Package: RMassBank (via r-universe)

August 22, 2026

Type Package

Title Workflow to process tandem MS files and build MassBank records

Version 3.23.0

Author Michael Stravs, Emma Schymanski, Steffen Neumann, Erik Mueller, Paul Stahlhofen, Tobias Schulze with contributions of Hendrik Treutler

Maintainer RMassBank at Eawag <massbank@eawag.ch>

Description Workflow to process tandem MS files and build MassBank records. Functions include automated extraction of tandem MS spectra, formula assignment to tandem MS fragments, recalibration of tandem MS spectra with assigned fragments, spectrum cleanup, automated retrieval of compound information from Internet databases, and export to MassBank records.

License Artistic-2.0

Depends Rcpp

Imports assertthat, Biobase, ChemmineR, data.table, digest, dplyr, enviPat, glue, httr, httr2, logger, methods, MSnbase, mzR, purrr, R.utils, rcdk, readJDX, readr, rjson, S4Vectors, tibble, tidyselect, webchem, XML, yaml

Suggests BiocStyle, CAMERA, gplots, knitr, magick, rmarkdown, RMassBankData (>= 1.33.1), RUnit, xcms (>= 1.37.1)

VignetteBuilder knitr

biocViews ImmunoOncology, Bioinformatics, MassSpectrometry, Metabolomics, Software

Encoding UTF-8

RoxygenNote 7.2.3

SystemRequirements OpenBabel

Collate 'alternateAnalyze.R' 'formulaCalculator.R' 'getSplash.R' 'leCsvAccess.R' 'settings_example.R' 'webAccess.R' 'deprofile.R' 'parseMassBank.R' 'Generics.R' 'SpectrumClasses.R' 'SpectrumMethods.R' 'RmbSpectrum2Update.R'

'readWriteMgfData.R' 'RmbWorkspace.R' 'RmbWorkspaceUpdate.R'
'SpectraSetMethods.R' 'AggregateMethods.R' 'leMsMs.r'
'leMsmsRaw.R' 'buildRecord.R' 'createMassBank.R'
'msmsRawExtensions.r' 'validateMassBank.R' 'tools.R'
'msmsRead.R' 'mergeSpectra.R' 'Isotopic_Annotation.R'
'fillback.R' 'parseMbRecord.R' 'zzz.R' 'log_wrapper.R'
'createCompoundlist.R'

Config/pak/sysreqs cmake libglpk-dev make default-jdk libicu-dev libpng-dev librsvg2-dev libuv1-dev libxml2-dev libnetcdf-dev libopenbabel-dev libssl-dev libx11-dev zlib1g-dev

Repository https://bioc.r-universe.dev

Date/Publication 2026-04-28 12:36:44 UTC

RemoteUrl https://github.com/bioc/RMassBank

RemoteRef HEAD

RemoteSha bca43b51ad069b7308671399c264ff99f692446a

Contents

-,RmbSpectraSet,ANY-method . . . 5
-,RmbSpectrum2List,ANY-method . . . 5
-,Spectrum,numeric-method . . . 6
.parseTitleString . . . 6
.updateObject.RmbSpectrum2.formulaSource . . . 7
+,RmbSpectraSet,ANY-method . . . 8
+,RmbSpectrum2List,ANY-method . . . 8
+,Spectrum,numeric-method . . . 9
add.formula . . . 9
addMB . . . 10
addPeaks . . . 11
addPeaksManually . . . 12
addProperty . . . 13
aggregateSpectra . . . 14
analyzeMsMs . . . 15
annotator.default . . . 18
archiveResults . . . 19
buildRecord . . . 19
CAS2SMILES . . . 21
checkIsotopes . . . 21
checkSpectra . . . 23
cleanElnoise . . . 24
combineMultiplicities . . . 25
compoundlist2SDF . . . 26
createCompoundlist . . . 27
createMolfile . . . 28
CTS.externalIdSubset . . . 29
CTS.externalIdTypes . . . 30
dbe . . . 30

deprofile . . . 31
exportMassbank . . . 33
fillback . . . 34
filterCompoundlist . . . 35
filterLowaccResults . . . 36
filterMultiplicity . . . 37
filterPeakSatellites . . . 38
filterPeaksMultiplicity . . . 39
findEIC . . . 40
findMass . . . 42
findMsMsHR . . . 43
findMsMsHR.direct . . . 46
findMsMsHR.ticms2 . . . 47
findMsMsHRperMsp . . . 48
findMsMsHRperxcms . . . 49
findMz . . . 50
findMz.formula . . . 52
findProgress . . . 53
flatten . . . 53
formulastring.to.list . . . 55
gatherCCTE . . . 56
gatherData . . . 57
gatherDataBabel . . . 58
gatherDataUnknown . . . 59
gatherPubChem . . . 60
getAnalyticalInfo . . . 61
getCactus . . . 61
getCASRN . . . 62
getCSID . . . 63
getCtsKey . . . 64
getCtsRecord . . . 65
getData . . . 66
getDTXCID . . . 66
getDTXSID . . . 67
getDTXSMILES . . . 68
getField . . . 69
getMolecule . . . 70
getPcId . . . 71
getPrefName . . . 72
is.valid.formula . . . 73
loadInfolists . . . 74
loadList . . . 75
makeMollist . . . 76
makePeaksCache . . . 76
makeRecalibration . . . 77
mbWorkflow . . . 79
mbWorkspace-class . . . 80
mergePeaks . . . 81

