

# An introduction to the nuCpos package

Hiroaki Kato,\* Takeshi Urano

July 4, 2024

## 1 About nuCpos

*nuCpos*, a derivative of *NuPoP*, is an R package for predicting **n**ucleosome **p**ositions. *nuCpos* calculates local and whole nucleosomal HBA scores for a given 147-bp sequence. This package was designed to demonstrate the use of chemical maps in prediction. As the parental package *NuPoP* now provides chemical-map-based prediction, the function for dHMM-based prediction was removed from this package. *nuCpos* continues to provide functions for HBA calculation. The models are based on chemical maps of nucleosomes from budding yeast *Saccharomyces cerevisiae* (Brogaard et al. (2012)), fission yeast *Schizosaccharomyces pombe* (Moyle-Heyrman et al. (2012)), or embryonic stem cells of house mouse *Mus musculus* (Voong et al. (2016)).

The parental package *NuPoP*, licensed under GPL-2, was developed by Ji-Ping Wang and Liqun Xi. Please refer to Xi et al. (2010) and Wang et al. (2008) for technical details of *NuPoP*. Their excellent codes were adapted in *nuCpos* to demonstrate the usefulness of chemical maps in prediction.

Note that when *nuCpos* was released, *NuPoP* only used an MNase-seq-based map of budding yeast nucleosomes to train a duration hidden Markov model. However, as *NuPoP* now provides chemical map-based prediction, users are encouraged to use *NuPoP* functions to conduct dHMM-based prediction in their original way.

## 2 nuCpos functions

*nuCpos* has two functions: `HBA`, and `localHBA`.

The functions `HBA` and `localHBA` receive a sequence of 147-bp DNA and calculate whole nucleosomal and local HBA scores. These functions invoke core Fortran codes for HBA calculation that were adapted from the excellent dHMM code of *NuPoP*.

*nuCpos* requires the *Biostrings* package, especially when DNA sequences are given as `DNAStrng` objects to the functions `HBA`, and `localHBA`. These functions can also receive DNA sequences as simple character string objects without loading the *Biostrings* package. Note: *nuCpos* requires the *NuPoP* package to perform some example runs.

Load the *nuCpos* package as follows:

```
> library(nuCpos)
```

---

\*hkato@med.shimane-u.ac.jp

### 3 Histone binding affinity score calculation with HBA

HBA score can be calculated for a given 147-bp sequence with the `HBA` function. In the examples below, a character string object `inseq` and a `DNAStrng` object `INSEQ` with the same 147-bp DNA sequences are given to `HBA`. Note: the *Biostrings* package is required for the latter case.

```
> load(system.file("extdata", "inseq.RData", package = "nuCpos"))
> HBA(inseq = inseq, species = "sc")
```

```
HBA
-2.460025
```

```
> for(i in 1:3) cat(substr(inseq, start = (i-1)*60+1,
+   stop = (i-1)*60+60), "\n")
```

```
ATCGAGAATCCCGGTGCCGAGGCCGCTCAATTGGTCGTAGACAGCTCTAGCACCGCTTAA
ACGCACGTACGCGCTGTCCCCGCGTTTTAACCGCCAAGGGGATTACTCCCTAGTCTCCA
GGCACGTGTCAGATATATACATCCGAT
```

```
> load(system.file("extdata", "INSEQ_DNAStrng.RData",
+   package = "nuCpos"))
> INSEQ
```

147-letter `DNAStrng` object

```
seq: ATCGAGAATCCCGGTGCCGAGGCCGCTCAATTGGTC...TAGTCTCCAGGCACGTGTCAGATATATACATCCGAT
```

```
> HBA(inseq = INSEQ, species = "sc")
```

```
HBA
-2.460025
```

The argument `inseq` is the character string object to be given. Alternatively, a `DNAStrng` object can be used here. The length of DNA must be 147 bp. The argument `species` can be specified as follows: `mm` = *M. musculus*; `sc` = *S. cerevisiae*; `sp` = *S. pombe*.

