

Package: VisiumIO (via r-universe)

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Title Import Visium data from the 10X Space Ranger pipeline

Version 1.9.5

Description The package allows users to readily import spatial data obtained from either the 10X website or from the Space Ranger pipeline. Supported formats include tar.gz, h5, and mtx files. Multiple files can be imported at once with *List type of functions. The package represents data mainly as SpatialExperiment objects.

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compareBarcodes	<i>Compare barcodes between raw and filtered data</i>
-----------------	---

Description

This function compares the barcodes between raw and filtered data **depending** on the order of processing. Typically, the "raw" barcodes are compared to the "filtered" ones. The presence of raw barcodes in the filtered data are marked as TRUE in the resulting `data.frame`.

Usage

```
compareBarcodes(  
  from_resource,  
  to_resource,  
  spacerangerOut,  
  format = c("mtx", "h5"),  
  processing = c("raw", "filtered"),  
  ...  
)
```

Arguments

- `from_resource` character(1) The path to the resource file whose barcodes are used as the basis of the comparison; typically, the "raw" feature barcodes are used.
- `to_resource` character(1) The path to the resource file whose barcodes are compared to the `from_resource`; typically, the "filtered" feature barcodes.

spacerangerOut	character(1) A single string specifying the path to the directory where the output of spaceranger count is located; typically (but not necessarily), this is the outs directory. The directory must contain the (processing)_feature_bc_matrix and spatial sub directories.
format	The format of the output. If missing and con is a file name, the format is derived from the file extension. This argument is unnecessary when con is a derivative of BiocFile .
processing	character(2) A vector of length 2 that corresponds to the processing type. The processing types are typically "raw" and "filtered". These are the prefixes of the folder names raw_feature_bc_matrix and filtered_feature_bc_matrix. The order of the vector determines the comparison. By default, processing = c("raw", "filtered"), which means barcodes in the raw data are compared to the filtered data.
...	Additional arguments passed to TENxH5 or TENxFileList.

Value

A data.frame with barcodes of the first element in the processing data type as the first column and a logical vector indicating whether the barcodes are found in the second element in processing. For example, if processing is c("raw", "filtered"), then the first column will be the barcodes in the raw data and the second column will be a logical vector indicating whether the barcodes are found in the filtered data.

Examples

```
if (interactive()) {
  compareBarcodes(
    from_resource = "V1_Adult_Mouse_Brain_raw_feature_bc_matrix.tar.gz",
    to_resource =
      "V1_Adult_Mouse_Brain_filtered_feature_bc_matrix.tar.gz",
  )

  compareBarcodes(
    from_resource =
      "V1_Adult_Mouse_Brain_raw_feature_bc_matrix.h5",
    to_resource =
      "V1_Adult_Mouse_Brain_filtered_feature_bc_matrix.h5"
  )

  compareBarcodes(spacerangerOut = "~/data/outs", format = "h5")

  compareBarcodes(
    spacerangerOut = "~/data/feature_bc_matrix", format = "mtx"
  )

  compareBarcodes(
    spacerangerOut = "~/data/folder_feature_bc_matrix", format = "mtx"
  )
}
```

st_invert_y	<i>Flip the Y-axis of cell or nucleus segmentations to align with H&E image</i>
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Description

This function flips the Y-axis of cell or nucleus segmentations stored in an sf object to align with the H&E image. The Y-axis flipping is necessary because the origin (0,0) in image coordinates is at the top-left corner, while in Cartesian coordinates, the origin is at the bottom-left corner. The function takes into account the image height and scaling factor to accurately flip the Y-coordinates of the segmentations.

