

# Package: RadioGx (via r-universe)

May 30, 2026

**Type** Package

**Title** Analysis of Large-Scale Radio-Genomic Data

**Version** 2.17.0

**Date** 2023-04-19

**Description** Computational tool box for radio-genomic analysis which integrates radio-response data, radio-biological modelling and comprehensive cell line annotations for hundreds of cancer cell lines. The 'RadioSet' class enables creation and manipulation of standardized datasets including information about cancer cells lines, radio-response assays and dose-response indicators. Included methods allow fitting and plotting dose-response data using established radio-biological models along with quality control to validate results. Additional functions related to fitting and plotting dose response curves, quantifying statistical correlation and calculating area under the curve (AUC) or survival fraction (SF) are included. For more details please see the included documentation, references, as well as: Manem, V. et al (2018) <[doi:10.1101/449793](https://doi.org/10.1101/449793)>.

**License** GPL-3

**VignetteBuilder** knitr

**VignetteEngine** knitr::rmarkdown

**Encoding** UTF-8

**LazyData** true

**Depends** R (>= 4.1), CoreGx

**Imports** SummarizedExperiment, BiocGenerics, data.table, S4Vectors, Biobase, parallel, BiocParallel, RColorBrewer, caTools, magicaxis, methods, reshape2, scales, grDevices, graphics, stats, utils, assertthat, matrixStats, downloader

**Suggests** rmarkdown, BiocStyle, knitr, pander, markdown

**biocViews** Software, Pharmacogenetics, QualityControl, Survival, Pharmacogenomics, Classification

**RoxygenNote** 7.2.3

**Collate** 'OERmodel.R' 'RadioSet-class.R' 'RadioSet-accessors.R'  
 'computeAUC.R' 'computeD10.R' 'computeSF2.R'  
 'computeSensitivity.R' 'datasets.R' 'doseResponseCurve.R'  
 'downloadRSet.R' 'geneDrugSensitivity.R' 'linearQuadratic.R'  
 'linearQuadraticInv.R' 'linearQuadraticModel.R'  
 'makeGrittyGuess.R' 'plotCurve.R' 'radSensitivitySig.R'  
 'rankGeneRadSensitivity.R' 'signatureClass.R'  
 'subsetTo-methods.R' 'summarizeMolecularProfiles-methods.R'  
 'summarizeSensitivityProfiles-methods.R'  
 'updateObject-methods.R' 'utilities.R' 'zzz.R'

**Config/pak/sysreqs** cmake libglpk-dev make libicu-dev libuv1-dev libxml2-dev zlib1g-dev

**Repository** <https://bioc.r-universe.dev>

**Date/Publication** 2026-04-28 12:52:59 UTC

**RemoteUrl** <https://github.com/bioc/RadioGx>

**RemoteRef** HEAD

**RemoteSha** 238a2e31cef8465cd3389f3f09b4f1ec857149bd

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[,RadioSet,ANY,ANY,ANY-method  
'['

---

### Description

'['

### Usage

```
## S4 method for signature 'RadioSet,ANY,ANY,ANY'  
x[i, j, ..., drop = FALSE]
```

### Arguments

x	object
i	Cell lines to keep in object
j	Drugs to keep in object
...	further arguments
drop	A boolean flag of whether to drop single dimensions or not

### Value

Returns the subsetted object

### Examples

```
data(clevelandSmall)  
clevelandSmall[sampleNames(clevelandSmall)[1], treatmentNames(clevelandSmall)[1]]
```

---

availableRSets            *Return a table of RadioSets available for download*

---

### Description

The function fetches a table of all RadioSets available for download from the PharmacoGx server. The table includes the names of the PharamcoSet, the types of data available in the object, and the date of last update.

### Usage

```
availableRSets(  
  saveDir = tempdir(),  
  fileName = "availableRadioSets.csv",  
  verbose = TRUE  
)
```

**Arguments**

saveDir	character	Directory to save the table of rSets
fileName	character	The filename for the table of rSets
verbose	bool	Should status messages be printed during download.

**Value**

A data.frame with details about the available RadioSet objects

**Examples**

```
if (interactive()){
  availableRSets()
}
```

---

checkRSetStructure     *A function to verify the structure of a RadioSet*

---

**Description**

This function checks the structure of a PharamcoSet, ensuring that the correct annotations are in place and all the required slots are filled so that matching of cells and radiations can be properly done across different types of data and with other studies.

**Usage**

```
checkRSetStructure(object, plotDist = FALSE, result.dir = tempdir())
```

**Arguments**

object	A RadioSet object
plotDist	Should the function also plot the distribution of molecular data?
result.dir	The path to the directory for saving the plots as a string, defaults to 'tempdir()'

**Value**

Prints out messages whenever describing the errors found in the structure of the pset object passed in.

**Examples**

```
checkRSetStructure(clevelandSmall)
```

---

clevelandSmall	<i>Cleveland_mut RadioSet subsetted</i>
----------------	---

---

**Description**

Documentation for this dataset will be added at a later date. For now I just need this package to pass the CRAN checks! This dataset powers the exampe usage in the roxygen2 documentation for CoreGx.

**Usage**

```
data(clevelandSmall)
```

**Format**

RadioSet object

**References**

Lamb et al. The Connectivity Map: using gene-expression signatures to connect small molecules, genes, and disease. Science, 2006.

---

computeAUC	<i>computeAUC: computes AUC</i>
------------	---------------------------------

---

**Description**

This function computes the area under a dose-response curve of the form survival fraction  $SF = \exp(-\alpha * D - \beta * D^2)$ .

**Usage**

```
computeAUC(  
  D,  
  SF,  
  pars,  
  lower,  
  upper,  
  trunc = TRUE,  
  SF_as_log = FALSE,  
  area.type = c("Fitted", "Actual"),  
  verbose = TRUE  
)
```

**Arguments**

D	vector of dosages
SF	vector of survival fractions
pars	parameters (alpha, beta) in equation $y = \exp(-\alpha * x - \beta * x^2)$
lower	lower bound of dose region to compute AUC over
upper	upper bound of dose region to compute AUC over
trunc	should survival fractions be truncated downward to 1 if they exceed 1?
SF_as_log	A boolean indicating whether survival fraction is displayed on a log axis. Defaults to FALSE
area.type	should the AUC of the raw (D, SF) points be returned, or should the AUC of a curve fit to said points be returned instead?
verbose	how detailed should error and warning messages be? See details.

