLoader = readESVObj,
  exprsGetter = getExprs,
  pDataGetter = getPData,
  fDataGetter = getFData,
  defaultAxisGetter = getAx,
  appName = "omicsViewer",
  appVersion = packageVersion("omicsViewer")
)
```

Arguments

dir	Character. Path to directory containing the ExpressionSet or SummarizedExperiment object saved as .RDS file. Provide only the directory path, not the full file path. The viewer will list all compatible files in this directory.
additionalTabs	List. Optional custom Shiny modules to add as tabs in the "Analyst" panel. Each element should be a list with: tabName (character), moduleName (character), moduleUi (UI function), and moduleServer (server function).
filePattern	Character. Regular expression pattern to filter files displayed in the directory. Default: ".(RDS|DB|SQLITE|SQLITE3)$" (case-insensitive).
ESVObj	ExpressionSet or SummarizedExperiment. Optional pre-loaded object to view directly without file selection. If provided, the file dropdown will show "ESVObj.RDS".
esetLoader	Function. Custom loader for reading saved objects. Default: readESVObj. Should accept a file path and return an ExpressionSet or SummarizedExperiment object.
exprsGetter	Function. Extracts expression matrix from loaded object. Default: getExprs. Should return a numeric matrix.
pDataGetter	Function. Extracts phenotype/sample metadata. Default: getPData. Should return a data.frame with rownames matching sample names.
fDataGetter	Function. Extracts feature metadata. Default: getFData. Should return a data.frame with rownames matching feature names.
defaultAxisGetter	Function. Determines default axes for plots. Takes two arguments: x (the loaded object) and what (one of "sx", "sy", "fx", "fy" for sample/feature space x/y axes). Should return column name from metadata.
appName	Character. Application title displayed in the UI. Default: "omicsViewer".
appVersion	Character or package_version. Version number displayed in UI. Default: current package version.

Value

NULL (invisibly). Launches the Shiny application. The app runs until stopped by the user.

See Also

prepOmicsViewer for preparing data objects for visualization. app_module for the main application module (developers only).

Examples

```
if (interactive()) {
  # Basic usage with example data
  omicsViewer(system.file("extdata", package = "omicsViewer"))

  # With pre-loaded object
  packdir <- system.file("extdata", package = "omicsViewer")
  eset <- readRDS(file.path(packdir, "exampleEset.RDS"))
  omicsViewer(packdir, ESVObj = eset)
}
```

---

parseDatTerm *Extract function annotation from uniprot .dat file*

Description

Extract function annotation from uniprot .dat file

Usage

```
parseDatTerm(file, outputDir = NULL, ...)
```

Arguments

file	the .dat or .dat.gz file
outputDir	dir of output file
...	other parameters passed to readLines

Value

a data.frame parse from .dat file

plot_roc_pr_module *Shiny module for ROC/PR plot - Module*

Description

Shiny module for ROC/PR plot - Module

Usage

```
plot_roc_pr_module(id, reactive_param, reactive_checkpoint = reactive(TRUE))
```

Arguments

id	module id
reactive_param	reactive value; argument pass to draw_roc_pr
reactive_checkpoint	reactive_value; check this value before render any plot/executing any calculation

Value

do not return any values

Examples

```
if (interactive()) {
  library(shiny)

  ui <- fluidPage(
    sliderInput("ngrp", label = "Number of groups", min = 2, max = 5, value = 2),
    plot_roc_pr_ui("testplot")
  )

  server <- function(input, output, session) {
    ng <- reactive(
      sample(letters[1:input$ngrp], size = 100, replace = TRUE)
    )
    plot_roc_pr_module("testplot",
      reactive_param = reactive(list(
        x = ng(),
        y = rnorm(100)
      ))
    )
  }
 shinyApp(ui, server)
}
```

plotDC *Draw dose-response curves*

Description

Draw dose-response curves

Usage

```
plotDC(mod, ylab = "Abundance", lty = 2, pch = 19, cex = 1, logx = FALSE)
```

Arguments

mod	an drc object
ylab	ylab in plot function
lty	lty in plot function
pch	pch in plot function
cex	cex in plot function
logx	whether the x-axis should be in log scale

plotDCMat *Draw dose response curve given parameters in the omicsViewer object*

Description

Draw dose response curve given the feature Data/rowData, phenotype data/colData and expression matrix. The function is usually used in shinyApp.

Usage