Usage

```
st_invert_y(sf, type = c("POINT", "POLYGON"), img_height, scalef)
```

Arguments

sf	sf an sf class object read from a .geojson file.
type	character(1) "POINT" for cell centroid, or "POLYGON" for cell segmentation mask. Default is "POINT".
img_height	numeric(1) The total length along the Y axis of the image. Obtained by reading in hires or lowres .png under /spatial folder with magick::image_read().
scalef	numeric(1) The scaling factor from a /spatial/scalefactors_json.json file

Value

an sf object with Y-axis of the points or polygons flipped

Author(s)

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Examples

```
geojson_file <- system.file(
  file.path("extdata", "segmented_outputs", "cell_segmentations.geojson"),
  mustWork = TRUE, package = "VisiumIO"
)
geo_data <- sf::st_read(geojson_file, quiet = TRUE)
st_invert_y(
  sf = geo_data, type = "POLYGON", img_height = 3886, scalef = 0.079
)
```

TENxGeoJSON-class *Import 10X Genomics GeoJSON files*

Description

TENxGeoJSON is a class to represent and import GeoJSON files from 10X Genomics. It is a composed class of [TENxIO::TENxFile](#).

Usage

```
TENxGeoJSON(resource)

## S4 method for signature 'TENxGeoJSON,ANY,ANY'
import(con, format, text, ...)
```

Arguments

resource	character(1) The path to the file
con	The connection from which data is loaded or to which data is saved. If this is a character vector, it is assumed to be a file name and a corresponding file connection is created and then closed after exporting the object. If it is a BiocFile derivative, the data is loaded from or saved to the underlying resource. If missing, the function will return the output as a character vector, rather than writing to a connection.
format	The format of the output. If missing and con is a file name, the format is derived from the file extension. This argument is unnecessary when con is a derivative of BiocFile .
text	If con is missing, this can be a character vector directly providing the string data to import.
...	Additional inputs to the low level class generator functions

Details

Typically, the user will not create an object of this class directly but rather use the [TENxVisium\(\)](#) constructor function to create an object of this class in the background.

Value

TENxGeoJSON(): An object of class [TENxGeoJSON](#)
import-method: An sf and data.frame with the GeoJSON data

See Also

<https://www.10xgenomics.com/support/software/xenium-ranger/3.0/analysis/segmentation-inputs>

Examples

```
segout_folder <- system.file(
  file.path("extdata", "segmented_outputs"),
  package = "VisiumIO"
)

## import cell boundaries
cellsegs <- file.path(segout_folder, "cell_segmentations.geojson")

TENxGeoJSON(cellsegs)

TENxGeoJSON(cellsegs) |>
  import()

## import nucleus boundaries
nucsegs <- file.path(segout_folder, "nucleus_segmentations.geojson")

TENxGeoJSON(nucsegs) |>
  import()
```

TENxParquet-class	<i>Represent and import Parquet data from 10X Genomics</i>
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Description

TENxParquet is a virtual class to represent and import Parquet files from 10X Genomics. It is a composed class of [TENxIO::TENxFile](#).

TENxSpatialParquet is a class to represent and import spatial Parquet files from 10X Genomics. It is a composed class of [TENxIO::TENxFile](#).

Usage

```
TENxParquet(resource, type = c("spatial", "mapping"))

TENxMappingParquet(resource, colnames = .MAPPING_POS_COLS)

## S4 method for signature 'TENxMappingParquet,ANY,ANY'
import(con, format, text, ...)
```

Arguments

resource	character(1) The path to the file
type	character(1) A scalar specifying the type of Parquet file to import, either "spatial" or "mapping". If not provided, the type is determined by whether or not there is a "mapping" keyword in the file name.
colnames	character() A vector specifying the column names of the Parquet, defaults to all columns in the dataset.

<code>con</code>	The connection from which data is loaded or to which data is saved. If this is a character vector, it is assumed to be a file name and a corresponding file connection is created and then closed after exporting the object. If it is a BiocFile derivative, the data is loaded from or saved to the underlying resource. If missing, the function will return the output as a character vector, rather than writing to a connection.
<code>format</code>	The format of the output. If missing and <code>con</code> is a file name, the format is derived from the file extension. This argument is unnecessary when <code>con</code> is a derivative of BiocFile .
<code>text</code>	If <code>con</code> is missing, this can be a character vector directly providing the string data to import.
<code>...</code>	Additional inputs to the low level class generator functions

Details

Typically, the user will not create an object of this class directly but rather use the `TENxParquet` constructor function to create an object of either [TENxSpatialParquet](#) or [TENxMappingParquet](#) class in the background.