**Details**

If lower and/or upper are missing, the function assumes their values to be the minimum and maximum D-values, respectively. For all warnings to be silent, set trunc = FALSE. For warnings to be output, set trunc = TRUE. For warnings to be output along with the arguments that triggered them, set trunc = 2.

**Value**

numeric The area under the ROC curve

**Examples**

```
computeAUC(D=c(0.1, 0.5, 0.7, 0.9), pars=c(0.2, 0.1), lower = 0,
upper = 1) # Returns 0.7039296
```

---

computeD10

*Compute D10*

---

**Description**

This function computes the radiation dose at which only 10 cancer cells survive under the exponential model  $SF = \exp(-\alpha * D - \beta * D^2)$  given alpha and beta, where D is the radiation dose given and SF is the fraction of cells surviving

**Usage**

```
computeD10(pars)
```

**Arguments**

pars	parameters (alpha, beta) in equation $y = \exp(-\alpha * x - \beta * x^2)$
------	--

**Details**

The units of the returned dose are the inverses of the units of the alpha and beta passed in.

**Value**

numeric The D10 value

**Examples**

computeD10(c(0.2, 0.1))

---

computeSF2

*Compute SF2*

---

**Description**

This function computes the survival fraction after administering 2 units of radiation, given alpha and beta in the equation  $SF = \exp(-\alpha * D - \beta * D ^ 2)$ .

**Usage**

computeSF2(pars)

**Arguments**

pars            parameters (alpha, beta) in equation  $y = \exp(-\alpha * x - \beta * x ^ 2)$

**Value**

numeric The survival fraction

**Examples**

computeSF2(c(0.2, 0.1))

---

dim, RadioSet-method    *Get the dimensions of a RadioSet*

---

### Description

Get the dimensions of a RadioSet

### Usage

```
## S4 method for signature 'RadioSet'  
dim(x)
```

### Arguments

x                      RadioSet

### Value

A named vector with the number of Cells and Drugs in the RadioSet

### Examples

```
data(clevelandSmall)  
dim(clevelandSmall)
```

---

doseResponseCurve    *Plot drug response curve of a given drug and a given cell for a list of rSets (objects of the RadioSet class).*

---

### Description

Given a list of RadioSets, the function will plot the drug\_response curve, for a given drug/cell pair. The y axis of the plot is the viability percentage and x axis is the log transformed Ds. If more than one rSet is provided, a light gray area would show the common concentration range between rSets. User can ask for type of sensitivity measurement to be shown in the plot legend. The user can also provide a list of their own Ds and viability values, as in the examples below, and it will be treated as experiments equivalent to values coming from a pset. The names of the concentration list determine the legend labels.

**Usage**

```
doseResponseCurve(
  rad.type = "radiation",
  cellline,
  rSets = list(),
  Ds = list(),
  SFs = list(),
  trunc = TRUE,
  legends.label = c("alpha", "beta", "rsquared"),
  ylim = c(0, 100),
  xlim,
  mycol,
  title,
  plot.type = c("Fitted", "Actual", "Both"),
  summarize.replicates = TRUE,
  lwd = 1,
  cex = 0.7,
  cex.main = 0.9,
  legend.loc = "topright",
  verbose = TRUE
)
```

**Arguments**

rad.type	‘character(1)’ The type(s) of radiation dosage to be plotted. If the plot is desirable for more than one radioset, A unique drug id should be provided.
cellline	‘character(1)’ A cell line name for which the radiation response curve should be plotted. If the plot is desirable for more than one radioset, a unique cell id should be provided.
rSets	‘list’ a list of RadioSet objects, for which the function should plot the curves.
Ds, SFs	‘list’ A list of Doses and SFs to plot, the function assumes that Ds[[i]] is plotted against SFs[[i]]. The names of the D list are used to create the legend labels
trunc	‘logical(1)’ Should the viability values be truncated to lie in [0-100] before doing the fitting
legends.label	‘numeric’ A vector of sensitivity measurement types which could be any combination of ic50_published, auc_published, auc_recomputed and auc_recomputed_star. A legend will be displayed on the top right of the plot which each line of the legend is the values of requested sensitivity measurements for one of the requested rSets. If this parameter is missed no legend would be provided for the plot.
ylim	‘numeric’ A vector of two numerical values to be used as ylim of the plot. If this parameter would be missed c(0,100) would be used as the ylim of the plot.
xlim	‘numeric’ A vector of two numerical values to be used as xlim of the plot. If this parameter would be missed the minimum and maximum concentrations between all the rSets would be used as plot xlim.
mycol	‘numeric’ A vector with the same length of the rSets parameter which will determine the color of the curve for the pharmaco sets. If this parameter is missed default colors from Rcolorbrewer package will be used as curves color.

<code>title</code>	‘character’ The title of the graph. If no title is provided, then it defaults to Drug’:’Cell Line’.
<code>plot.type</code>	‘character’ Plot type which can be the actual one ("Actual") or the one fitted by log1 logistic regression ("Fitted") or both of them ("Both"). If this parameter is missed by default actual curve is plotted.
<code>summarize.replicates</code>	‘character’ If this parameter is set to true replicates are summarized and replicates are plotted individually otherwise
<code>lwd</code>	‘numeric’ The line width to plot with
<code>cex</code>	‘numeric’ The cex parameter passed to plot
<code>cex.main</code>	‘numeric’ The cex.main parameter passed to plot, controls the size of the titles
<code>legend.loc</code>	And argument passable to xy.coords for the position to place the legend.
<code>verbose</code>	‘logical(1)’ Should warning messages about the data passed in be printed?

**Value**

Plots to the active graphics device and returns an invisible NULL.