mergeSpectra . . . 82
msmsRead . . . 83
msmsRead.RAW . . . 85
msmsWorkflow . . . 86
msmsWorkspace-class . . . 88
newMbWorkspace . . . 89
newMsmsWorkspace . . . 89
normalize,RmbSpectrum2-method . . . 90
normalize,RmbSpectrum2List-method . . . 91
order.formula . . . 91
parseMassBank . . . 92
parseMbRecord . . . 93
peaksMatched . . . 94
peaksUnmatched . . . 94
plotMbWorkspaces . . . 95
plotRecalibration . . . 96
ppm . . . 97
problematicPeaks . . . 98
processProblematicPeaks . . . 99
progressBarHook . . . 99
property . . . 100
property<- . . . 101
reanalyzeFailpeaks . . . 102
recalibrate . . . 103
recalibrate.addMS1data . . . 105
rmb_log_debug . . . 106
rmb_log_error . . . 106
rmb_log_fatal . . . 107
rmb_log_info . . . 107
rmb_log_success . . . 108
rmb_log_trace . . . 108
rmb_log_warn . . . 109
RmbDefaultSettings . . . 109
RmbSettings . . . 110
RmbSpectraSet-class . . . 113
RmbSpectrum2-class . . . 114
RmbSpectrum2List-class . . . 115
selectPeaks . . . 115
selectSpectra . . . 116
setAccessionBuilder . . . 117
setData . . . 118
smiles2mass . . . 118
spectraCount . . . 119
to.limits.rcdk . . . 120
toMassbank . . . 121
toRMB . . . 123
updateHeader . . . 124
updateSettings . . . 124

validate 125

Index **127**

`-,RmbSpectraSet,ANY-method`

Add a negative mass shift to a list of spectra

Description

Shifts both ‘parent‘ and ‘children‘ spectra of the ‘RmbSpectraSet‘ by the same mass.

Usage

```
## S4 method for signature 'RmbSpectraSet,ANY'
e1 - e2
```

Arguments

`e1`	a ‘RmbSpectraSet‘ object containing zero or more ‘children‘ spectra and a ‘parent‘ spectrum
`e2`	a numeric mass shift

`-,RmbSpectrum2List,ANY-method`

Add a negative mass shift to a list of spectra

Description

Shifts all spectra in a ‘RmbSpectrum2List‘ by the same mass

Usage

```
## S4 method for signature 'RmbSpectrum2List,ANY'
e1 - e2
```

Arguments

`e1`	a ‘RmbSpectrum2List‘ object containing zero or more ‘RmbSpectrum2‘ spectra
`e2`	a numeric mass shift

`-,Spectrum,numeric-method` *Add a negative mass shift to a spectrum*

Description

Add a negative mass shift to a spectrum

Usage

```
## S4 method for signature 'Spectrum,numeric'
e1 - e2
```

Arguments

`e1`	a ‘MSnbase::Spectrum‘ object
`e2`	a numeric mass shift

`.parseTitleString` *Parse record title*

Description

Parses a title for a single MassBank record using the title format specified in the option titleFormat. Internally used, not exported.

Usage

```
.parseTitleString(mbdata)
```

Arguments

`mbdata`	list The information data block for the record header, as stored in `mbdata_relisted` after loading an infolist.

Details

If the option is not set, a standard title format is used (for record definition version 1 or 2).

Value

A string with the title.

Author(s)

Michael Stravs, Eawag

References

MassBank record format: http://www.massbank.jp/manuals/MassBankRecord_en.pdf

See Also

buildRecord

Examples

```
## Not run:
  # used in buildRecord()
  title <- .parseTitleString(mbdata)

## End(Not run)
```

`.updateObject.RmbSpectrum2.formulaSource` *Add formulaSource column to spectrum.*

Description

TODO: consider whether to add functionality to move reanalysis stuff from legacy data back in.

Usage

```
.updateObject.RmbSpectrum2.formulaSource(w)
```

Arguments

w	RmbSpectrum2 The object to be updated

Value

The updated RmbSpectrum2 object

Author(s)

stravsmi

`+,RmbSpectraSet,ANY-method`
Add a mass shift to a list of spectra

Description

Shifts both ‘parent‘ and ‘children‘ spectra of the ‘RmbSpectraSet‘ by the same mass.

Usage

```
## S4 method for signature 'RmbSpectraSet,ANY'
e1 + e2
```

Arguments

e1	a ‘RmbSpectraSet‘ object containing zero or more ‘children‘ spectra and a ‘parent‘ spectrum
e2	a numeric mass shift

`+,RmbSpectrum2List,ANY-method`
Add a mass shift to a list of spectra

Description

Shifts all spectra in a ‘RmbSpectrum2List‘ by the same mass

Usage

```
## S4 method for signature 'RmbSpectrum2List,ANY'
e1 + e2
```

Arguments

e1	a ‘RmbSpectrum2List‘ object containing zero or more ‘RmbSpectrum2‘ spectra
e2	a numeric mass shift

+,Spectrum,numeric-method *Add a mass shift to a spectrum*

Description

Add a mass shift to a spectrum

Usage

```
## S4 method for signature 'Spectrum,numeric'
e1 + e2
```

Arguments

e1	a 'MSnbase::Spectrum' object
e2	a numeric mass shift

add.formula *Calculations on molecular formulas*

Description

Add, subtract, and multiply molecular formulas.

Usage

```
add.formula(f1, f2, as.formula = TRUE, as.list = FALSE)
multiply.formula(f1, n, as.formula = TRUE, as.list = FALSE)
```

Arguments

f1, f2	Molecular formulas (in list form or in text form) to calculate with.
as.formula	Return the result as a text formula (e.g. "C6H12O6"). This is the default
as.list	Return the result in list format (e.g. list(C=6, H=12,O=6)).
n	Multiplier (positive or negative, integer or non-integer.)

Details

Note that the results are not checked for plausibility at any stage, nor reordered.

Value

The resulting formula, as specified above.

Author(s)

Michael Stravs

See Also

formulastring.to.list, is.valid.formula, order.formula

Examples

```
##

add.formula("C6H12O6", "C3H3")
add.formula("C6H12O6", "C-3H-3")
add.formula("C6H12O6", multiply.formula("C3H3", -1))
```

---

addMB *MassBank-record Addition*

Description

Adds the peaklist of a MassBank-Record to the specs of an msmsWorkspace

Usage

```
addMB(w, cpdID, fileName, mode)
```

Arguments

w	The msmsWorkspace that the peaklist should be added to.
cpdID	The compoundID of the compound that has been used for the record
fileName	The path to the record
mode	The ionization mode that has been used to create the record

Value

The msmsWorkspace with the additional peaklist from the record

Author(s)

Erik Mueller

See Also

addPeaksManually

Examples

```
## Not run:
  addMB("filepath_to_records/RC00001.txt")

## End(Not run)
```

---

addPeaks *Add additional peaks to spectra*

Description

Loads a table with additional peaks to add to the MassBank spectra. Required columns are `cpdID`, `scan`, `int`, `mzFound`, `OK`.