```
plotDCMat(
  expr,
  pd,
  fd,
  featid,
  dose.var,
  curve.var = NULL,
  only.par = FALSE,
  ...
)
```

Arguments

expr	expression matrix
pd	phenotype data or colData
fd	feature data or rowData
featid	feature id to be visualized
dose.var	the column header indicating the dose/time/concentration
curve.var	the column header indicating the curve ids
only.par	logical value. If true, no plot generated, the function only returns the parameters of models.
...	other parameters passed to plot function, except col, pch, xlab, ylab

---

plotly_boxplot_module *Shiny module for boxplot using plotly - Module*

Description

Shiny module for boxplot using plotly - Module

Usage

```
plotly_boxplot_module(
  id,
  reactive_param_plotly_boxplot,
  reactive_checkpoint = reactive(TRUE)
)
```

Arguments

id	module id
reactive_param_plotly_boxplot	reactive value; argument passed to plotly_boxplot
reactive_checkpoint	reactive_value; check this value before render any plot/executing any calculation

Value

do not return any values

Examples

```
if (interactive()) {

  library(shiny)

  ui <- fluidPage(
    plotly_boxplot_ui("testplotly")
  )

  server <- function(input, output, session) {

    x <- cbind(matrix(rnorm(10000, mean = 3), 1000, 10), matrix(rnorm(20000), 1000, 20))
    x[sample(1:length(x), size = 0.3*length(x))] <- NA
    rownames(x) <- paste("R", 1:nrow(x), sep = "")
    colnames(x) <- paste("C", 1:ncol(x), sep = "")
    plotly_boxplot_module("testplotly",
                reactive_param_plotly_boxplot = reactive(list(
                  x = x# , i  = c(4, 20, 80)# , highlight = c(1, 4, 5, 20), extvar = 1:30
                ))
    )
  }

  shinyApp(ui, server)
}
```

---

plotly_boxplot_ui *Shiny module for boxplot using plotly - UI*

Description

Function should only be used for the developers

Usage

```
plotly_boxplot_ui(id)
```

Arguments

id	id

Value

a tagList of UI components

a tagList of UI components

Examples

```
if (interactive()) {

  library(shiny)

  ui <- fluidPage(
    plotly_boxplot_ui("testplotly")
  )

  server <- function(input, output, session) {

    x <- cbind(matrix(rnorm(10000, mean = 3), 1000, 10), matrix(rnorm(20000), 1000, 20))
    x[sample(1:length(x), size = 0.3*length(x))] <- NA
    rownames(x) <- paste("R", 1:nrow(x), sep = "")
    colnames(x) <- paste("C", 1:ncol(x), sep = "")
    plotly_boxplot_module("testplotly",
                reactive_param_plotly_boxplot = reactive(list(
                  x = x# , i  = c(4, 20, 80)# , highlight = c(1, 4, 5, 20), extvar = 1:30
                ))
    )
  }

  shinyApp(ui, server)
}
```

---

plotly_scatter_module *Shiny module for scatter plot using plotly - Module*

Description

Function should only be used for the developers

Usage

```
plotly_scatter_module(
  id,
  reactive_param_plotly_scatter,
  reactive_regLine = reactive(FALSE),
  reactive_checkpoint = reactive(TRUE),
  htest_var1 = reactive(NULL),
  htest_var2 = reactive(NULL)
)
```

Arguments

id	module id
reactive_param_plotly_scatter	reactive parammeters for plotly_scatter

reactive_regLine
: logical show or hide the regression line

reactive_checkpoint
: checkpoint

htest_var1
: when the plot is a beeswarmplot, two groups could be selected for two group comparison, this argument gives the default value. Mainly used for restoring the saved session.

htest_var2
: see above

Value

a list containing the information about the selected data points

an reactive object containing the information of selected, brushed points.

Examples

```
if (interactive()) {
  library(shiny)

