

# Introduction to RBM package

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August 30, 2024

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## 1 Overview

This document provides an introduction to the RBM package. The RBM package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the RBM package computes the moderated t-statistics based on the observed data set for each feature using the lmFit and eBayes function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

## 2 Getting started

The `RBM` package can be installed and loaded through the following R code.  
Install the `RBM` package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the `RBM` package with:

```
> library(RBM)
```

## 3 RBM\_T and RBM\_F functions

There are two functions in the `RBM` package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The *p*-values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1), 1000, 6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata, mydesign, 100, 0.05)
> summary(myresult)

      Length Class  Mode
ordfit_t     1000 -none- numeric
ordfit_pvalue 1000 -none- numeric
ordfit_beta0  1000 -none- numeric
ordfit_beta1  1000 -none- numeric
permutation_p 1000 -none- numeric
bootstrap_p    1000 -none- numeric

> sum(myresult$permutation_p<=0.05)
```

```

[1] 25

> which(myresult$permutation_p<=0.05)

[1] 57 97 116 128 162 182 238 272 346 358 445 560 624 640 653 663 739 768 775
[20] 822 827 889 894 900 994

> sum(myresult$bootstrap_p<=0.05)

[1] 10

> which(myresult$bootstrap_p<=0.05)

[1] 36 97 162 339 346 444 477 663 754 943

> permutation_adjp <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adjp<=0.05)

[1] 4

> bootstrap_adjp <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adjp<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7, 0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutation_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 57

> which(myresult2$bootstrap_p<=0.05)

[1] 34 41 57 74 80 106 107 119 131 156 162 170 188 199 207 335 336 347 354
[20] 357 396 460 492 516 531 562 563 568 582 588 601 605 665 667 672 700 708 730
[39] 742 745 779 788 792 806 807 830 834 848 860 884 916 917 930 936 947 973 976

> bootstrap2_adjp <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adjp<=0.05)

[1] 1

```

- Examples using the RBM\_F function: normdata\_F simulates a standardized gene expression data and unifdata\_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

      Length Class  Mode
ordfit_t     3000 -none- numeric
ordfit_pvalue 3000 -none- numeric
ordfit_beta1  3000 -none- numeric
permutation_p 3000 -none- numeric
bootstrap_p   3000 -none- numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)
[1] 49

> sum(myresult_F$permutation_p[, 2]<=0.05)
[1] 50

> sum(myresult_F$permutation_p[, 3]<=0.05)
[1] 58

> which(myresult_F$permutation_p[, 1]<=0.05)
[1]    9   15   22   24   40   63   65   86   90  116  162  179  181  185  204  219  297  336  343
[20] 380  388  456  487  521  527  539  544  549  575  584  585  596  600  605  613  640  655  699
[39] 701  704  713  730  845  866  869  910  941  943  957

> which(myresult_F$permutation_p[, 2]<=0.05)
[1]    1    9   15   22   24   40   63   66   86   90  116  162  179  181  182  185  219  297  335
[20] 343  351  380  388  407  422  482  487  521  527  539  584  585  596  600  605  613  640  655
[39] 699  701  713  730  807  845  866  869  910  943  957  972

> which(myresult_F$permutation_p[, 3]<=0.05)
[1]    1     5    9   15   22   24   40   63   65   66   69   86   90  116  162  179  181  185  219
[20] 278  297  305  323  336  343  380  388  407  428  512  521  527  539  544  549  575  584  585
[39] 596  600  605  640  655  701  704  713  722  740  807  819  845  869  881  910  941  943  957
[58] 972

```

```

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

[1] 8

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 8

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 10

> which(con2_adjp<=0.05/3)

[1] 63 179 527 600 605 701 845 910

> which(con3_adjp<=0.05/3)

[1] 24 63 86 90 179 343 521 605 701 910

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

      Length Class  Mode
ordfit_t     3000 -none- numeric
ordfit_pvalue 3000 -none- numeric
ordfit_beta1  3000 -none- numeric
permutation_p 3000 -none- numeric
bootstrap_p   3000 -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 52

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 61

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 47

```

```

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

[1]   6   9  20  46  64  70 115 123 135 136 150 157 181 237 240 269 298 301 325
[20] 330 380 382 384 447 461 464 469 550 563 564 590 592 602 630 633 636 677 694
[39] 722 730 740 742 743 779 812 845 885 901 915 936 947 951

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1]   6   9  44  46  64  65  70  96 112 123 135 140 150 157 171 178 181 188 220
[20] 237 240 269 301 323 330 367 368 380 384 447 461 464 469 514 550 553 563 590
[39] 592 594 624 630 636 677 722 730 740 742 746 749 751 779 812 817 845 856 885
[58] 901 915 936 951

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1]   6   9  46  58  64  96 119 123 135 150 181 188 240 301 325 330 362 367 368
[20] 380 384 447 461 464 469 514 529 550 563 602 630 633 636 677 710 722 730 740
[39] 742 749 779 812 817 885 901 915 951

> con21_adjp <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adjp<=0.05/3)

[1] 3

> con22_adjp <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adjp<=0.05/3)

[1] 6

> con23_adjp <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adjp<=0.05/3)

[1] 4