Value

`TENxParquet()`: An object of class [TENxSpatialParquet](#) or [TENxMappingParquet](#)

`TENxMappingParquet()`: An object of class [TENxMappingParquet](#)

Examples

```
sample_dir <- system.file(
  file.path("extdata", "binned_outputs", "square_002um", "spatial"),
  package = "VisiumIO"
)
spatial_dir <- Filter(
  function(x) endsWith(x, "spatial"), list.dirs(sample_dir)
)
parquetres <- file.path(spatial_dir, "tissue_positions.parquet")
TENxParquet(parquetres)
parq_file <- system.file(
  "extdata", "barcode_mappings.parquet",
  package = "VisiumIO", mustWork = TRUE
)
TENxMappingParquet(parq_file)
```

TENxSpatialCSV-class *Represent and import spatial CSV data from 10X Genomics*

Description

`TENxSpatialCSV` is a class to represent and import spatial CSV files with specific column names. It is a composed class of [TENxIO::TENxFile](#) and contains additional slots for the column names and whether the CSV is a list-type of file.

Usage

```
TENxSpatialCSV(resource, colnames = .TISSUE_POS_COLS)

## S4 method for signature 'TENxSpatialCSV,ANY,ANY'
import(con, format, text, ...)
```

Arguments

resource	character(1) The path to the file
colnames	character() A vector specifying the column names of the CSV, defaults to <code>c("barcode", "in_tissue", "array_row", "array_col", "pxl_row_in_fullres", "pxl_col_in_fullres")</code> . Mainly used for the "positions" CSV type of file which does not include column names in the file.
con	The connection from which data is loaded or to which data is saved. If this is a character vector, it is assumed to be a file name and a corresponding file connection is created and then closed after exporting the object. If it is a BiocFile derivative, the data is loaded from or saved to the underlying resource. If missing, the function will return the output as a character vector, rather than writing to a connection.
format	The format of the output. If missing and con is a file name, the format is derived from the file extension. This argument is unnecessary when con is a derivative of BiocFile .
text	If con is missing, this can be a character vector directly providing the string data to import.
...	Additional inputs to the low level class generator functions

Details

Typically, the user will not create an object of this class directly but rather use the [TENxVisium\(\)](#) constructor function to create an object of this class in the background. The column names are set to the default values of `c("barcode", "in_tissue", "array_row", "array_col", "pxl_row_in_fullres", "pxl_col_in_fullres")`. The column names can be changed by specifying the `colnames` argument in the constructor function.

Set the option "VisiumIO.csvreader" to either "data.table" or "readr" to use the `data.table::fread` or `readr::read_csv` functions, respectively. These options are useful when the CSV file is relatively large and the user wants to use faster read-in options. Note that the outputs will still be converted to `DataFrame` when incorporated to the `SpatialExperiment` or `SingleCellExperiment` object.

Value

TENxSpatialCSV: An object of class [TENxSpatialCSV](#)

import-method: A `DataFrame` object containing the data from the CSV file

Slots

isList logical(1) A scalar specifying whether the CSV is a list-type of file
 colnames character() A vector specifying the column names of the CSV
 variant character(1) A scalar specifying the variant of the CSV file "positions", "cell_boundaries", or "other". The variant is determined by the name of the CSV file within the constructor function. Values include "positions", "cell_boundaries", and "other".
 compressed logical(1) A scalar specifying whether the CSV is compressed (mainly with a .gz file extension).

Examples

```
sample_dir <- system.file(
  file.path("extdata", "10xVisium", "section1"),
  package = "VisiumIO"
)
spatial_dir <- Filter(
  function(x) endsWith(x, "spatial"), list.dirs(sample_dir)
)
csvresource <- file.path(spatial_dir, "tissue_positions_list.csv")
TENxSpatialCSV(csvresource)
head(import(TENxSpatialCSV(csvresource)), 4)

import(TENxSpatialCSV(csvresource)) |>
  attr("metadata") |>
  lapply(names)
```

TENxSpatialList-class *A class to represent and import spatial Visium data*

Description

This class is a composed class of [TENxFileList](#), which can contain a list of [TENxFile](#) objects, and a [TENxSpatialList](#) object. It is meant to handle spatial Visium data from 10X Genomics.