  # two random variables
  x <- rnorm(30)
  y <- x + rnorm(30, sd = 0.5)

  # variables mapped to color, shape and size
  cc <- sample(letters[1:4], replace = TRUE, size = 30)
  shape <- sample(c("S1", "S2", "S3"), replace = TRUE, size = 30)
  sz <- sample(c(10, 20, 30, replace = TRUE, size = 30))

  ui <- fluidPage(
    plotly_scatter_ui("test_scatter")
  )

  server <- function(input, output, session) {
    v <- plotly_scatter_module("test_scatter",
                    # reactive_checkpoint = reactive(FALSE),
                    reactive_param_plotly_scatter = reactive(list(
                      x = x, y = y,
                      color = cc,
                      shape = shape,
                      size = sz,
                      tooltips = paste("A", 1:30)
                    )))
    observe(print(v()))
  }
  shinyApp(ui, server)



  # example beeswarm horizontal
  x <- rnorm(30)
  y <- sample(c("x", "y", "z"), size = 30, replace = TRUE)
```

```
  shinyApp(ui, server)

  # example beeswarm vertical
  x <- sample(c("x", "y", "z"), size = 30, replace = TRUE)
  y <- rnorm(30)
  shinyApp(ui, server)

  # return values
  x <- c(5, 6, 3, 4, 1, 2)
  y <- c(5, 6, 3, 4, 1, 2)
  ui <- fluidPage(
    plotly_scatter_ui("test_scatter")
  )
  server <- function(input, output, session) {
    v <- plotly_scatter_module("test_scatter",
                    reactive_param_plotly_scatter = reactive(list(
                      x = x, y = y, tooltips = paste("A", 1:6), highlight = 2:4
                    )))

    observe(print(v()))
  }
  shinyApp(ui, server)
}
```

`plotly_scatter_ui` *Shiny module for scatter plot using plotly - UI*

Description

Function should only be used for the developers

Usage

```
plotly_scatter_ui(id, height = "400px")
```

Arguments

`id`	id
`height`	figure height

Value

a tagList of UI components

Examples

```
if (interactive()) {
  library(shiny)

  # two random variables
  x <- rnorm(30)
  y <- x + rnorm(30, sd = 0.5)

  # variables mapped to color, shape and size
  cc <- sample(letters[1:4], replace = TRUE, size = 30)
  shape <- sample(c("S1", "S2", "S3"), replace = TRUE, size = 30)
  sz <- sample(c(10, 20, 30, replace = TRUE, size = 30))

  ui <- fluidPage(
    plotly_scatter_ui("test_scatter")
  )

  server <- function(input, output, session) {
    v <- callModule(plotly_scatter_module, id = "test_scatter",
                    # reactive_checkpoint = reactive(FALSE),
                    reactive_param_plotly_scatter = reactive(list(
                      x = x, y = y,
                      color = cc,
                      shape = shape,
                      size = sz,
                      tooltips = paste("A", 1:30)
                    )))
    observe(print(v()))
  }
  shinyApp(ui, server)



  # example beeswarm horizontal
  x <- rnorm(30)
  y <- sample(c("x", "y", "z"), size = 30, replace = TRUE)
  shinyApp(ui, server)

  # example beeswarm vertical
  x <- sample(c("x", "y", "z"), size = 30, replace = TRUE)
  y <- rnorm(30)
  shinyApp(ui, server)

  # return values
  x <- c(5, 6, 3, 4, 1, 2)
  y <- c(5, 6, 3, 4, 1, 2)
  ui <- fluidPage(
    plotly_scatter_ui("test_scatter")
  )
  server <- function(input, output, session) {
    v <- callModule(plotly_scatter_module, id = "test_scatter",
                    reactive_param_plotly_scatter = reactive(list(
```

```
                    x = x, y = y, tooltips = paste("A", 1:6), highlight = 2:4
                  )))

    observe(print(v()))
  }
  shinyApp(ui, server)
}
```

`prepOmicsViewer` *Prepare Omics Data for Visualization with omicsViewer*

Description

A comprehensive data preparation function that processes expression matrices and associated metadata for interactive visualization with `omicsViewer`. Automatically performs dimensionality reduction (PCA), statistical testing (t-tests), and integrates gene set annotations, STRING database IDs, and survival data.

Usage