Usage

```
addPeaks(mb, filename_or_dataframe)
```

Arguments

`mb`	The `mbWorkspace` to load the peaks into.
`filename_or_dataframe`	Filename of the csv file, or name of the R dataframe containing the peaklist.

Details

All peaks with OK=1 will be included in the spectra.

Value

The `mbWorkspace` with loaded additional peaks.

Author(s)

Michael Stravs

See Also

`mbWorkflow`

Examples

```
## Not run: addPeaks("myrun_additionalPeaks.csv")
```

addPeaksManually *Addition of manual peaklists*

Description

Adds a manual peaklist in matrix-format

Usage

```
addPeaksManually(w, cpdID, handSpec, mode)
```

Arguments

w	The msmsWorkspace that the peaklist should be added to.
cpdID	The compoundID of the compound that has been used for the peaklist
handSpec	A peaklist with 2 columns, one with "mz", one with "int"
mode	The ionization mode that has been used for the spectrum represented by the peaklist

Value

The msmsWorkspace with the additional peaklist added to the right spectrum

Author(s)

Erik Mueller

See Also

msmsWorkflow

Examples

```
## Not run:
 handSpec <- cbind(mz=c(274.986685367956, 259.012401087427, 95.9493025990907, 96.9573002472772),
                                int=c(357,761, 2821, 3446))
  addPeaksManually(w, cpdID, handSpec)

## End(Not run)
```

addProperty *Add and initialize dataframe column*

Description

Adds a new column of a defined type to a `data.frame` and initializes it to a value. The advantage of doing this over adding it with `$` or `[,""]` is that the case `nrow(o) == 0` is adequately handled and doesn't raise an error.

Usage

```
addProperty(o, name, type, value = NA)

## S4 method for signature 'RmbSpectrum2,character,character'
addProperty(o, name, type, value = NA)

## S4 method for signature 'data.frame,character,character'
addProperty(o, name, type, value = NA)
```

Arguments

`o`	`data.frame` to add the column to
`name`	Name of the new column
`type`	Data type of the new column
`value`	Initial value of the new column (`NA` if not given)

Value

Expanded data frame.

Methods (by class)

- `addProperty(o = data.frame, name = character, type = character)`: Add a new column to a data.frame

Author(s)

stravsmi

`aggregateSpectra` *Aggregate analyzed spectra*

Description

Groups an array of analyzed spectra and creates aggregated peak tables

Usage

```
aggregateSpectra(spec,  addIncomplete=FALSE)
```

Arguments

`spec`	The `RmbSpectraSetList` of spectra sets (`RmbSpectraSet` objects) to aggregate
`addIncomplete`	Whether or not the peaks from incomplete files (files for which less than the maximal number of spectra are present)

Details

addIncomplete is relevant for recalibration. For recalibration, we want to use only high-confidence peaks, therefore we set *addIncomplete* to `FALSE`. When we want to generate a peak list for actually generating MassBank records, we want to include all peaks into the peak tables.

Value

A summary `data.frame` with all peaks (possibly multiple rows for one m/z value from a spectrum, see below) with columns:

`mzFound`, `intensity`
: Mass and intensity of the peak

`good`
: if the peak passes filter criteria

`mzCalc`, `formula`, `dbe`, `dppm`
: calculated mass, formula, dbe and ppm deviation of the assigned formula

`formulaCount`, `dppmBest`
: Number of matched formulae for this m/z value, and ppm deviation of the best match

`scan`, `cpdID`, `parentScan`
: Scan number of the child and parent spectrum in the raw file, also the compound ID to which the peak belongs

`dppmRc`
: ppm deviation recalculated from the aggregation function

`index`
: Aggregate-table peak index, so the table can be subsetted, edited and results reinserted back into this table easily

Further columns are later added by workflow steps 6 (electronic noise culler), 7 and 8.

Author(s)

Michael Stravs

See Also

msmsWorkflow, analyzeMsMs

Examples

```
## As used in the workflow:
## Not run: %
 w@spectra <- lapply(w@spectra, function(spec)
  analyzeMsMs(spec, mode="pH", detail=TRUE, run="recalibrated", cut=0, cut_ratio=0 ) )
 w@aggregate <- aggregateSpectra(w@spectra)

## End(Not run)
```

---

analyzeMsMs *Analyze MSMS spectra*

Description

Analyzes MSMS spectra of a compound by fitting formulas to each subpeak.

Usage

```
analyzeMsMs(
  msmsPeaks,
  mode = "pH",
  detail = FALSE,
  run = "preliminary",
  filterSettings = getOption("RMassBank")$filterSettings,
  spectraList = getOption("RMassBank")$spectraList,
  method = "formula"
)

analyzeMsMs.formula(
  msmsPeaks,
  mode = "pH",
  detail = FALSE,
  run = "preliminary",
  filterSettings = getOption("RMassBank")$filterSettings
)

analyzeMsMs.intensity(
  msmsPeaks,
```

```
  mode = "pH",
  detail = FALSE,
  run = "preliminary",
  filterSettings = getOption("RMassBank")$filterSettings
)
```