**Examples**

```
doseResponseCurve(Ds=list("Experiment 1" = c(0, 2, 4, 6)),
  SFs=list("Experiment 1" = c(1,.6,.4,.2)), plot.type="Both")
```

---

downloadRSet

*Download a RadioSet object*

---

**Description**

This function allows you to download a RadioSet object for use with this package. The RadioSets have been extensively curated and organized within a PharacoSet class, enabling use with all the analysis tools provided in PharmacGx.

**Usage**

```
downloadRSet(
  name,
  saveDir = tempdir(),
  rSetFileName = NULL,
  verbose = TRUE,
  timeout = 600
)
```

**Arguments**

name	Character string, the name of the PhamracoSet to download.
saveDir	Character string with the folder path where the RadioSet should be saved. Defaults to './rSets/'. Will create directory if it does not exist.
rSetFileName	character string, the file name to save the dataset under
verbose	bool Should status messages be printed during download. Defaults to TRUE.
timeout	'numeric(1)' How long to wait before erroring due to timeout, in seconds. Default it 600s (10 minutes).

**Value**

A rSet object with the dataset, downloaded from our server

**Examples**

```
if (interactive()) {
  cleveland <- downloadRSet("Cleveland")
}
```

---

linearQuadraticModel *Fit linear-quadratic curves to dose-response data*

---

**Description**

This function fits a linear-quadratic curve to dose-response data.

**Usage**

```
linearQuadraticModel(
  D,
  SF,
  lower_bounds = c(0, 0),
  upper_bounds = c(1, 1),
  scale = 5,
  family = c("normal", "Cauchy"),
  median_n = 1,
  trunc = FALSE,
  verbose = FALSE
)
```

**Arguments**

D	vector of radiation doses
SF	vector of survival fractions corresponding to the doses
lower_bounds	vector of length 2 containing minimum allowed values of fitted alpha and beta, respectively
upper_bounds	vector of length 2 containing maximum allowed values of fitted alpha and beta, respectively
scale	parameter of the assumed error distribution of the data; see sdetails
family	family of distributions of the error terms in the data; currently supported options are "normal" and "cauchy"
median_n	see details
trunc	should survival fractions be truncated downward to 1? Defaults to FALSE.
verbose	'verbose' outputs warnings that are otherwise suppressed when the function sanity-checks user inputs. 'median_n' denotes the number of distributions from family 'family' that are medianned. (Note that setting n = 1 (the default) is equivalent to using a simple normal or cauchy distribution without taking any medians.)

**Value**

numeric The estimated alpha and beta values

**Examples**

```
linearQuadraticModel(c(0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10),
  c(1.1, 0.8, 0.7, 0.45, 0.15, -0.1, -0.1, -0.4, -0.65, -0.75, -1.1))
```

---

plotCurve

*Plot radiation dose-response curve*

---

**Description**

This function plots doses of radiation against the cancer cell survival fractions thereby observed.

**Usage**

```
plotCurve(
  D,
  SF,
  pars,
  filename = "dose_response_plot.pdf",
  fit_curve = TRUE,
  SF_as_log = TRUE
)
```

**Arguments**

D	vector of radiation doses
SF	vector of survival fractions corresponding to the doses
pars	parameters (alpha, beta) in the equation $SF = \exp(-\alpha * D - \beta * D^2)$
filename	name of PDF which will be created by the function
fit_curve	should the graph include a linear-quadratic curve of best fit? Defaults to TRUE
SF_as_log	should SF be expressed in log10 on the graph? Defaults to TRUE

**Value**

nothing Function works by side effects only

**Examples**

```
plotCurve(c(0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10),
  c(1.1, 0.8, 0.7, 0.45, 0.15, -0.1, -0.1, -0.4, -0.65, -0.75, -1.1),
  filename = NULL)
```

---

RadioSet

*RadioSet constructor*


---

**Description**

A constructor that simplifies the process of creating RadioSets, as well as creates empty objects for data not provided to the constructor. Only objects returned by this constructor are expected to work with the RadioSet methods. For a much more detailed instruction on creating RadioSets, please see the "CreatingRadioSet" vignette.

**Usage**

```
RadioSet(
  name,
  molecularProfiles = list(),
  sample = data.frame(),
  treatment = data.frame(),
  sensitivityInfo = data.frame(),
  sensitivityRaw = array(dim = c(0, 0, 0)),
  sensitivityProfiles = matrix(),
  sensitivityN = matrix(nrow = 0, ncol = 0),
  perturbationN = array(NA, dim = c(0, 0, 0)),
  curationSample = data.frame(),
  curationTissue = data.frame(),
  curationTreatment = data.frame(),
  datasetType = c("sensitivity", "perturbation", "both"),
  verify = TRUE
)
```

**Arguments**

name	A character string detailing the name of the dataset
molecularProfiles	A list of SummarizedExperiment objects containing molecular profiles for each molecular data type.
sample	A data.frame containing the annotations for all the sample profiled in the data set, across all data types. Must contain the mandatory sampleid column which uniquely identifies each sample in the object.
treatment	A data.frame containing annotations for all treatments profiled in the dataset. Must contain the mandatory treatmentid column which uniquely identifies each treatment in the object.
sensitivityInfo	A data.frame containing the information for the sensitivity experiments. Must contain a 'sampleid' column with unique identifiers to each sample, matching the sample object and a 'treatmentid' columns with unique indenifiers for each treatment, matching the treatment object.
sensitivityRaw	A 3 Dimensional array containng the raw drug dose response data for the sensitivity experiments
sensitivityProfiles	data.frame containing drug sensitivity profile statistics such as IC50 and AUC
sensitivityN, perturbationN	A data.frame summarizing the available sensitivity/perturbation data
curationSample, curationTissue, curationTreatment	A data.frame mapping the names for samples, tissues and treatments used in the data set to universal identifiers used between different CoreSet objects
datasetType	A character(1) string of 'sensitivity', 'preturbation', or 'both' detailing what type of data can be found in the CoreSet, for proper processing of the data
verify	logical(1)Should the function verify the CoreSet and print out any errors it finds after construction?