```
prepOmicsViewer(
  expr,
  pData,
  fData,
  PCA = TRUE,
  ncomp = min(8, ncol(expr)),
  pca.fillNA = TRUE,
  t.test = NULL,
  ttest.fillNA = FALSE,
  ...,
  gs = NULL,
  stringDB = NULL,
  surv = NULL,
  SummarizedExperiment = TRUE
)
```

Arguments

`expr`	Numeric matrix. Expression data with features in rows and samples in columns. Should be log-transformed (e.g., log2 or log10). Row and column names must be unique. Missing values (NA) are permitted if `pca.fillNA` or `ttest.fillNA` are TRUE.
`pData`	Data.frame. Sample/phenotype metadata with one row per sample. Row names must match column names of `expr`. Should contain grouping variables for statistical tests.
`fData`	Data.frame. Feature metadata with one row per feature. Row names must match row names of `expr`. Can include gene symbols, descriptions, database IDs, etc.

`PCA` Logical. Whether to perform Principal Component Analysis. Default: TRUE. Results are added to both sample and feature metadata.

`ncomp` Integer. Number of principal components to compute. Default: minimum of 8 or the number of samples. Ignored if `PCA = FALSE`.

`pca.fillNA` Logical. If TRUE, missing values in `expr` are imputed before PCA by replacing with minimum value * 0.9. Default: TRUE. Two PCAs are performed: one with imputation and one without (if possible).

`t.test` Matrix or NULL. Definition of t-tests to perform. Should be an n×3 matrix where each row specifies: [column_name, group1, group2]. The column should exist in `pData`. Example: `rbind(c("Treatment", "Drug", "Control"), c("Genotype", "WT", "KO"))`. Results are added as columns to `fData`. NULL = no t-tests.

`ttest.fillNA` Logical. Whether to impute missing values before t-tests. Default: FALSE (features with NAs are excluded from testing).

`...` Additional arguments passed to `t.test`, such as `paired = TRUE` for paired t-tests or `var.equal = TRUE` for equal variance assumption.

`gs` Gene set annotations in one of two formats:

- Data.frame with columns: `featureId` (indices), `gsId` (gene set IDs), `weight` (optional weights). See `gsAnnotIdList`.
- Matrix or sparse matrix (dgCMatrix) with features in rows and gene sets in columns. Values indicate membership (0/1 or weights).

NULL = no gene set annotations. Enables ORA and GSEA analyses in viewer.

`stringDB` Character vector of length `nrow(expr)`. Protein/gene identifiers compatible with STRING database queries (e.g., Ensembl protein IDs, gene names). NULL = STRING network analysis disabled.

`surv` Survival data in one of three formats:

- Vector of length `ncol(expr)`: single survival time with censoring indicated by "+" suffix (e.g., "120+", "45").
- Matrix/data.frame: multiple survival endpoints with samples in rows. Column names will be prefixed with "Surv|all|". Values must be numeric with optional "+" suffix.

NULL = no survival analysis.

`SummarizedExperiment` Logical. If TRUE, returns a `SummarizedExperiment` object; if FALSE, returns an `ExpressionSet`. Default: TRUE.

Details

The function performs the following processing steps:

1. Validates dimensions and ensures unique row/column names
2. Standardizes column names by prefixing with data type (e.g., "General|All|")
3. Performs PCA on expression data (with and without imputation)
4. Conducts statistical tests (t-tests) between specified groups
5. Computes feature rankings across samples

6. Integrates gene set, STRING, and survival annotations
7. Sets sensible default axes for visualization

All metadata columns are prefixed with standardized headers following the pattern "Category|Subcategory|Variable" to organize variables in the viewer interface.

Value

A `SummarizedExperiment` or `ExpressionSet` object ready for visualization with `omicsViewer`.
The object includes:

- Expression matrix (and optionally imputed matrix)
- Enhanced metadata with PCA results, t-test statistics, rankings
- Gene set annotations (as attributes)
- Default axis selections (as attributes: "sx", "sy", "fx", "fy")

an object of `ExpressionSet` or `SummarizedExperiment` that can be visualized using `omicsViewer`

See Also

`omicsViewer` for launching the viewer. `multi.t.test` for details on t-test implementation. `gsAnnotIdList` for gene set annotation formatting.

Examples