Arguments

msmsPeaks	A RmbSpectraSet object. Corresponds to a parent spectrum and children MSMS spectra of one compound (plus some metadata). The objects are typically generated with findMsMsHR, and populate the @spectrum slot in a msmsWorkspace (refer to the corresponding documentation for the precise format specifications).
mode	Specifies the processing mode, i.e. which molecule species the spectra contain. *pH* (positive H) specifies [M+H]+, *pNa* specifies [M+Na]+, *pM* specifies [M]+, *mH* and *mNa* specify [M-H]- and [M-Na]-, respectively. (I apologize for the naming of *pH* which has absolutely nothing to do with chemical *pH* values.)
detail	Whether detailed return information should be provided (defaults to FALSE). See below.
run	"preliminary" or "recalibrated". In the preliminary run, mass tolerance is set to 10 ppm (above m/z 120) and 15 ppm (below m/z 120), the default intensity cutoff is 10^4 for positive mode (no default cutoff in negative mode), and the column "mz" from the spectra is used as data source. In the recalibrated run, the mass tolerance is set to 5 ppm over the whole mass range, the default cutoff is 0 and the column "mzRecal" is used as source for the m/z values. Defaults to "preliminary".
filterSettings	Settings for the filter parameters, by default loaded from the RMassBank settings set with e.g. loadRmbSettings. Must contain: • ppmHighMass, allowed ppm deviation before recalibration for high mass range • ppmLowMass, allowed ppm deviation before recalibration for low mass range • massRangeDivision, division point between high and low mass range (before recalibration) • ppmFine, allowed ppm deviation overall after recalibration • prelimCut, intensity cutoff for peaks in preliminary run • prelimCutRatio, relative intensity cutoff for peaks in preliminary run, e.g. 0.01 = 1 • fineCut, intensity cutoff for peaks in second run • fineCutRatio, relative intensity cutoff for peaks in second run • specOkLimit, minimum intensity of base peak for spectrum to be accepted for processing • dbeMinLimit, minimum double bond equivalent for accepted molecular subformula. • satelliteMzLimit, for satellite peak filtering (filterPeakSatellites: mass window to use for satellite removal • satelliteIntLimit, the relative intensity below which to discard "satellites". (refer to filterPeakSatellites).

`spectraList` The list of MS/MS spectra present in each data block. As also defined in the settings file.

`method` Selects which function to actually use for data evaluation. The default "formula" runs a full analysis via formula assignment to fragment peaks. The alternative setting "intensity" calls a "mock" implementation which circumvents formula assignment and filters peaks purely based on intensity cutoffs and the satellite filtering. (In this case, the ppm and dbe related settings in filterSettings are ignored.)

Details

The analysis function uses Rcdk. Note that in this step, *satellite peaks* are removed by a simple heuristic rule (refer to the documentation of `filterPeakSatellites` for details.)

Value

The processed `RmbSpectraSet` object. Added (or filled, respectively, since the slots are present before) data include

`list("complete")`
whether all spectra have useful value

`list("empty")` whether there are no useful spectra

`list("children")`
The processed `RmbSpectrum2` objects (in a `RmbSpectrum2List`).

- `ok` if the spectrum was successfully processed with at least one resulting peak
- `mz`, `intensity`: note that mz/int pairs can be duplicated when multiple matches are found for one mz value, therefore the two slots are not necessarily unchanged from before
- `rawOK` (logical) whether the m/z peak passes satellite/low removal
- `low`, `satellite` if `TRUE`, the peak failed cutoff (`low`) or was removed as `satellite`
- `formula`, `mzCalc`, `dppm`, `dbe` Formula, calculated mass, ppm deviation and dbe assigned to a peak
- `formulaCount`, `dppmBest` Number of formulae matched for this m/z value and ppm deviation of the best match
- `info` Spectrum identifying information (collision energy, resolution, collision mode) from the `spectraList`
- All other entries are retained from the original `RmbSpectrum2`.

Functions

- `analyzeMsMs.formula()`: Analyze the peaks using formula annotation
- `analyzeMsMs.intensity()`: Analyze the peaks going only by intensity values

Author(s)

Michael Stravs

See Also

msmsWorkflow, filterLowaccResults, filterPeakSatellites, reanalyzeFailpeaks

Examples

```
## Not run: analyzed <- analyzeMsMs(spec, "pH", TRUE)
```

---

annotator.default *Generate peak annotation from peaklist*

Description

Generates the PK$ANNOTATION entry from the peaklist obtained. This function is overridable by using the "annotator" option in the settings file.

Usage

```
annotator.default(annotation, formulaTag)
```

Arguments

annotation	A peak list to be annotated. Contains columns: "cpdID","formula","mzFound" ,"scan","mzCalc","dppm", "dbe","mz","int","formulaCount","parentScan","fM_factor","d "formulaMultiplicity","intrel","mzSpec"
formulaTag	The ion type to be added to annotated formulas ("+" or "-" usually)

Value

The annotated peak table. Table colnames() will be used for the titles (preferrably don't use spaces in the column titles; however no format is strictly enforced by the MassBank data format.

Author(s)

Michele Stravs, Eawag <stravsmi@eawag.ch>

Examples

```
## Not run:
annotation <- annotator.default(annotation)

## End(Not run)
```

archiveResults *Backup* msmsWorkflow *results*

Description

Writes the results from different msmsWorkflow steps to a file.

Usage

```
archiveResults(w, fileName, settings = getOption("RMassBank"))
```

Arguments

w	The msmsWorkspace to be saved.
fileName	The filename to store the results under.
settings	The settings to be stored into the msmsWorkspace image.

Examples

```
# This doesn't really make a lot of sense,
# it stores an empty workspace.
RmbDefaultSettings()
w <- newMsmsWorkspace()
archiveResults(w, "narcotics.RData")
```

buildRecord *Build MassBank records*

Description

Takes a spectra block for a compound, as returned from analyzeMsMs, and an aggregated cleaned peak table, together with a MassBank information block, as stored in the infolists and loaded via loadInfolist/readMbdata and processes them to a MassBank record

Usage

```
buildRecord(o, ..., cpd, mbdata, analyticalInfo, additionalPeaks)

## S4 method for signature 'RmbSpectraSet'
buildRecord(o, ..., cpd, mbdata, analyticalInfo, additionalPeaks)

## S4 method for signature 'RmbSpectrum2'
buildRecord(
  o,
  ...,
```

```
  cpd = NULL,
  mbdata = list(),
  analyticalInfo = list(),
  additionalPeaks = NULL
)
```

Arguments

o	RmbSpectraSet or RmbSpectrum2 The spectra (or single spectrum) should be taken from a compound after analysis (analyzeMsMs). Note that **peaks are not read from this object anymore**: Peaks come from the aggregated dataframe (and from the global additionalPeaks dataframe; cf. addPeaks for usage information.)
...	keyword arguments for intensity normalization and peak selection (see normalize and selectPeaks)
cpd	RmbSpectraSet or missing In case o is an RmbSpectrum2, this represents the RmbSpectraSet it belongs to
mbdata	list The information data block for the record header, as stored in mbdata_relisted after loading an infolist.
analyticalInfo	A list containing information for the 'AC$' section of a MassBank record, with elements 'ai, ac_lc, ac_ms' for general, LC and MS info respectively.
additionalPeaks	data.frame If present, a table with additional peaks to add into the spectra. As loaded with addPeaks.