Usage

```
TENxSpatialList(
  resources,
  sample_id = "sample01",
  images = c("lowres", "hires", "detected", "aligned", "aligned_fiducials", "cytassist"),
  jsonFile = .SCALE_JSON_FILE,
  tissuePattern = "tissue_positions.*",
  bin_size = c("008", "016", "002"),
  loadImage = FALSE,
  ...
)

## S4 method for signature 'TENxSpatialList,ANY,ANY'
import(con, format, text, ...)
```

Arguments

resources	A TENxFileList object or a file path to the tarball containing the matrix / assay data resources.
sample_id	character(1) A single string specifying the sample ID.
images	character() A vector specifying the images to be imported; can be one or multiple of "lowres", "hires", "detected", "aligned".
jsonFile	character(1) A single string specifying the name of the JSON file containing the scale factors.
tissuePattern	character(1) A single string specifying the pattern to match the tissue positions file.
bin_size	character(1) An <i>optional</i> scalar indicating the image bin size in microns, e.g., '008' for 8 microns. When provided, the function will look for a subfolder in the binned_outputs folder that corresponds to the specified bin size, e.g., square_008um for an input value of '008'. Bin sizes '002', '008', and '016' are typical of the space ranger pipeline but custom bin sizes may be used. If the bin_size argument is not provided, it is set to an empty character value i.e., "" and the function will not look for any data in binned_outputs and will only import data from the main spacerangerOut directory.
loadImage	logical(1) A single logical value indicating whether to load the image data into the resulting SpatialExperiment object. This argument requires the magick package.
...	Parameters to pass to the format-specific method.
con	The connection from which data is loaded or to which data is saved. If this is a character vector, it is assumed to be a file name and a corresponding file connection is created and then closed after exporting the object. If it is a BiocFile derivative, the data is loaded from or saved to the underlying resource. If missing, the function will return the output as a character vector, rather than writing to a connection.
format	The format of the output. If missing and con is a file name, the format is derived from the file extension. This argument is unnecessary when con is a derivative of BiocFile .
text	If con is missing, this can be a character vector directly providing the string data to import.

Details

Typically, the user will not create an object of this class directly but rather use the [TENxVisium\(\)](#) constructor function to create an object of this class.

Value

A SpatialExperiment object

Methods (by generic)

- import(con = TENxSpatialList, format = ANY, text = ANY): Import a TENxSpatialList object

Slots

- `images` `character()` The image name(s) to use with `grep` and include in the list of files. Can be one of "lowres", "hires", "lowres", "hires", "detected", "aligned", "aligned_fiducials", or "cytassist".
- `scaleJSON` `character(1)` The file name of the scale factors JSON file, defaults to 'scalefactors_json.json'.
- `tissuePos` `character(1)` An optional slot indicating the file name of the tissue positions file; typically a .parquet or .csv file. To avoid import, this slot can be set to an empty character value i.e., "".
- `sampleId` `character(1)` A scalar specifying the sample identifier.
- `binSize` `character(1)` An optional slot to store the image bin size when importing. The default slot value is an empty character i.e., "". When present, the value must be a character scalar, e.g., '008' for 8 microns, and will match the directory name e.g., square_008um.
- `loadImage` `logical(1)` Whether to load the images into memory as `SpatialImage` objects. If FALSE, the images are stored as file paths and loaded as `StoredSpatialImage` objects when the `TENxSpatialList` object is imported. The default is FALSE to avoid loading large images into memory. This functionality requires the `magick` package.

Examples

```
spatial_dir <- system.file(
  file.path("extdata", "10xVisium", "section1", "outs", "spatial"),
  package = "VisiumIO"
)

TENxSpatialList(resources = spatial_dir, images = "lowres")

TENxSpatialList(resources = spatial_dir, images = "lowres") |>
  metadata() |> lapply(names)

TENxSpatialList(resources = spatial_dir, images = "lowres") |>
  import()
```

TENxSpatialParquet-class

Represent and import spatial Parquet data from 10X Genomics

Description

TENxSpatialParquet is a class to represent and import spatial Parquet files with specific column names. It is a composed class of [TENxIO::TENxFile](#) and contains additional slots for the column names and whether the Parquet is a list-type of file.