**Value**

An object of class 'RadioSet'

---

RadioSet-accessors      *Accessing and modifying information in a 'RadioSet'*

---

**Description**

Documentation for the various setters and getters which allow manipulation of data in the slots of a 'RadioSet' object.

**Usage**

```
radiationInfo(...)

radiationInfo(...) <- value

radiationTypes(...)

radiationTypes(...) <- value

## S4 method for signature 'RadioSet'
annotation(object)

## S4 replacement method for signature 'RadioSet,list'
annotation(object) <- value

## S4 method for signature 'RadioSet'
dateCreated(object)

## S4 replacement method for signature 'RadioSet,character'
dateCreated(object) <- value

## S4 method for signature 'RadioSet'
name(object)

## S4 replacement method for signature 'RadioSet'
name(object) <- value

## S4 method for signature 'RadioSet'
sampleInfo(object)

## S4 replacement method for signature 'RadioSet,data.frame'
sampleInfo(object) <- value

## S4 method for signature 'RadioSet'
sampleNames(object)

## S4 replacement method for signature 'RadioSet,character'
sampleNames(object) <- value

## S4 method for signature 'RadioSet'
curation(object)

## S4 replacement method for signature 'RadioSet,list'
curation(object) <- value

## S4 method for signature 'RadioSet'
datasetType(object)
```

```
## S4 replacement method for signature 'RadioSet,character'  
datasetType(object) <- value  
  
## S4 method for signature 'RadioSet'  
molecularProfiles(object, mDataType, assay)  
  
## S4 replacement method for signature 'RadioSet,character,character,matrix'  
molecularProfiles(object, mDataType, assay) <- value  
  
## S4 method for signature 'RadioSet'  
featureInfo(object, mDataType)  
  
## S4 replacement method for signature 'RadioSet,character,data.frame'  
featureInfo(object, mDataType) <- value  
  
## S4 method for signature 'RadioSet,character'  
phenoInfo(object, mDataType)  
  
## S4 replacement method for signature 'RadioSet,character,data.frame'  
phenoInfo(object, mDataType) <- value  
  
## S4 method for signature 'RadioSet,character'  
fNames(object, mDataType)  
  
## S4 replacement method for signature 'RadioSet,character,character'  
fNames(object, mDataType) <- value  
  
## S4 method for signature 'RadioSet'  
mDataNames(object)  
  
## S4 replacement method for signature 'RadioSet'  
mDataNames(object) <- value  
  
## S4 method for signature 'RadioSet'  
molecularProfilesSlot(object)  
  
## S4 replacement method for signature 'RadioSet,list_OR_MAE'  
molecularProfilesSlot(object) <- value  
  
## S4 method for signature 'RadioSet'  
sensitivityInfo(object, dimension, ...)  
  
## S4 replacement method for signature 'RadioSet,data.frame'  
sensitivityInfo(object, dimension, ...) <- value  
  
## S4 method for signature 'RadioSet'  
sensitivityMeasures(object)
```

```

## S4 replacement method for signature 'RadioSet,character'
sensitivityMeasures(object) <- value

## S4 method for signature 'RadioSet'
sensitivityProfiles(object)

## S4 replacement method for signature 'RadioSet,data.frame'
sensitivityProfiles(object) <- value

## S4 method for signature 'RadioSet'
sensitivityRaw(object)

## S4 replacement method for signature 'RadioSet,array'
sensitivityRaw(object) <- value

## S4 method for signature 'RadioSet'
treatmentResponse(object)

## S4 replacement method for signature 'RadioSet,list_OR_LongTable'
treatmentResponse(object) <- value

## S4 method for signature 'RadioSet'
sensNumber(object)

## S4 replacement method for signature 'RadioSet,matrix'
sensNumber(object) <- value

## S4 method for signature 'RadioSet'
pertNumber(object)

## S4 replacement method for signature 'RadioSet,array'
pertNumber(object) <- value

```

## Arguments

...	See details.
value	See details.
object	A 'RadioSet' object.
mDataType	'character(1)' The name of a molecular datatype to access from the 'molecularProfiles' of a 'RadioSet' object.
assay	'character(1)' A valid assay name in the 'SummarizedExperiment' of '@molecularProfiles' of a RadioSet object for data type 'mDataType'.
dimension	See details.

## Details

**treatmentInfo:** data.frame Metadata for all treatments in a RadioSet object. Arguments:

- **object:** RadioSet An object to retrieve treatment metadata from.

**treatmentInfo<-:** RadioSet object with updated treatment metadata. **object.** Arguments:

- **object:** RadioSet An object to set treatment metadata for.
- **value:** data.frame A new table of treatment metadata for object.

**treatmentNames:** character Names for all treatments in a RadioSet object. Arguments:

- **object:** RadioSet An object to retrieve treatment names from.

**treatmentNames<-:** RadioSet Object with updates treatment names. **object.** Arguments:

- **object:** RadioSet An object to set treatment names from.
- **value:** character A character vector of updated treatment names.

#### **@annotation:**

**annotation:** A list of RadioSet annotations with items: 'name', the name of the object; 'dateCreated', date the object was created; 'sessionInfo', the sessionInfo() when the object was created; 'call', the R constructor call; and 'version', the object version.

**annotation<-:** Setter method for the annotation slot. Arguments:

- **value:** a list of annotations to update the RadioSet with.

#### **@dateCreated:**

**dateCreated:** character(1) The date the RadioSet object was created, as returned by the date() function.

**dateCreated<-:** Update the 'dateCreated' item in the annotation slot of a RadioSet object. Arguments:

- **value:** A character(1) vector, as returned by the date() function.

**name:** character(1) The name of the RadioSet, retrieved from the @annotation slot.

**name<-:** Update the @annotation\$name value in a RadioSet object.

- **value:** character(1) The name of the RadioSet object.

**cellInfo:** data.frame Metadata for all sample in a RadioSet object.

**sampleInfo<-:** assign updated sample annotations to the RadioSet object. Arguments:

- **value:** a data.frame object.