```
packdir <- system.file("extdata", package = "omicsViewer")
# reading expression
expr <- read.delim(file.path(packdir, "expressionMatrix.tsv"), stringsAsFactors = FALSE)
colnames(expr) <- make.names(colnames(expr))
rownames(expr) <- make.names(rownames(expr))
# reading feature data
fd <- read.delim(file.path(packdir, "featureGeneral.tsv"), stringsAsFactors = FALSE)
# reading phenotype data
pd <- read.delim(file.path(packdir, "sampleGeneral.tsv"), stringsAsFactors = FALSE)

#  reading other datasets
drugData <- read.delim(file.path(packdir, "sampleDrug.tsv"))
# survival data
# this data is from cell line, the survival data are fake data to
# show how to use the survival data in #' omicsViewer
surv <- read.delim(file.path(packdir, "sampleSurv.tsv"))
# gene set information
genesets <- read_gmt(file.path(packdir, "geneset.gmt"), data.frame = TRUE)
gsannot <- gsAnnotIdList(idList = rownames(fd), gsIdMap = genesets, data.frame = TRUE)

# Define t-test to be done, a matrix nx3
# every row define a t-test, the format
# [column header] [group 1 in the test] [group 2 in the test]
tests <- rbind(
 c("Origin", "RE", "ME"),
 c("Origin", "RE", "LE"),
```

```
 c('TP53.Status', "MT", "WT")
 )
# prepare column for stringDB query
strid <- sapply(strsplit(fd$Protein.ID, ";|-"), "[", 1)
###
d <- prepOmicsViewer(
  expr = expr, pData = pd, fData = fd,
  PCA = TRUE, pca.fillNA = TRUE,
  t.test = tests, ttest.fillNA = FALSE,
  gs = gsannot, stringDB = strid, surv = surv)
# feature space - default x axis
attr(d, "fx") <- "ttest|RE_vs_ME|mean.diff"
# feature space - default y axis
attr(d, "fy") <- "ttest|RE_vs_ME|log.fdr"
# sample space - default x axis
attr(d, "sx") <- "PCA|All|PC1("
# sample space - default y axis
attr(d, "sy") <- "PCA|All|PC2("
# Save object and view
# saveRDS(d, file = "dtest.RDS")
##  to open the viewer
# omicsViewer("./")
```

`read.proteinGroups` *Reading proteinGroup table of MaxQuant output*

Description

A convenience function to read the proteinGroups table of MaxQuant output. The function organize the result into different tables, e.g. iBAQ.

Usage

```
read.proteinGroups(x, quant = c("LF", "TMT")[1])
```

Arguments

x	the proteinGroup.txt file returned by MaxQuant search
quant	the quantification method, LF or TMT

Value

a list of tables extracted from proteinGroups.txt file

read.proteinGroups.lf *Read protein groups output of maxquant output and split it to columns*

Description

Read protein groups output of maxquant output and split it to columns

Usage

```
read.proteinGroups.lf(file)
```

Arguments

file	Maxquant proteinGroup.txt file path

Value

a list of tables extracted from proteinGroups.txt file

read_gmt *Reading gene set .gmt file*

Description

Frequently the .gmt files are downloaed from MSigDB database

Usage

```
read_gmt(x, id = NA, data.frame = FALSE)
```

Arguments

x	the name/path of the gmt file to be read
id	the id used in gene sets, if is not NA, it should be either "SYMBOL" or "ENTREZ". Usually only used when reading the .gmt file downloaded from MSigDB.
data.frame	logical; whether to organize the data in data.frame format. Default is FALSE, a list will be returned.

Value

a list or data frame of gene set. When data.frame = TRUE, the returned object is a data.frame with two columns: id and term.

Examples