Value

An object of the same type as was used for the input with new information added to it

Author(s)

Michael Stravs

References

MassBank record format: http://www.massbank.jp/manuals/MassBankRecord_en.pdf

See Also

mbWorkflow, addPeaks, toMassbank

CAS2SMILES *Convert CAS to SMILES*

Description

This is a wrapper for `webchem::cir_query`, using the CACTUS API at https://cactus.nci.nih.gov/chemical/structure_docume for the conversion. Before converting the CAS number, the name is checked whether it contains the word 'derivative'. If so, the conversion is stopped and NA is returned. Also, a warning will be printed in this case.

Usage

```
CAS2SMILES(CAS_number, name)
```

Arguments

`CAS_number`	character The CAS registry number of a compound
`name`	character The compound's name

Details

The API allows only one query per second. This is a hard- coded feature

Value

The SMILES code of the compound as character-string

Author(s)

pstahlhofen

Examples

```
SMILES_ethanol <- CAS2SMILES("64-17-5", "Ethanol")
```

checkIsotopes *Checks for isotopes in a* `msmsWorkspace`

Description

Checks for isotopes in a `msmsWorkspace`

Usage

```
checkIsotopes(
  w,
  mode = "pH",
  intensity_cutoff = 0,
  intensity_precision = "none",
  conflict = "strict",
  isolationWindow = 2,
  evalMode = "complete",
  plotSpectrum = TRUE,
  settings = getOption("RMassBank")
)
```

Arguments

w	A msmsWorkspace to work with.
mode	"pH", "pNa", "pM", "pNH4", "mH", "mM", "mFA" for different ions ([M+H]+, [M+Na]+, [M]+, [M+NH4]+, [M-H]-, [M]-, [M+FA]-).
intensity_cutoff	The cutoff (as an absolute intensity value) under which isotopic peaks shouldn't be checked for or accepted as valid. Please note: The cutoff is not hard in the sense that it interacts with the intensity_precision parameter.
intensity_precision	The difference that is accepted between the calculated and observed intensity of a possible isotopic peak. Further details down below.
conflict	Either "isotopic"(Peak formulas are always chosen if they fit the requirements for an isotopic peak) or "strict"(Peaks are only marked as isotopic when there hasn't been a formula assigned before.)
isolationWindow	Half of the width of the isolation window in Da
evalMode	Currently no function yet, but planned. Currently must be "complete"
plotSpectrum	A boolean specifiying whether the spectrumshould be plotted
settings	Options to be used for processing. Defaults to the options loaded via loadRmbSettings et al. Refer to there for specific settings.

Details

text describing parameter inputs in more detail.

- intensity_precisionThis parameter determines how strict the intensity values should adhere to the calculated intensity in relation to the parent peak. Options for this parameter are "none", where the intensity is irrelevant, "low", which has an error margin of 70% and "high", where the error margin is set to 35%. The recommended setting is "low", but can be changed to adjust to the intensity precision of the mass spectrometer.

Value

The msmsWorkspace with annotated isolation peaks

Author(s)

Michael Stravs, Eawag <michael.stravs@eawag.ch>

Erik Mueller, UFZ

`checkSpectra` *Check if a spectra set is found, complete, empty*

Description

Checks if a specific compound (`RmbSpectraSet`) was found with child spectra in the raw file (`found`), has a complete set of MS2 spectra with useful peaks (`complete`), or is empty (note: spectra are currently not ever marked empty - empty should mean found, but no useful peaks at all. This is not yet currently tested.)

Usage

```
checkSpectra(s, property)

## S4 method for signature 'RmbSpectraSet,character'
checkSpectra(s, property)
```

Arguments

`s`	The (`RmbSpectraSet`) to check
`property`	The property to check (`found`, `complete` or `empty`)

Value

`TRUE` or `FALSE`

Methods (by class)

- `checkSpectra(s = RmbSpectraSet, property = character)`:

Author(s)

stravsmi

`cleanElnoise` *Remove electronic noise*

Description

Removes known electronic noise peaks from a peak table

Usage

```
cleanElnoise(peaks, noise=getOption("RMassBank")$electronicNoise,
  width = getOption("RMassBank")$electronicNoiseWidth)

## S4 method for signature 'data.frame,numeric,numeric'
cleanElnoise(
  peaks,
  noise = getOption("RMassBank")$electronicNoise,
  width = getOption("RMassBank")$electronicNoiseWidth
)

## S4 method for signature 'RmbSpectrum2,numeric,numeric'
cleanElnoise(
  peaks,
  noise = getOption("RMassBank")$electronicNoise,
  width = getOption("RMassBank")$electronicNoiseWidth
)

## S4 method for signature 'RmbSpectrum2List,numeric,numeric'
cleanElnoise(
  peaks,
  noise = getOption("RMassBank")$electronicNoise,
  width = getOption("RMassBank")$electronicNoiseWidth
)

## S4 method for signature 'RmbSpectraSet,numeric,numeric'
cleanElnoise(
  peaks,
  noise = getOption("RMassBank")$electronicNoise,
  width = getOption("RMassBank")$electronicNoiseWidth
)
```

Arguments

`peaks`	An aggregated peak frame as described in `aggregateSpectra`. Columns `mzFound`, `dppm` and `dppmBest` are needed.
`noise`	A numeric vector of known m/z of electronic noise peaks from the instrument Defaults to the entries in the RMassBank settings.