**sampleNames:** character Retrieve the rownames of the data.frame in the sample slot from a RadioSet object.

**sampleNames<-:** assign new rownames to the sampleInfo data.frame for a RadioSet object. Arguments:

- **value:** character vector of rownames for the sampleInfo(object) data.frame.

**@curation:**

**curation:** A list of curated mappings between identifiers in the RadioSet object and the original data publication. Contains three data.frames, 'cell' with cell-line ids and 'tissue' with tissue ids and 'radiation' with radiation ids.

**curation<-:** Update the curation slot of a RadioSet object. Arguments:

- value: A list of data.frames, one for each type of curated identifier. For a RadioSet object the slot should contain tissue, cell-line and radiation id data.frames.

**datasetType slot:**

**datasetType:** character(1) The type treatment response in the sensitivity slot. Valid values are 'sensitivity', 'perturbation' or 'both'.

**datasetType<-:** Update the datasetType slot of a RadioSet object. Arguments:

- value: A character(1) vector with one of 'sensitivity', 'perturbation' or 'both'

**@molecularProfiles:**

**molecularProfiles:** matrix() Retrieve an assay in a SummarizedExperiment from the molecularProfiles slot of a RadioSet object with the specified mDataType. Valid mDataType arguments can be found with mDataNames(object). Exclude mDataType and assay to access the entire slot. Arguments:

- assay: Optional character(1) vector specifying an assay in the SummarizedExperiment of the molecularProfiles slot of the RadioSet object for the specified mDataType. If excluded, defaults to modifying the first assay in the SummarizedExperiment for the given mDataType.

**molecularProfiles<-:** Update an assay in a SummarizedExperiment from the molecularProfiles slot of a RadioSet object with the specified mDataType. Valid mDataType arguments can be found with mDataNames(object). Omit mDataType and assay to update the slot.

- assay: Optional character(1) vector specifying an assay in the SummarizedExperiment of the molecularProfiles slot of the RadioSet object for the specified mDataType. If excluded, defaults to modifying the first assay in the SummarizedExperiment for the given mDataType.
- value: A matrix of values to assign to the assay slot of the SummarizedExperiment for the selected mDataType. The rownames and column names must match the associated SummarizedExperiment.

**featureInfo:** Retrieve a DataFrame of feature metadata for the specified mDataType from the molecularProfiles slot of a RadioSet object. More specifically, retrieve the @rowData slot from the SummarizedExperiment from the @molecularProfiles of a RadioSet object with the name mDataType.

**\_\_featureInfo<-\_\_:** Update the 'featureInfo(object, mDataType)' 'DataFrame' with new feature metadata. Arguments: - value: A 'data.frame' or 'DataFrame' with updated feature metadata for the specified molecular profile in the 'molecularProfiles' slot of a 'RadioSet' object.

**phenoInfo:** Return the @colData slot from the SummarizedExperiment of mDataType, containing sample-level metadata, from a RadioSet object.

**phenoInfo<-:** Update the @colData slot of the SummarizedExperiment of mDataType in the @molecularProfiles slot of a RadioSet object. This updates the sample-level metadata in-place.

- value: A data.frame or DataFrame object where rows are samples and columns are sample metadata.

**fNames:** character() The features names from the rowData slot of a SummarizedExperiment of mDataType within a RadioSet object.

**fNames:** Updates the rownames of the feature metadata (i.e., rowData) for a SummarizedExperiment of mDataType within a RadioSet object.

- value: character() A character vector of new features names for the rowData of the SummarizedExperiment of mDataType in the @molecularProfiles slot of a RadioSet object. Must be the same length as nrow(featureInfo(object, mDataType)), the number of rows in the feature metadata.

**mDataNames:** character Retrieve the names of the molecular data types available in the molecularProfiles slot of a RadioSet object. These are the options which can be used in the mDataType parameter of various molecularProfiles slot accessors methods.

**mDataNames:** Update the molecular data type names of the molecularProfiles slot of a RadioSet object. Arguments:

- value: character vector of molecular datatype names, with length equal to length(molecularProfilesSlot(object

**molecularProfilesSlot:** Return the contents of the @molecularProfiles slot of a RadioSet object. This will either be a list or MultiAssayExperiment of SummarizedExperiments.

**molecularProfilesSlot<-:** Update the contents of the @molecularProfiles slot of a RadioSet object. Arguments:

- value: A list or MultiAssayExperiment of SummarizedExperiments. The list and assays should be named for the molecular datatype in each SummarizedExperiment.

#### **@treatmentResponse:**

*Arguments::*

- dimension: Optional character(1) One of 'treatment', 'sample' or 'assay' to retrieve rowData, colData or the 'assay\_metadata' assay from the RadioSet @sensitivity LongTable object, respectively. Ignored with warning if @treatmentResponse is not a LongTable object.
- ...: Additional arguments to the rowData or colData. LongTable methods. Only used if the sensitivity slot contains a LongTable object instead of a list and the dimension argument is specified.

*Methods::*

**sensitivityInfo:** DataFrame or data.frame of sensitivity treatment combo by sample metadata for the RadioSet object. When the dimension parameter is used, it allows retrieval of the dimension specific metadata from the LongTable object in @treatmentResponse of a RadioSet object.

**sensitivityInfo<-:** Update the @treatmentResponse slot metadata for a RadioSet object. When used without the dimension argument it behaves similar to the old RadioSet implementation, where the @treatmentResponse slot contained a list with a \$info data.frame item. When the dimension argument is used, more complicated assignments can occur where 'sample' modifies the @sensitivity LongTable colData, 'treatment' the rowData and 'assay' the 'assay\_metadata' assay. Arguments:

- value: A `data.frame` of treatment response experiment metadata, documenting experiment level metadata (mapping to treatments and samples). If the `@treatmentResponse` slot doesn't contain a `LongTable` and dimension is not specified, you can only modify existing columns as returned by `sensitivityInfo(object)`.