```
file <- system.file("extdata", package = "omicsViewer")
file <- file.path(file, "geneset.gmt")
gs <- read_gmt(file)
```

readESVObj — *Read the object of SummarizedExperiment or ExpressetSet to be visualized using omicsViewer*

Description

This function accept a path to a sqlite database or RDS object. If an RDS file to be read, The function is similar to `readRDS`. It reads the object to R working environment and perform extra two things. 1. If the loaded data an class of `SummarizedExperiment`, it will be converted to `ExpressionSet`; 2. If the gene set annotatio is in matrix format, the gene set annotation is converted to `data.frame` format.

Usage

```
readESVObj(x)
```

Arguments

x	the path of an object of `SummarizedExperiment` or `ExpressionSet`, passed to readRDS

Value

an object of class `ExpressionSet` or `SummarizedExperiment` to be visualzied.

Examples

```
file <- system.file("extdata/demo.RDS", package = "omicsViewer")
obj <- readESVObj(file)
```

removeVarQC — *Removing variance of reference samples*

Description

This normalization removes the variance in reference samples. The method do not need to specific the batch assignment but cannot work with data contains less than five common reference samples. A typical use of this normalization is to correct some drifting effect in mass spec based label free proteomics or untargeted metabolomics experiment. Usually, this is a very strong normalization should only be used with good reasons.

Usage

```
removeVarQC(x, ref, positive = TRUE, ...)
```

Arguments

x	an expression matrix
ref	the index of reference samples
positive	logical; force only positive values in the resulted matrix
...	if given, normalize.nQuantiles will be called first, the arguments here will be passed to normalize.nQuantiles

Value

a normalized matrix

Examples

```
e1 <- matrix(rnorm(5000), 100, 50)+10
e2 <- removeVarQC(x = e1, ref = seq(5, 45, by = 10))
boxplot(e2)
```

---

rowshift *Row-wise normalization of expression matrix with or without reference sample*

Description

Row-wise normalization of expression matrix with or without reference sample

Usage

```
rowshift(x, batch, ref = NULL, useMean = FALSE)
```

Arguments

x	an expression matrix where rows are features, e.g. genes, proteins and columns are samples. The values in the matrix are usually log transformed.
batch	a factor or vector has the same length as ncol(x) to indicate the batch assignment of samples.
ref	a logical vector has the same length as ncol(x) to indicated which columns are the common references among batches. If it is NULL (by default), the mean of all channels will be used as batch reference. When NA present in the reference channels, the mean values will be used in correction.
useMean	logical; whether to use means of batches, usually set to TRUE when no reference available

Value

a matrix (hopefully without/with less batch effect)

Examples

```
e1 <- matrix(rnorm(5000), 500, 10)
e1[, 6:10] <- e1[, 6:10] + 3
boxplot(e1)
f <- rep(c("a", "b"), each = 5)
e2 <- rowshift(x = e1, batch = f)
boxplot(e2)
```

---

saveOmicsViewerDb *Save the xcmsViewer result object as sqlite database*

Description

Save the xcmsViewer result object as sqlite database

Usage

```
saveOmicsViewerDb(obj, db.file, overwrite = TRUE)

## S4 method for signature 'SummarizedExperiment,character'
saveOmicsViewerDb(obj, db.file, overwrite = TRUE)

## S4 method for signature 'ExpressionSet,character'
saveOmicsViewerDb(obj, db.file, overwrite = TRUE)
```

Arguments

obj	an object of class ExpressionSet or SummarizedExperiment
db.file	a character indicate file name of the database file
overwrite	logical. whether the database should be overwritten if exist already.

Value

the directory where the database saved

Examples

```
f <- system.file("extdata", "demo.RDS", package = "omicsViewer")
es <- readRDS(f)
# The following line will write a database file on your disk
# saveOmicsViewerDb(es, db.file = "./omicsViewerData.db")
```

stringD3Net *Drawing network given network and gene set result*

Description

Drawing network given network and gene set result

Usage

```
stringD3Net(ntwk, gsa, i, label = FALSE)
```

Arguments

ntwk	network result, often returned by "stringNetwork" function.
gsa	gene set result, often returned by "stringGSA" function
i	row index of gsa, which should be highlighted in the network
label	whether the point should be labelled in the network

stringGSA *Performing gene set analysis using string-db*

Description

Performing gene set analysis using string-db

Usage