Usage

```
TENxSpatialParquet(resource, colnames)

## S4 method for signature 'TENxSpatialParquet,ANY,ANY'
import(con, format, text, ...)
```

Arguments

resource	character(1) The path to the file
colnames	character() A vector specifying the column names of the Parquet, defaults to all columns in the dataset.
con	The connection from which data is loaded or to which data is saved. If this is a character vector, it is assumed to be a file name and a corresponding file connection is created and then closed after exporting the object. If it is a BiocFile derivative, the data is loaded from or saved to the underlying resource. If missing, the function will return the output as a character vector, rather than writing to a connection.
format	The format of the output. If missing and con is a file name, the format is derived from the file extension. This argument is unnecessary when con is a derivative of BiocFile .
text	If con is missing, this can be a character vector directly providing the string data to import.
...	Additional inputs to the low level class generator functions

Details

Typically, the user will not create an object of this class directly but rather use the [TENxVisium\(\)](#) constructor function to create an object of this class in the background. The column names are set to the default values of `c("barcode", "in_tissue", "array_row", "array_col", "pxl_row_in_fullres", "pxl_col_in_fullres")`. The column names can be changed by specifying the `colnames` argument in the constructor function.

Value

`TENxSpatialParquet()`: An object of class [TENxSpatialParquet](#)
`import-method`: A tibble object containing the data from the Parquet file

Examples

```
sample_dir <- system.file(
  file.path("extdata", "binned_outputs", "square_002um", "spatial"),
  package = "VisiumIO"
)
spatial_dir <- Filter(
  function(x) endsWith(x, "spatial"), list.dirs(sample_dir)
)
parquetres <- file.path(spatial_dir, "tissue_positions.parquet")
TENxSpatialParquet(parquetres)
```

```
TENxSpatialParquet(parquetres) |>
  import()

## metadata in attributes
TENxSpatialParquet(parquetres) |>
  import() |>
  attr("metadata") |>
  lapply(names)
```

TENxVisium-class	<i>A class to represent and import a single Visium Sample</i>
------------------	---

Description

This class is a composed class of [TENxFileList](#) which can contain a list of [TENxFile](#) objects and a [TENxSpatialList](#) object. It is meant to handle a single Visium sample from 10X Genomics.

Usage

```
TENxVisium(
  resources,
  spatialResource,
  spacerangerOut,
  sample_id = "sample01",
  processing = c("filtered", "raw"),
  format = c("mtx", "h5"),
  images = c("lowres", "hires", "detected", "aligned", "cytassist"),
  jsonFile = .SCALE_JSON_FILE,
  tissuePattern = "tissue_positions.*\\.csv",
  spatialCoordsNames = c("pxl_col_in_fullres", "pxl_row_in_fullres"),
  loadImage = FALSE,
  ...
)

## S4 method for signature 'TENxVisium,ANY,ANY'
import(con, format, text, ...)
```

Arguments

resources	A TENxFileList object or a file path to the tarball containing the matrix / assay data resources.
spatialResource	A TENxSpatialList object or a file path to the tarball containing the spatial data.
spacerangerOut	character(1) A single string specifying the path to the directory where the output of spaceranger count is located; typically (but not necessarily), this is the outs directory. The directory must contain the (processing)_feature_bc_matrix and spatial sub directories.

sample_id	character(1) A single string specifying the sample ID.
processing	character(1) A single string indicating the processing folder available e.g., "filtered_feature_barcode_matrix" in the spacerangerOut folder. It can be either "filtered" or "raw" (default "filtered"). Only used when spacerangerOut is specified.
format	The format of the output. If missing and con is a file name, the format is derived from the file extension. This argument is unnecessary when con is a derivative of BiocFile .
images	character() A vector specifying the images to be imported; can be one or multiple of "lowres", "hires", "detected", "aligned".
jsonFile	character(1) A single string specifying the name of the JSON file containing the scale factors.
tissuePattern	character(1) A single string specifying the pattern to match the tissue positions file.
spatialCoordsNames	character() A vector of strings specifying the names of the columns in the spatial data containing the spatial coordinates.
loadImage	logical(1) A single logical value indicating whether to load the image data into the resulting SpatialExperiment object. This argument requires the magick package.
...	In the constructor, additional arguments passed to TENxFileList ; otherwise, not used.
con	The connection from which data is loaded or to which data is saved. If this is a character vector, it is assumed to be a file name and a corresponding file connection is created and then closed after exporting the object. If it is a BiocFile derivative, the data is loaded from or saved to the underlying resource. If missing, the function will return the output as a character vector, rather than writing to a connection.
text	If con is missing, this can be a character vector directly providing the string data to import.