**sensitivityMeasures:** Get the 'sensitivityMeasures' available in a `RadioSet` object. Each measure represents some summary of sample sensitivity to a given treatment, such as `ic50`, `ec50`, `AUC`, `AAC`, etc. The results are returned as a character vector with all available metrics for the `PSet` object.

**sensitivityMeasures:** Update the sensitivity measure in a `RadioSet` object. These values are the column names of the 'profiles' assay and represent various computed sensitivity metrics such as `ic50`, `ec50`, `AUC`, `AAC`, etc.

- value: A character vector of new sensitivity measure names, the then length of the character vector must match the number of columns of the 'profiles' assay, excluding metadata and key columns.

**sensitivityProfiles:** Return the sensitivity profile summaries from the sensitivity slot. This `data.frame` contains various sensitivity summary metrics, such as `ic50`, `amax`, `EC50`, `aac`, `HS`, etc as columns, with rows as treatment by sample experiments.

**sensitivityProfiles<-:** Update the sensitivity profile summaries the sensitivity slot. Arguments: - value: A `data.frame` the the same number of rows as as returned by `sensitivityProfiles(object)`, but potentially modified columns, such as the computation of additional summary metrics.

**sensitivityRaw:** Access the raw sensitivity measurements for a `RadioSet` object. A 3D array where rows are `experiment_ids`, columns are doses and the third dimension is metric, either 'Dose' for the doses used or 'Viability' for the sample viability at that dose.

**sensitivityRaw<-:** Update the raw dose and viability data in a `RadioSet`.

- value: A 3D array object where rows are `experiment_ids`, columns are replicates and pages are `c('Dose', 'Viability')`, with the corresponding dose or viability measurement for that `experiment_id` and replicate.

**sensNumber:** Return a count of viability observations in a `RadioSet` object for each treatment-combo by sample combination.

**sensNumber<-:** Update the 'n' item, which holds a matrix with a count of treatment by sample-line experiment counts, in the `list` in `@treatmentResponse` slot of a `RadioSet` object. Will error when `@sensitivity` contains a `LongTable` object, since the counts are computed on the fly. Arguments:

- value: A matrix where rows are samples and columns are treatments, with a count of the number of experiments for each combination as the values.

**pertNumber:** array Summary of available perturbation experiments from in a `RadioSet` object. Returns a 3D array with the number of perturbation experiments per treatment and sample, and data type.

**pertNumber<-:** Update the `@perturbation$n` value in a `RadioSet` object, which stores a summary of the available perturbation experiments. Arguments:

- value: A new 3D array with the number of perturbation experiments per treatment and sample, and data type

**Value**

Accessors: See details.

Setters: An updated 'RadioSet' object, returned invisibly.

**Examples**

```
data(clevelandSmall)
treatmentInfo(clevelandSmall)

treatmentInfo(clevelandSmall) <- treatmentInfo(clevelandSmall)

treatmentNames(clevelandSmall)

treatmentNames(clevelandSmall) <- treatmentNames(clevelandSmall)

## @annotation
annotation(clevelandSmall)

annotation(clevelandSmall) <- annotation(clevelandSmall)

dateCreated(clevelandSmall)

## dateCreated
dateCreated(clevelandSmall) <- date()

name(clevelandSmall)

name(clevelandSmall) <- 'new_name'

sampleInfo(clevelandSmall) <- sampleInfo(clevelandSmall)

sampleNames(clevelandSmall)

sampleNames(clevelandSmall) <- sampleNames(clevelandSmall)

## curation
curation(clevelandSmall)

curation(clevelandSmall) <- curation(clevelandSmall)

datasetType(clevelandSmall)

datasetType(clevelandSmall) <- 'both'

# No assay specified
molecularProfiles(clevelandSmall, 'rna') <- molecularProfiles(clevelandSmall, 'rna')

# Specific assay
molecularProfiles(clevelandSmall, 'rna', 'exprs') <-
  molecularProfiles(clevelandSmall, 'rna', 'exprs')
```

```
# Replace the whole slot
molecularProfiles(clevelandSmall) <- molecularProfiles(clevelandSmall)

featureInfo(clevelandSmall, 'rna')

featureInfo(clevelandSmall, 'rna') <- featureInfo(clevelandSmall, 'rna')

phenoInfo(clevelandSmall, 'rna')

phenoInfo(clevelandSmall, 'rna') <- phenoInfo(clevelandSmall, 'rna')

fNames(clevelandSmall, 'rna')

fNames(clevelandSmall, 'rna') <- fNames(clevelandSmall, 'rna')

mDataNames(clevelandSmall)

mDataNames(clevelandSmall) <- mDataNames(clevelandSmall)

molecularProfilesSlot(clevelandSmall)

molecularProfilesSlot(clevelandSmall) <- molecularProfilesSlot(clevelandSmall)

sensitivityInfo(clevelandSmall)

sensitivityInfo(clevelandSmall) <- sensitivityInfo(clevelandSmall)

sensitivityMeasures(clevelandSmall) <- sensitivityMeasures(clevelandSmall)

sensitivityMeasures(clevelandSmall) <- sensitivityMeasures(clevelandSmall)

sensitivityProfiles(clevelandSmall)

sensitivityProfiles(clevelandSmall) <- sensitivityProfiles(clevelandSmall)

head(sensitivityRaw(clevelandSmall))

sensitivityRaw(clevelandSmall) <- sensitivityRaw(clevelandSmall)

treatmentResponse(clevelandSmall)

treatmentResponse(clevelandSmall) <- treatmentResponse(clevelandSmall)

sensNumber(clevelandSmall)

sensNumber(clevelandSmall) <- sensNumber(clevelandSmall)

pertNumber(clevelandSmall)

pertNumber(clevelandSmall) <- pertNumber(clevelandSmall)
```

---

RadioSet-class	<i>A Class to Contain RadioGenomic datasets together with their curations</i>
----------------	---

---

### Description

The RadioSet (RSet) class was developed to contain and organise large RadioGenomic datasets, and aid in their metanalysis. It was designed primarily to allow bioinformaticians and biologists to work with data at the level of genes and cell lines, providing a more naturally intuitive interface and simplifying analyses between several datasets. As such, it was designed to be flexible enough to hold datasets of two different natures while providing a common interface. The class can accommodate datasets containing both radiation dose response data, as well as datasets containing genetic profiles of cell lines pre and post treatment with compounds, known respectively as sensitivity and perturbation datasets.