```
stringGSA(
  genes,
  taxid = 9606,
  background = NULL,
  backgroundStringId = FALSE,
  caller = "omicsViewer"
)
```

Arguments

genes	a character vector of gene ids
taxid	taxonomy ids (species NCBI identifier)
background	the background genes
backgroundStringId	logical value, whether the identifier given in background is already stringid, only used when background != NULL. You can use getStringId to convet your id to string id.
caller	your identifier for string-db.org

Note

https://string-db.org/cgi/help.pl?subpage=api

Examples

```
1
# gg = c('P04637', 'P00533', 'P04626', "Q8IYB3", "O75494", "Q9Y696")
# u <- stringGSA(gg)
```

---

stringNetwork *Retrieve string network*

Description

get string network

Usage

```
stringNetwork(genes, taxid = 9606, caller = "omicsViewer")
```

Arguments

genes	the gene ids
taxid	taxonomy ids
caller	your identifier for string-db.org

---

triselector_module *The three-step selector - the module function*

Description

The selector is used to select columns of phenotype and feature data. Function should only be used for the developers.

Usage

```
triselector_module(
  id,
  reactive_x,
  reactive_selector1 = reactive(NULL),
  reactive_selector2 = reactive(NULL),
  reactive_selector3 = reactive(NULL),
  label = "Group Label:"
)
```

Arguments

id	module id
reactive_x	an nx3 matrix
reactive_selector1	default value for selector 1
reactive_selector2	default value for selector 2
reactive_selector3	default value for selector 3
label	of the triselector

Value

an reactive object containing the selected values

Examples

```
if (interactive()) {
  library(shiny)
  library(Biobase)

  file <- system.file("extdata/demo.RDS", package = "omicsViewer")
  dat <- readRDS(file)
  fData <- fData(dat)
  triset <- stringr::str_split_fixed(colnames(fData), '\\|', n= 3)

  ui <- fluidPage(
    triselector_ui("tres"),
    triselector_ui("tres2")
  )
  server <- function(input, output, session) {
    v1 <- triselector_module("tres", reactive_x = reactive(triset),
                    reactive_selector1 = reactive("ttest"),
                    reactive_selector2 = reactive("RE_vs_ME"),
                    reactive_selector3 = reactive("mean.diff")
    )
    v2 <- triselector_module("tres2", reactive_x = reactive(triset),
                    reactive_selector1 = reactive("ttest"),
                    reactive_selector2 = reactive("RE_vs_ME"),
                    reactive_selector3 = reactive("log.fdr"))
    observe({
      print("//////////////////////////")
      print(v1())
    })
  }

  shinyApp(ui, server)
}
```

`triselector_ui` *The three-step selector - the ui function*

Description

Function should only be used for the developers

Usage

```
triselector_ui(id, right_margin = "20")
```

Arguments

`id`	id
`right_margin`	margin on the right side, in px. For example, "20" translates to "20px".

Value

a tagList of UI components

Examples

```
if (interactive()) {
  library(shiny)
  library(Biobase)

  file <- system.file("extdata/demo.RDS", package = "omicsViewer")
  dat <- readRDS(file)
  fData <- fData(dat)
  triset <- stringr::str_split_fixed(colnames(fData), '\\|', n= 3)

  ui <- fluidPage(
    triselector_ui("tres"),
    triselector_ui("tres2")
  )
  server <- function(input, output, session) {
    v1 <- triselector_module("tres", reactive_x = reactive(triset),
                    reactive_selector1 = reactive("ttest"),
                    reactive_selector2 = reactive("RE_vs_ME"),
                    reactive_selector3 = reactive("mean.diff")
    )
    v2 <- triselector_module("tres2", reactive_x = reactive(triset),
                    reactive_selector1 = reactive("ttest"),
                    reactive_selector2 = reactive("RE_vs_ME"),
                    reactive_selector3 = reactive("log.fdr"))
    observe({
      print("//////////////////////////")
      print(v1())
    })
```