### 4 Local histone binding affinity score calculation with localHBA

Local HBA scores are defined as HBA scores for 13 overlapping subnucleosomal segments named A to M. They can be calculated for a given 147-bp sequence with the `localHBA` function. Like `HBA`, this function can receive either a character string object or a `DNAStrng` object. The segment G corresponds to the central 21 bp region, in which the dyad axis passes through the 11th base position. This means that the local HBA score for the G segment implies the relationship between DNA and histone proteins at around superhelical locations -0.5 and +0.5. The neighboring F segment, which is 20 bp in length, is for SHLs -1.5 and -0.5. The result of example run shown below suggests that subsequence of `inseq` around SHL -3.5 and -2.5 is suitable for nucleosome formation.

```
> localHBA(inseq = inseq, species = "sc")
```

```

      1HBA_A      1HBA_B      1HBA_C      1HBA_D      1HBA_E      1HBA_F
-1.56140949 -1.62502354  0.48885990  2.37615568  2.90458625 -1.35195919
      1HBA_G      1HBA_H      1HBA_I      1HBA_J      1HBA_K      1HBA_L
-3.13228907 -0.32208031  0.27650871  0.01922002  0.49787625 -0.17151500
      1HBA_M
-1.27186158

```

```

> barplot(localHBA(inseq = inseq, species = "sc"),
+   names.arg = LETTERS[1:13], xlab = "Nucleosomal subsegments",
+   ylab = "local HBA", main = "Local HBA scores for inseq")

```

## 5 Acknowledgements

We would like to thank Drs. Shimizu, Fuse and Ichikawa for sharing DNA sequences and *in vivo* data, and giving fruitful comments. We would like to thank Dr. Ji-Ping Wang and his colleagues for distributing NuPoP under the GPL-2 license. In this package, their excellent code for dHMM-based prediction was adapted for chemical map-based prediction to demonstrate the usefulness of chemical maps in prediction. As we noticed that canceling of HBA smoothing helps predicting rotational settings, predNuCpos in the earlier version provided this option. However, for those who want to predict nucleosome occupancy in the original way with chemical maps, we encourage users to use NuPoP functions as it now provides chemical map-based predictions. In our functions HBA and localHBA, their excellent code was also adapted to calculate the scores of given 147-bp sequences independently of the genomic context. The function HBA now runs without invoking a fortran subroutine.

## References

- Wang JP, Fondufe-Mittendorf Y, Xi L, Tsai GF, Segal E and Widom J (2008). Preferentially quantized linker DNA lengths in *Saccharomyces cerevisiae*. *PLoS Computational Biology*, 4(9):e1000175.
- Xi L, Fondufe-Mittendorf Y, Xia L, Flatow J, Widom J and Wang JP (2010). Predicting nucleosome positioning using a duration hidden markov model. *BMC Bioinformatics*, 11:346.
- Brogaard K, Xi L, and Widom J (2012). A map of nucleosome positions in yeast at base-pair resolution. *Nature*, 486(7404):496-501.
- Moyle-Heyrman G, Zaichuk T, Xi L, Zhang Q, Uhlenbeck OC, Holmgren R, Widom J and Wang JP (2013). Chemical map of *Schizosaccharomyces pombe* reveals species-specific features in nucleosome positioning. *Proc. Natl. Acad. Sci. U. S. A.*, 110(50):20158-63.
- Ichikawa Y, Morohoshi K, Nishimura Y, Kurumizaka H and Shimizu M (2014). Telomeric repeats act as nucleosome-disfavouring sequences in vivo. *Nucleic Acids Res.*, 42(3):1541-1552.
- Voong LN, Xi L, Sebeson AC, Xiong B, Wang JP and Wang X (2016). Insights into Nucleosome Organization in Mouse Embryonic Stem Cells through Chemical Mapping. *Cell*, 167(6):1555-1570.
- Fuse T, Katsumata K, Morohoshi K, Mukai Y, Ichikawa Y, Kurumizaka H, Yanagida A, Urano T, Kato H, and Shimizu M (2017). Parallel mapping with site-directed hydroxyl radicals and micrococcal nuclease reveals structural features of positioned nucleosomes in vivo. *Plos One*, 12(10):e0186974.