Details

Typically, the user will not create an object of this class directly but rather use [TENxVisiumList](#) constructor function for multiple samples. Note that the images, jsonFile, tissuePattern, and spatialCoordsNames arguments are only considered when the spacerangerOut argument or both the resources and spatialResource arguments are paths to files.

Value

A [SpatialExperiment](#) object

Functions

- `import(con = TENxVisium, format = ANY, text = ANY)`: Import Visium data

Slots

resources A [TENxFileList](#) or [TENxH5](#) object containing the Visium data.

spatialList A [TENxSpatialList](#) object containing the spatial

coordNames character() A vector specifying the names of the columns in the spatial data containing the spatial coordinates.

sampleId character(1) A scalar specifying the sample identifier.

loadImage logical(1) Whether to load the images into memory as SpatialImage objects. If FALSE, the images are stored as file paths and loaded as StoredSpatialImage objects when the TENxSpatialList object is imported. The default is FALSE to avoid loading large images into memory. This functionality requires the magick package.

See Also

<https://support.10xgenomics.com/spatial-gene-expression/software/pipelines/latest/output/overview>

Examples

```
outs_dir <- system.file(
  file.path("extdata", "10xVisium", "section1", "outs"),
  package = "VisiumIO"
)

## using spacerangerOut folder
tv <- TENxVisium(
  spacerangerOut = outs_dir, processing = "raw", images = "lowres"
)

import(tv)

## with TENxFileList spacerangerOut input
tvfl <- TENxVisium(
  spacerangerOut = TENxFileList(outs_dir),
  format = "mtx",
  processing = "raw",
  images = "lowres"
)

import(tvfl)

## check metadata of the object
import(tvfl) |>
  metadata() |>
  lapply(names)

## importing h5 format
tvfl <- TENxVisium(
  spacerangerOut = outs_dir,
  format = "h5",
  processing = "raw",
```

```

    images = "lowres"
)

import(tvfl)

rffolder <- file.path(outs_dir, "raw_feature_bc_matrix")
## using resources and spatialResource inputs
tvfl <- TENxVisium(
  resources = rffolder,
  spatialResource = file.path(dirname(rffolder), "spatial"),
  format = "mtx",
  processing = "raw",
  images = "lowres"
)

import(tvfl)

```

TENxVisiumHD-class	<i>A class to represent and import multiple Visium HD samples</i>
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Description

This class contains a SimpleList of [TENxVisiumHD](#) objects each corresponding to one sample. The provided spacerangerOut folder should contain a binned_outputs folder where multiple bin_size subfolders are present, e.g., square_002um. Note that the sf package is required to import cell segmentation and nucleus segmentation data. If the sf package is not installed, the cell segmentation data will not be imported and an error is issued.

Usage

```

TENxVisiumHD(
  resources,
  spatialResource,
  spacerangerOut,
  segmented_outputs,
  sample_id = "sample01",
  processing = c("filtered", "raw"),
  format = c("mtx", "h5"),
  images = c("lowres", "hires", "detected", "aligned_fiducials"),
  bin_size = c("008", "016", "002"),
  jsonFile = .SCALE_JSON_FILE,
  tissuePattern = "tissue_positions\\.parquet",
  spatialCoordsNames = c("pxl_col_in_fullres", "pxl_row_in_fullres"),
  mappingPattern = "barcode_mappings\\.parquet",
  boundary = c("cell_segmentations", "nucleus_segmentations", "both"),
  loadImage = FALSE,
  ...