```
  }

  shinyApp(ui, server)
}
```

`trisetter` *Create a nx3 matrix that can be use for triselector given a meta and expression table*

Description

only used inside reactive

Usage

```
trisetter(meta, expr = NULL, combine)
```

Arguments

`meta`	a meta data, usually either phenotype data or feature data
`expr`	expression matrix, optional.
`combine`	how the meta and expression to be combined. Should be either "pheno" or "feature' or "none".

Value

a nx3 matrix

a `data.frame` with 3 columns

`validate_character_vector` *Validate character vector input*

Description

Validate character vector input

Usage

```
validate_character_vector(
  vec,
  name = "vector",
  required = TRUE,
  min_length = 1,
  max_length = Inf,
  allow_empty = FALSE
)
```

Arguments

vec	Reactive expression or vector to validate
name	Character. Name of the vector for error messages
required	Logical. Whether NULL values are allowed. Default: TRUE
min_length	Integer. Minimum length required. Default: 1
max_length	Integer. Maximum length allowed. Default: Inf
allow_empty	Logical. Whether empty strings are allowed. Default: FALSE

Value

List with $valid (logical) and $message (character) fields

---

validate_matching_dimensions
Validate matching dimensions

Description

Validate matching dimensions

Usage

```
validate_matching_dimensions(
  obj1,
  obj2,
  dimension = c("rows", "cols"),
  name1 = "object1",
  name2 = "object2"
)
```

Arguments

obj1	First object (matrix or data.frame)
obj2	Second object (matrix or data.frame)
dimension	Character. Either "rows" or "cols" to check
name1	Character. Name of first object for error messages
name2	Character. Name of second object for error messages

Value

List with $valid (logical) and $message (character) fields

validate_numeric_matrix *Validate numeric matrix input*

Description

Validate numeric matrix input

Usage

```
validate_numeric_matrix(
  mat,
  name = "matrix",
  required = TRUE,
  min_rows = 1,
  min_cols = 1,
  allow_na = TRUE
)
```

Arguments

mat	Reactive expression or matrix to validate
name	Character. Name of the matrix for error messages
required	Logical. Whether NULL values are allowed. Default: TRUE
min_rows	Integer. Minimum number of rows required. Default: 1
min_cols	Integer. Minimum number of columns required. Default: 1
allow_na	Logical. Whether NA values are allowed. Default: TRUE

Value

List with $valid (logical) and $message (character) fields

validate_numeric_range *Validate numeric range*

Description

Validate numeric range

Usage

```
validate_numeric_range(
  value,
  name = "value",
  min = -Inf,
  max = Inf,
  allow_na = FALSE
)
```

Arguments

value	Numeric value to validate
name	Character. Name for error messages
min	Numeric. Minimum allowed value (inclusive). Default: -Inf
max	Numeric. Maximum allowed value (inclusive). Default: Inf
allow_na	Logical. Whether NA is allowed. Default: FALSE

Value

List with $valid (logical) and $message (character) fields

---

validate_triselector_input

Validate triselector input matrix

Description

Validate triselector input matrix

Usage

```
validate_triselector_input(
  triset,
  name = "triselector data",
  allow_null = FALSE
)
```

Arguments

triset	Matrix or reactive expression. Should be nx3 matrix from str_split_fixed
name	Character. Name for error messages
allow_null	Logical. Whether NULL is allowed (e.g., during app startup). Default: FALSE

Value

List with $valid (logical) and $message (character) fields

validMQFolder *MQ folder validator Validate whether a folder is a MQ output folder*

Description

MQ folder validator Validate whether a folder is a MQ output folder

Usage

```
validMQFolder(dir)
```

Arguments

dir	the directory to check

Details

from the root level, these files exist: mqpar.xml [[combined/]txt/]proteinGroups.txt

Value

a list containing the info about MQ folder check

varSelector *variable selector*

Description

variable selector

Usage

```
varSelector(x, expr, meta, alternative = NULL)
```

Arguments

x	variable return by triselector, a list of length three named as "analysis", "subset" and "variable'
expr	the expression matrix
meta	a meta matrix
alternative	alternative value to be returned when nothing to select

Value

the selected values in input argument x

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