### Value

An object of the RadioSet class

### Slots

`annotation` A list of annotation data about the RadioSet, including the `$name` and the session information for how the object was creating, detailing the exact versions of R and all the packages used

`molecularProfiles` A list containing 4 SummarizedExperiment type object for holding data for RNA, DNA, SNP and Copy Number Variation measurements respectively, with associated `fData` and `pData` containing the row and column metadata

`sample` A `data.frame` containing the annotations for all the cell lines profiled in the data set, across all data types

`treatment` A `data.frame` containing the annotations for all the radiation treatment types used in the in the dataset, across all data types

`sensitivity` A list containing all the data for the sensitivity experiments, including `$info`, a `data.frame` containing the experimental info, `$raw` a 3D array containing raw data, `$profiles`, a `data.frame` containing sensitivity profiles statistics, and `$n`, a `data.frame` detailing the number of experiments for each cell-radiation type pair

`perturbation` A list containing `$n`, a `data.frame` summarizing the available perturbation data,

`curation` A list containing mappings for cell and tissue names used in the data set to universal identifiers used between different RadioSet objects

`datasetType` A character string of 'sensitivity', 'perturbation', or both detailing what type of data can be found in the RadioSet, for proper processing of the data

RadioSig

*Radiation Signature Class Constructor***Description**

A documented constructor to provide user friendly interface to .RadioSig

**Usage**

```
RadioSig(
  Data = array(NA, dim = c(0, 0, 0)),
  RSetName = "",
  DateCreated = date(),
  SigType = "sensitivity",
  SessionInfo = sessionInfo(),
  Call = "No Call Recorded"
)
```

**Arguments**

Data	The data
RSetName	The name of the pSet
DateCreated	The date the object was created
SigType	The type of sensitivy signature
SessionInfo	The package version used to generate the object
Call	The calls for sensitivity vs not

**Value**

A RadioSig object

radSensitivitySig

*Creates a signature representing the association between gene expression (or other molecular profile) and radiation dose response, for use in radiation sensitivity analysis.*

**Description**

Given a RadioSet of the sensitivity experiment type, and a list of radiation types, the function will compute a signature for the effect of gene expression on the molecular profile of a cell. The function returns the estimated coefficient, the t-stat, the p-value and the false discovery rate associated with that coefficient, in a 3 dimensional array, with genes in the first direction, drugs in the second, and the selected return values in the third.

**Usage**

```
radSensitivitySig(
  rSet,
  mDataType,
  radiation.types,
  features,
  sensitivity.measure = "AUC_recomputed",
  molecular.summary.stat = c("mean", "median", "first", "last", "or", "and"),
  sensitivity.summary.stat = c("mean", "median", "first", "last"),
  returnValues = c("estimate", "pvalue", "fdr"),
  sensitivity.cutoff = NA,
  standardize = c("SD", "rescale", "none"),
  nthread = 1,
  verbose = TRUE,
  ...
)
```

**Arguments**

rSet	A RadioSet of the perturbation experiment type
mDataType	character which one of the molecular data types to use in the analysis, out of dna, rna, rnaseq, snp, cnv
radiation.types	character a vector of radiation.types for which to compute the signatures. Should match the names used in the PharmacoSet.
features	character a vector of features for which to compute the signatures. Should match the names used in correspondant molecular data in PharmacoSet.
sensitivity.measure	character which measure of the radiation sensitivity should the function use for its computations? Use the sensitivityMeasures function to find out what measures are available for each PSet.
molecular.summary.stat	What summary statistic should be used to summarize duplicates for cell line molecular profile measurements?
sensitivity.summary.stat	What summary statistic should be used to summarize duplicates for cell line sensitivity measurements?
returnValues	character Which of estimate, t-stat, p-value and fdr should the function return for each gene?
sensitivity.cutoff	Allows to provide upper and lower bounds to sensitivity measures in the cases where the values exceed physical values due to numerical or other errors.
standardize	character One of "SD", "rescale", or "none", for the form of standardization of the data to use. If "SD", the the data is scaled so that SD = 1. If rescale, then the data is scaled so that the 95 interquantile range lies in [0,1]. If none no rescaling is done.

nthread	numeric if multiple cores are available, how many cores should the computation be parallelized over?
verbose	boolean 'TRUE' if the warnings and other informative message should be displayed
...	additional arguments not currently fully supported by the function

**Value**

list a 3D array with genes in the first dimension, radiation.types in the second, and return values in the third.

**Examples**

```
data(clevelandSmall)
rad.sensitivity <- radSensitivitySig(clevelandSmall, mDataType="rna",
  nthread=1, features = fNames(clevelandSmall, "rna")[1],
  radiation.types=treatmentNames(clevelandSmall))
print(rad.sensitivity)
```

---

show, RadioSet-method    *Show a RadioSet*

---

**Description**

Show a RadioSet

**Usage**

```
## S4 method for signature 'RadioSet'
show(object)
```

**Arguments**

object            A RadioSet object

**Value**

Prints the RadioSet object to the output stream, and returns invisible NULL.

**Examples**

```
data(clevelandSmall)
clevelandSmall
```

---

show, RadioSig-method    *Show RadioGx Signatures*

---

### Description

Show RadioGx Signatures

### Usage

```
## S4 method for signature 'RadioSig'
show(object)
```

### Arguments

object                  RadioSig

### Value

Prints the RadioGx Signatures object to the output stream, and returns invisible NULL.

### Examples

```
data(clevelandSmall)
rad.sensitivity <- radSensitivitySig(clevelandSmall, mDataType="rna",
                                   nthread=1, features = fName(clevelandSmall, "rna")[1])
rad.sensitivity
```

---

showSigAnnot, RadioSig-method  
*Show the Annotations of a signature object*

---

### Description

This function prints out the information about the call used to compute the rad signatures, and the session info for the session in which the computation was done. Useful for determining the exact conditions used to generate signatures.