```

```
)

## S4 method for signature 'TENxVisiumHD,ANY,ANY'
import(con, format, text, ...)
```

Arguments

resources	A TENxFileList object or a file path to the tarball containing the matrix / assay data resources.
spatialResource	A TENxSpatialList object or a file path to the tarball containing the spatial data.
spacerangerOut	character(1) A single string specifying the path to the directory where the output of spaceranger count is located; typically (but not necessarily), this is the outs directory. The directory must contain the (processing)_feature_bc_matrix and spatial sub directories.
segmented_outputs	character(1) The path to the segmented_outputs directory
sample_id	character(1) A single string specifying the sample ID.
processing	character(1) A single string indicating the processing folder available e.g., "filtered_feature_barcode_matrix" in the spacerangerOut folder. It can be either "filtered" or "raw" (default "filtered"). Only used when spacerangerOut is specified.
format	The format of the output. If missing and con is a file name, the format is derived from the file extension. This argument is unnecessary when con is a derivative of BiocFile .
images	character() A vector specifying the images to be imported; can be one or multiple of "lowres", "hires", "detected", "aligned".
bin_size	character(1) An <i>optional</i> scalar indicating the image bin size in microns, e.g., '008' for 8 microns. When provided, the function will look for a subfolder in the binned_outputs folder that corresponds to the specified bin size, e.g., square_008um for an input value of '008'. Bin sizes '002', '008', and '016' are typical of the space ranger pipeline but custom bin sizes may be used. If the bin_size argument is not provided, it is set to an empty string "" and the function will not look for any data in binned_outputs and will only import data from the main spacerangerOut directory.
jsonFile	character(1) A single string specifying the name of the JSON file containing the scale factors.
tissuePattern	character(1) A single string specifying the pattern to match the tissue positions file.
spatialCoordsNames	character() A vector of strings specifying the names of the columns in the spatial data containing the spatial coordinates.
mappingPattern	character(1) The pattern used in list.files that identifies the mapping file. The default is "barcode_mappings\\.parquet".

boundary	character(1) The type of segmentation boundary to use. The options are "cell_segmentations" (default), "nucleus_segmentations", or "both". When "both" is specified, "cell_segmentations" are added to the spatialCoords and the nucleus_segmentations centroids (labeled x.nuc and y.nuc) are added to the colData of the returned object.
loadImage	logical(1) A single logical value indicating whether to load the image data into the resulting SpatialExperiment object. This argument requires the magick package.
...	In the constructor, additional arguments passed to TENxFileList ; otherwise, not used.
con	The connection from which data is loaded or to which data is saved. If this is a character vector, it is assumed to be a file name and a corresponding file connection is created and then closed after exporting the object. If it is a BiocFile derivative, the data is loaded from or saved to the underlying resource. If missing, the function will return the output as a character vector, rather than writing to a connection.
text	If con is missing, this can be a character vector directly providing the string data to import.

Details

Typically, the user will provide a path to a directory containing the output of the `spaceranger count` command. The `spaceranger count` command outputs a folder containing the "raw" or "filtered" `()_feature_bc_matrix`.

Note that `nucleus_segmentations.geojson` file must be in the same folder as the `cell_segmentations.geojson` file for the nucleus centroids to be imported correctly when selecting the "both" for the boundary argument.

Value

A [SpatialExperiment](#) object

Functions

- `import(con = TENxVisiumHD, format = ANY, text = ANY)`: Import Visium HD data from multiple bin sizes

Author(s)