`width` The window for the noise peak in m/z units. Defaults to the entries in the RMassBank settings.

Value

Extends the aggregate data frame by column `noise` (logical), which is `TRUE` if the peak is marked as noise.

Methods (by class)

- `cleanElnoise(peaks = data.frame, noise = numeric, width = numeric)`: Remove known electronic noise peaks
- `cleanElnoise(peaks = RmbSpectrum2, noise = numeric, width = numeric)`: Remove known electronic noise peaks
- `cleanElnoise(peaks = RmbSpectrum2List, noise = numeric, width = numeric)`: Remove known electronic noise peaks
- `cleanElnoise(peaks = RmbSpectraSet, noise = numeric, width = numeric)`: Remove known electronic noise peaks

Author(s)

Michael Stravs

See Also

`msmsWorkflow`

Examples

```
# As used in the workflow:
## Not run:
     w@aggregated <-
  cleanElnoise(w@aggregated)

## End(Not run)
```

---

combineMultiplicities *Combine workspaces for multiplicity filtering*

Description

Combines multiple msmsWorkspace items to one workspace which is used for multiplicity filtering.

Usage

```
combineMultiplicities(workspaces)
```

Arguments

workspaces	A vector of msmsWorkspace items. The first item is taken as the "authoritative" workspace, i.e. the one which will be used for the record generation. The subsequent workspaces will only be used for multiplicity filtering.

Details

This feature is particularily meant to be used in conjunction with the confirmMode option of msmsWorkflow: a file can be analyzed with confirmMode = 0 (default) and subsequently with confirmMode = 1 (take second highest scan). The second analysis should contain "the same" spectra as the first one (but less intense) and can be used to confirm the peaks in the first spectra.

TO DO: Enable the combination of workspaces for combining e.g. multiple energy settings measured separately.

Value

A msmsWorkspace object prepared for step 8 processing.

Author(s)

Stravs MA, Eawag <michael.stravs@eawag.ch>

See Also

msmsWorkspace-class

Examples

```
## Not run:
 w <- newMsmsWorkspace
 w@files <- c("spec1", "spec2")
 w1 <- msmsWorkflow(w, steps=c(1:7), mode="pH")
 w2 <- msmsWorkflow(w, steps=c(1:7), mode="pH", confirmMode = 1)
 wTotal <- combineMultiplicities(c(w1, w2))
 wTotal <- msmsWorkflow(wTotal, steps=8, mode="pH", archivename = "output")
 # continue here with mbWorkflow

## End(Not run)
```

---

compoundlist2SDF *Convert a Compoundlist into an SDF*

Description

The resulting SDF will be written to a file named 'Compoundlist.sdf'. The header for each block is the chemical name, tags for ID, SMILES and CAS are added in the description block

Usage

```
compoundlist2SDF(filename)
```

Arguments

filename	character The name of the csv-file to be read. Note that the compoundlist has to be filtered already.

Value

This method has no return value.

Author(s)

pstahlhofen

Examples

```
## Not run:
   compoundlist2SDF("Compoundlist_filtered.csv")

## End(Not run)
```

---

createCompoundlist *Create a Compoundlist from JCAMP files*

Description

This method will automatically look for all single-block JCAMP files in the directory by picking all files ending in '.dx' (and not '.jdx'). A csv-file named 'Compoundlist.csv' will be created in the same directory. The Compoundlist contains columns 'ID', 'Name', 'SMILES' and 'CAS' where 'SMILES' might be empty if the compound is a derivative or if the CAS number could not be converted (see CAS2SMILES).

Usage

```
createCompoundlist()
```

Value

This method has no return value.

Author(s)

pstahlhofen

See Also

CAS2SMILES

Examples

```
## Not run:
   # Prepare the compoundlist-creation
   splitMultiblockDX('my_multiblock_jcamp.jdx')
   createCompoundlist()

## End(Not run)
```

`createMolfile` *Create MOL file for a chemical structure*

Description

Creates a MOL file (in memory or on disk) for a compound specified by the compound ID or by a SMILES code.

Usage

```
createMolfile(id_or_smiles, fileName = FALSE)
```

Arguments

`id_or_smiles`	The compound ID or a SMILES code.
`fileName`	If the filename is set, the file is written directly to disk using the specified filename. Otherwise, it is returned as a text array.

Details

The function invokes OpenBabel (and therefore needs a correctly set OpenBabel path in the RMassBank settings), using the SMILES code retrieved with `findSmiles` or using the SMILES code directly. The current implementation of the workflow uses the latter version, reading the SMILES code directly from the MassBank record itself.

Value

A character array containing the MOL/SDF format file, ready to be written to disk.

Author(s)

Michael Stravs

References

OpenBabel: http://openbabel.org

See Also

`findSmiles`

Examples

```
# Benzene:
## Not run:
createMolfile("C1=CC=CC=C1")

## End(Not run)
```

`CTS.externalIdSubset` *Select a subset of external IDs from a CTS record.*

Description

Select a subset of external IDs from a CTS record.

Usage

```
CTS.externalIdSubset(data, database)
```

Arguments

`data`	The complete CTS record as retrieved by `getCtsRecord`.
`database`	The database for which keys should be returned.

Value

Returns an array of all external identifiers stored in the record for the given database.

Author(s)

Michele Stravs, Eawag <stravsmi@eawag.ch>

Examples

```
## Not run:
# Return all CAS registry numbers stored for benzene.
data <- getCtsRecord("UHOVQNZJYSORNB-UHFFFAOYSA-N")
cas <- CTS.externalIdSubset(data, "CAS")

## End(Not run)
```

CTS.externalIdTypes *Find all available databases for a CTS record*

Description

Find all available databases for a CTS record

Usage

```
CTS.externalIdTypes(data)
```

Arguments

data	The complete CTS record as retrieved by getCtsRecord.

Value

Returns an array of all database names for which there are external identifiers stored in the record.