```

## 4 Ovarian cancer methylation example using the RBM\_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of `RBM_T` in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the `RBM_T` function and presenting the results for further validation and investigations.

```

> system.file("data", package = "RBM")
[1] "/tmp/RtmpsmqDNI/Rinst3582f1f84a4/RBM/data"

> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)

    IlmnID      Beta      exmdata2[, 2]      exmdata3[, 2]
cg00000292: 1   Min.   :0.01058   Min.   :0.01187   Min.   :0.009103
cg00002426: 1   1st Qu.:0.04111   1st Qu.:0.04407   1st Qu.:0.041543
cg00003994: 1   Median  :0.08284   Median  :0.09531   Median  :0.087042
cg00005847: 1   Mean    :0.27397   Mean    :0.28872   Mean    :0.283729
cg00006414: 1   3rd Qu.:0.52135   3rd Qu.:0.59032   3rd Qu.:0.558575
cg00007981: 1   Max.    :0.97069   Max.    :0.96937   Max.    :0.970155
(Other)     :994          NA's    :4
exmdata4[, 2]  exmdata5[, 2]  exmdata6[, 2]  exmdata7[, 2]
Min.   :0.01019   Min.   :0.01108   Min.   :0.01937   Min.   :0.01278
1st Qu.:0.04092   1st Qu.:0.04059   1st Qu.:0.05060   1st Qu.:0.04260
Median  :0.09042   Median  :0.08527   Median  :0.09502   Median  :0.09362
Mean    :0.28508   Mean    :0.28482   Mean    :0.27348   Mean    :0.27563
3rd Qu.:0.57502   3rd Qu.:0.57300   3rd Qu.:0.52099   3rd Qu.:0.52240
Max.    :0.96658   Max.    :0.97516   Max.    :0.96681   Max.    :0.95974
NA's    :1

exmdata8[, 2]
Min.   :0.01357
1st Qu.:0.04387
Median  :0.09282
Mean    :0.28679
3rd Qu.:0.57217
Max.    :0.96268

> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)

      Length Class  Mode
ordfit_t     1000  -none- numeric
ordfit_pvalue 1000  -none- numeric
ordfit_beta0  1000  -none- numeric
ordfit_beta1  1000  -none- numeric
permutation_p 1000  -none- numeric
bootstrap_p   1000  -none- numeric

> sum(diff_results$ordfit_pvalue<=0.05)
[1] 47

```

```

> sum(diff_results$permutation_p<=0.05)
[1] 43

> sum(diff_results$bootstrap_p<=0.05)
[1] 43

> ordfit_adjp <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adjp<=0.05)

[1] 0

> perm_adjp <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adjp<=0.05)

[1] 2

> boot_adjp <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adjp<=0.05)

[1] 4

> diff_list_perm <- which(perm_adjp<=0.05)
> diff_list_boot <- which(boot_adjp<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[, diff_results$ordfit_t[diff_list_perm]], diff_results$permutation_p[diff_list_perm])
> print(sig_results_perm)

   IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
83  cg00072216 0.04505377    0.04598964    0.04000674    0.03231534
450  cg00432979 0.03681359    0.04515700    0.04374394    0.03683598
     exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
83      0.04965089    0.04833366    0.03466159    0.04390894
450      0.04419125    0.04409653    0.02839263    0.03410020
  diff_results$ordfit_t[diff_list_perm]
83                      1.947226
450                      1.274731
  diff_results$permutation_p[diff_list_perm]
83                      0
450                      0

> sig_results_boot <- cbind(ovarian_cancer_methylation[, diff_results$ordfit_t[diff_list_boot]], diff_results$permutation_p[diff_list_boot])
> print(sig_results_boot)

   IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
95  cg00081975 0.03633894    0.04975194    0.06024723    0.05598723
146  cg00134539 0.61101320    0.53321780    0.45999340    0.46787420
280  cg00260778 0.64319890    0.60488960    0.56735060    0.53150910

```

```
979 cg00945507 0.13432250      0.23854600      0.34749760      0.28903340
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
95      0.04561792      0.05115624      0.06068253      0.06168212
146     0.67191510      0.63137380      0.47929610      0.45428300
280     0.61920530      0.61925200      0.46753250      0.55632410
979     0.11848510      0.16653850      0.30718420      0.26624740
      diff_results$ordfit_t[diff_list_boot]
95                  -2.654324
146                  5.636263
280                  4.337628
979                 -4.968792
      diff_results$bootstrap_p[diff_list_boot]
95                      0
146                      0
280                      0
979                      0
```