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Examples

```
vdir <- system.file(
  "extdata", package = "VisiumIO", mustWork = TRUE
)

## with spacerangerOut folder
TENxVisiumHD(spacerangerOut = vdir, bin_size = "002", images = "lowres")
```

```

TENxVisiumHD(spacerangerOut = vdir, bin_size = "002", images = "lowres") |>
  import()

## indicate h5 format
TENxVisiumHD(
  spacerangerOut = vdir, bin_size = "002",
  images = "lowres", format = "h5"
)

TENxVisiumHD(
  spacerangerOut = vdir, bin_size = "002",
  images = "lowres", format = "h5"
) |>
  import()

## use resources and spatialResource arguments as file paths
TENxVisiumHD(
  resources = file.path(
    vdir, "binned_outputs", "square_002um",
    "filtered_feature_bc_matrix.h5"
  ),
  spatialResource = file.path(
    vdir, "binned_outputs", "square_002um",
    "spatial"
  ),
  bin_size = "002", processing = "filtered",
  images = "lowres", format = "h5"
) |>
  import()

## provide the spatialResource argument as a TENxFileList
TENxVisiumHD(
  resources = file.path(
    vdir, "binned_outputs", "square_002um",
    "filtered_feature_bc_matrix.h5"
  ),
  spatialResource = TENxFileList(
    file.path(
      vdir, "binned_outputs", "square_002um",
      "spatial"
    )
  ),
  bin_size = "002", images = "lowres", format = "h5"
) |>
  import()

## with segmented_outputs folder
seg_outs <- system.file(
  "extdata", "segmented_outputs", package = "VisiumIO", mustWork = TRUE
)
TENxVisiumHD(
  segmented_outputs = seg_outs,

```

```

    format = "h5",
    images = "lowres"
) |>
  import()

```

TENxVisiumList-class *A class to represent and import multiple Visium samples*

Description

This class contains a SimpleList of [TENxVisium](#) objects each corresponding to one sample.

Usage

```

TENxVisiumList(
  sampleFolders,
  sample_ids,
  processing = c("filtered", "raw"),
  images = c("lowres", "hires", "detected", "aligned"),
  format = c("mtx", "h5"),
  jsonFile = .SCALE_JSON_FILE,
  tissuePattern = "tissue_positions.*\\.csv",
  spatialCoordsNames = c("pxl_col_in_fullres", "pxl_row_in_fullres"),
  ...
)

## S4 method for signature 'TENxVisiumList,ANY,ANY'
import(con, format, text, ...)

```

Arguments

sampleFolders	character() A vector of strings specifying the directories containing the output of the spaceranger count command.
sample_ids	character() A vector of strings specifying the sample IDs. If not provided, the sample IDs will be the names of the sampleFolders. Therefore, the sample_ids must be the same length as sampleFolders.
processing	character(1) A single string indicating the processing folder available e.g., "filtered_feature_barcode_matrix" in the spacerangerOut folder. It can be either "filtered" or "raw" (default "filtered"). Only used when spacerangerOut is specified.
images	character() A vector specifying the images to be imported; can be one or multiple of "lowres", "hires", "detected", "aligned".
format	The format of the output. If missing and con is a file name, the format is derived from the file extension. This argument is unnecessary when con is a derivative of BiocFile .

jsonFile	character(1) A single string specifying the name of the JSON file containing the scale factors.
tissuePattern	character(1) A single string specifying the pattern to match the tissue positions file.
spatialCoordsNames	character() A vector of strings specifying the names of the columns in the spatial data containing the spatial coordinates.
...	In the constructor, additional arguments passed to TENxFileList ; otherwise, not used.
con	The connection from which data is loaded or to which data is saved. If this is a character vector, it is assumed to be a file name and a corresponding file connection is created and then closed after exporting the object. If it is a BiocFile derivative, the data is loaded from or saved to the underlying resource. If missing, the function will return the output as a character vector, rather than writing to a connection.
text	If con is missing, this can be a character vector directly providing the string data to import.

Details

Typically, the user will provide a path to a directory containing the output of the `spaceranger count` command. The `spaceranger count` command outputs a folder containing the "raw" or "filtered" `()_feature_bc_matrix`.

Value

A [SpatialExperiment](#) object

Functions

- `import(con = TENxVisiumList, format = ANY, text = ANY)`: Import multiple Visium samples

See Also

<https://support.10xgenomics.com/spatial-gene-expression/software/pipelines/latest/output/overview>

Examples

```
sample_dirs <- list.dirs(
  system.file(
    file.path("extdata", "10xVisium"),
    package = "VisiumIO"
  ),
  recursive = FALSE, full.names = TRUE
)

tv1 <- TENxVisiumList(
```

```
        sampleFolders = sample_dirs,  
        sample_ids = c("sample01", "sample02"),  
        processing = "raw",  
        images = "lowres",  
        format = "mtx"  
    )  
  
import(tv1)
```

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