Author(s)

Michele Stravs, Eawag <stravsmi@eawag.ch>

Examples

```
## Not run:
# Return all databases for which the benzene entry has
# links in the CTS record.

data <- getCTS("UHOVQNZJYSORNB-UHFFFAOYSA-N")
databases <- CTS.externalIdTypes(data)

## End(Not run)
```

dbe *Calculate Double Bond Equivalents*

Description

Calculates the Ring and Double Bond Equivalents for a chemical formula. The highest valence state of each atom is used, such that the returned DBE should never be below 0.

Usage

```
dbe(formula)
```

Arguments

formula	A molecular formula in text or list representation (e.g. "C6H12O6" or list(C=6, H=12, O=6)).

Value

Returns the DBE for the given formula.

Author(s)

Michael Stravs

Examples

```
#
 dbe("C6H12O6")
```

---

deprofile *De-profile a high-resolution MS scan in profile mode.*

Description

The deprofile functions convert profile-mode high-resolution input data to "centroid"-mode data amenable to i.e. centWave. This is done using full-width, half-height algorithm, spline algorithm or local maximum method.

Usage

```
deprofile.scan(scan, noise = NA, method = "deprofile.fwhm",
     colnames = TRUE, ...)

    deprofile(df, noise, method, ...)

    deprofile.fwhm(df, noise = NA, cut = 0.5)

    deprofile.localMax(df, noise = NA)

    deprofile.spline(df, noise=NA, minPts = 5, step = 0.00001)
```

Arguments

df	A dataframe with at least the columns mz and int to perform deprofiling on.
noise	The noise cutoff. A peak is not included if the maximum stick intensity of the peak is below the noise cutoff.
method	"deprofile.fwhm" for full-width half-maximum or "deprofile.localMax" for local maximum.

...	Arguments to the workhorse functions deprofile.fwhm etc.
scan	A matrix with 2 columns for m/z and intensity; from profile-mode high-resolution data. Corresponds to the spectra obtained with xcms::getScan or mzR::peaks.
colnames	For deprofile.scan: return matrix with column names (xcms-style, TRUE, default) or plain (mzR-style, FALSE).
cut	A parameter for deprofile.fwhm indicating where the peak flanks should be taken. Standard is 0.5 (as the algorithm name says, at half maximum.) Setting cut = 0.75 would instead determine the peak width at 3/4 maximum, which might give a better accuracy for merged peaks, but could be less accurate if too few data points are present.
minPts	The minimal points count in a peak to build a spline; for peaks with less points the local maximum will be used instead. Note: The minimum value is 4!
step	The interpolation step for the calculated spline, which limits the maximum precision which can be achieved.

Details

The deprofile.fwhm method is basically an R-semantic version of the "Exact Mass" m/z deprofiler from MZmine. It takes the center between the m/z values at half-maximum intensity for the exact peak mass. The logic is stolen verbatim from the Java MZmine algorithm, but it has been rewritten to use the fast R vector operations instead of loops wherever possible. It's slower than the Java implementation, but still decently fast IMO. Using matrices instead of the data frame would be more memory-efficient and also faster, probably.

The deprofile.localMax method uses local maxima and is probably the same used by e.g. Xcalibur. For well-formed peaks, "deprofile.fwhm" gives more accurate mass results; for some applications, deprofile.localMax might be better (e.g. for fine isotopic structure peaks which are not separated by a valley and also not at half maximum.) For MS2 peaks, which have no isotopes, deprofile.fwhm is probably the better choice generally.

deprofile.spline calculates the mass using a cubic spline, as the HiRes peak detection in OpenMS does.

The word "centroid" is used for convenience to denote not-profile-mode data. The data points are NOT mathematical centroids; we would like to have a better word for it.

The noise parameter was only included for completeness, I personally don't use it.

deprofile.fwhm and deprofile.localMax are the workhorses; deprofile.scan takes a 2-column scan as input. deprofile dispatches the call to the appropriate worker method.

Value

deprofile.scan: a matrix with 2 columns for m/z and intensity

Note

Known limitations: If the absolute leftmost stick or the absolute rightmost stick in a scan are maxima, they will be discarded! However, I don't think this will ever present a practical problem.

Author(s)

Michael Stravs, Eawag <michael.stravs@eawag.ch>

References

mzMine source code http://sourceforge.net/svn/?group_id=139835

Examples

```
## Not run:
mzrFile <- openMSfile("myfile.mzML")
acqNo <- xraw@acquisitionNum[[50]]
scan.mzML.profile <- mzR::peaks(mzrFile, acqNo)
scan.mzML <- deprofile.scan(scan.mzML.profile)
close(mzrFile)

## End(Not run)
```

---

exportMassbank *Export internally stored MassBank data to files*

Description

Exports MassBank recfile data arrays and corresponding molfiles to physical files on hard disk, for one compound.

Usage

```
exportMassbank(compiled, molfile = NULL)
```

Arguments

compiled	RmbSpectraSet the spectra of one compound for which files should be exported
molfile	A molfile from createMolfile; deprecated since molfiles are not used by MassBank anymore.

Details

The data from compiled is still used here, because it contains the "visible" accession number. In the plain-text format contained in files, the accession number is not "accessible" anymore since it's in the file.

Value

No return value.

Note

An improvement would be to write the accession numbers into `names(compiled)` and later into `names(files)` so `compiled` wouldn't be needed here anymore. (The compound ID would have to go into `names(molfile)`, since it is also retrieved from `compiled`.)

Author(s)

Michael Stravs

References

MassBank record format: http://www.massbank.jp/manuals/MassBankRecord_en.pdf

See Also

`createMolfile`, `toMassbank`, `mbWorkflow`

---

fillback *Fill back reanalyzed / refiltered peak info into spectra*

Description

This method takes the info which is added to the aggregated table in the reanalysis and multiplicity filtering steps of the workflow, and adds it back into the spectra.