### Usage

```
## S4 method for signature 'RadioSig'
showSigAnnot(object)
```

### Arguments

object                  An object of the RadioSig Class, as returned by radPerturbationSig or radSensitivitySig

**Value**

Prints the RadioGx Signatures annotations to the output stream, and returns invisible NULL.

**Examples**

```
data(clevelandSmall)
rad.sensitivity <- radSensitivitySig(clevelandSmall, mDataType="rna",
  nthread=1, features = fName(clevelandSmall, "rna")[1])
showSigAnnot(rad.sensitivity)
```

---

subsetTo, RadioSet-method

*A function to subset a RadioSet to data containing only specified radiations, cells and genes*

---

**Description**

This is the preferred method of subsetting a RadioSet. This function allows abstraction of the data to the level of biologically relevant objects: radiations and cells. The function will automatically go through all of the combined data in the RadioSet and ensure only the requested radiations and cell lines are found in any of the slots. This allows quickly picking out all the experiments for a radiation or cell of interest, as well removes the need to keep track of all the metadata conventions between different datasets.

**Usage**

```
## S4 method for signature 'RadioSet'
subsetTo(
  object,
  cells = NULL,
  radiationTypes = NULL,
  molecular.data.cells = NULL,
  keep.controls = TRUE,
  ...
)
```

**Arguments**

object	A RadioSet to be subsetted
cells	A list or vector of cell names as used in the dataset to which the object will be subsetted. If left blank, then all cells will be left in the dataset.
radiationTypes	A list or vector of radiation names as used in the dataset to which the object will be subsetted. If left blank, then all radiationTypes will be left in the dataset.
molecular.data.cells	A list or vector of cell names to keep in the molecular data

keep.controls If the dataset has perturbation type experiments, should the controls be kept in the dataset? Defaults to true.

... Other arguments passed by other function within the package

### Value

A RadioSet with only the selected radiation types and cells

### Examples

```
clevelandRadiationTypes <- treatmentNames(clevelandSmall)
clevelandCells <- sampleNames(clevelandSmall)
RSet <- subsetTo(clevelandSmall, radiationTypes = clevelandRadiationTypes[1],
  cells = clevelandCells[1])
RSet
```

---

summarizeMolecularProfiles, RadioSet-method

*Takes molecular data from a RadioSet, and summarises them into one entry per drug*

---

### Description

Given a RadioSet with molecular data, this function will summarize the data into one profile per cell line, using the chosen summary.stat. Note that this does not really make sense with perturbation type data, and will combine experiments and controls when doing the summary if run on a perturbation dataset.

### Usage

```
## S4 method for signature 'RadioSet'
summarizeMolecularProfiles(
  object,
  mDataType,
  cell.lines,
  features,
  summary.stat = c("mean", "median", "first", "last", "and", "or"),
  fill.missing = TRUE,
  summarize = TRUE,
  verbose = TRUE
)
```

**Arguments**

object	RadioSet The RadioSet to summarize
mDataType	character which one of the molecular data types to use in the analysis, out of all the molecular data types available for the rSet for example: rna, rnaseq, snp
cell.lines	character The cell lines to be summarized. If any cell.line has no data, missing values will be created
features	character A vector of the feature names to include in the summary
summary.stat	character which summary method to use if there are repeated cell.lines? Choices are "mean", "median", "first", or "last" In case molecular data type is mutation or fusion "and" and "or" choices are available
fill.missing	boolean should the missing cell lines not in the molecular data object be filled in with missing values?
summarize	A flag which when set to FALSE (defaults to TRUE) disables summarizing and returns the data unchanged as a ExpressionSet
verbose	boolean should messages be printed

**Value**

matrix An updated RadioSet with the molecular data summarized per cell line.

**Examples**

```
data(clevelandSmall)
clevelandSmall <- summarizeMolecularProfiles(clevelandSmall,
      mDataType = "rna", cell.lines=sampleNames(clevelandSmall),
      summary.stat = 'median', fill.missing = TRUE, verbose=TRUE)
clevelandSmall
```

---

summarizeSensitivityProfiles, RadioSet-method

*Takes the sensitivity data from a RadioSet, and summarises them into a drug vs cell line table*

---

**Description**

This function creates a table with cell lines as rows and radiation types as columns, summarising the drug sensitivity data of a RadioSet into drug-cell line pairs

## Usage

```
## S4 method for signature 'RadioSet'
summarizeSensitivityProfiles(
  object,
  sensitivity.measure = "AUC_recomputed",
  cell.lines,
  radiation.types,
  summary.stat = c("mean", "median", "first", "last", "max", "min"),
  fill.missing = TRUE,
  verbose = TRUE
)
```

## Arguments

object	'RadioSet' The RadioSet from which to extract the data
sensitivity.measure	'character' which sensitivity measure to use? Use the sensitivityMeasures function to find out what measures are available for each PSet.
cell.lines	'character' The cell lines to be summarized. If any cell lines has no data, it will be filled with missing values
radiation.types	'character' The radiation types to be summarized. If any radiation type has no data, it will be filled with missing values
summary.stat	'character' which summary method to use if there are repeated cell line-drug experiments? Choices are "mean", "median", "first", or "last"
fill.missing	'logical(1)' should the missing cell lines not in the molecular data object be filled in with missing values?
verbose	Should the function print progress messages?

## Value

[matrix] A matrix with cell lines going down the rows, radiation types across the columns, with the selected sensitivity statistic for each pair.

## Examples

```
data(clevelandSmall)
GDSCauc <- summarizeSensitivityProfiles(clevelandSmall, sensitivity.measure='AUC_published')
```

---

updateObject, RadioSet-method

*Update the RadioSet class after changes in it structure or API*

---

**Description**

Update the RadioSet class after changes in it structure or API

**Usage**

```
## S4 method for signature 'RadioSet'  
updateObject(object)
```

**Arguments**

object            A RadioSet object to update the class structure for.

**Value**

RadioSet with update class structure.

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