Usage

```
fillback(o, id, aggregated)

## S4 method for signature 'msmsWorkspace,missing,missing'
fillback(o)

## S4 method for signature 'RmbSpectraSet,missing,data.frame'
fillback(o, aggregated)

## S4 method for signature 'RmbSpectrum2,character,data.frame'
fillback(o, id, aggregated)
```

Arguments

`o`	msmsWorkspace, RmbSpectraSet or RmbSpectrum2 The object information is filled back into. If applied to an RmbSpectraSet, information is added to all its RmbSpectrum2 children. If applied to the whole msmsWorkspace, information is added to all SpectraSets.
`id`	character or missing The id of the parent RmbSpectraSet if applied to RmbSpectrum2
`aggregated`	data.frame or missing The aggregated table of the parent msmsWorkspace if applied to RmbSpectraSet or RmbSpectrum2

Value

o msmsWorkspace, RmbSpectraSet or Rmbspectrum2 The same object that was given as input with new information filled into it

---

`filterCompoundlist` *Filter a Compoundlist for missing SMILES values*

Description

Read the Compoundlist given by the filename and write a 'Compoundlist_filtered.csv', containing only the lines with a SMILES string

Usage

```
filterCompoundlist(filename)
```

Arguments

`filename`	character The name of the csv-file to be read

Value

This method has no return value.

Author(s)

pstahlhofen

Examples

```
## Not run:
   filterCompoundlist('Compoundlist.csv')

## End(Not run)
```

`filterLowaccResults` *Filter peaks with low accuracy*

Description

Filters a peak table (with annotated formulas) for accuracy. Low-accuracy peaks are removed.

Usage

```
filterLowaccResults(peaks, mode="fine", filterSettings = getOption("RMassBank")$filterSettings)
```

Arguments

`peaks`	A data frame with at least the columns `mzFound` and `dppm`.
`mode`	`coarse` or `fine`, see below.
`filterSettings`	Settings for filtering. For details, see documentation of `analyzeMsMs`

Details

In the `coarse` mode, mass tolerance is set to 10 ppm (above m/z 120) and 15 ppm (below m/z 120). This is useful for formula assignment before recalibration, where a wide window is desirable to accomodate the high mass deviations at low m/z values, so we get a nice recalibration curve.

In the `fine` run, the mass tolerance is set to 5 ppm over the whole mass range. This should be applied after recalibration.

Value

A `list(TRUE = goodPeakDataframe, FALSE = badPeakDataframe)` is returned: A data frame with all peaks which are "good" is in `return[["TRUE"]]`.

Author(s)

Michael Stravs

See Also

`analyzeMsMs`, `filterPeakSatellites`

Examples

```
# from analyzeMsMs:
## Not run: childPeaksFilt <- filterLowaccResults(childPeaksInt, filterMode)
```

filterMultiplicity *filterMultiplicity*

Description

Multiplicity filtering: Removes peaks which occur only once in a n-spectra set.

Usage

```
filterMultiplicity(w, archivename=NA, mode="pH", recalcBest = TRUE,
  multiplicityFilter = getOption("RMassBank")$multiplicityFilter)
```

Arguments

w	Workspace containing the data to be processed (aggregate table and RmbSpectraSet objects)
archivename	The archive name, used for generation of archivename_Failpeaks.csv
mode	Mode of ion analysis
recalcBest	Boolean, whether to recalculate the formula multiplicity after the first multiplicity filtering step. Sometimes, setting this to FALSE can be a solution if you have many compounds with e.g. fluorine atoms, which often have multiple assigned formulas per peak and might occasionally lose peaks because of that.
multiplicityFilter	Threshold for the multiplicity filter. If set to 1, no filtering will apply (minimum 1 occurrence of peak). 2 equals minimum 2 occurrences etc.

Details

This function executes multiplicity filtering for a set of spectra using the workhorse function filterPeaksMultiplicity (see details there) and retrieves problematic filtered peaks (peaks which are of high intensity but were discarded, because either no formula was assigned or it was not present at least 2x), using the workhorse function problematicPeaks. The results are returned in a format ready for further processing with mbWorkflow.

Value

A list object with values:

peaksOK	Peaks with >1-fold formula multiplicity from the "normal" peak analysis.
peaksReanOK	Peaks with >1-fold formula multiplicity from peak reanalysis.
peaksFiltered	All peaks with annotated formula multiplicity from first analysis.
peaksFilteredReanalysis	All peaks with annotated formula multiplicity from peak reanalysis.
peaksProblematic	Peaks with high intensity which do not match inclusion criteria -> possible false negatives. The list will be exported into archivename_failpeaks.csv.

Author(s)

Michael Stravs

See Also

filterPeaksMultiplicity,problematicPeaks

Examples

```
## Not run:
    refilteredRcSpecs <- filterMultiplicity(
   w, "myarchive", "pH")

## End(Not run)
```

---

filterPeakSatellites *Filter satellite peaks*

Description

Filters satellite peaks in FT spectra which arise from FT artifacts and from conversion to stick mode. A very simple rule is used which holds mostly true for MSMS spectra (and shouldn't be applied to MS1 spectra which contain isotope structures...)

Usage

```
filterPeakSatellites(peaks, filterSettings = getOption("RMassBank")$filterSettings)
```

Arguments

peaks	A peak dataframe with at least the columns mz, int. Note that mz is used even for the recalibrated spectra, i.e. the desatellited spectrum is identical for both the unrecalibrated and the recalibrated spectra.
filterSettings	The settings used for filtering. Refer to analyzeMsMs documentation for filter settings.

Details

The function cuts off all peaks within 0.5 m/z from every peak, in decreasing intensity order, which are below 5 intensity. E.g. for peaks m/z=100, int=100; m/z=100.2, int=2, m/z=100.3, int=6, m/z 150, int=10: The most intense peak (m/z=100) is selected, all neighborhood peaks below 5 peak) and the next less intense peak is selected. Here this is the m/z=150 peak. All low-intensity neighborhood peaks are removed (nothing). The next less intense peak is selected (m/z=100.3) and again neighborhood peaks are cut away (nothing to cut here. Note that the m/z = 100.2 peak was alredy removed.)

Value

Returns the peak table with satellite peaks removed.

Note

This is a very crude rule, but works remarkably well for our spectra.

Author(s)

Michael Stravs

See Also

analyzeMsMs, filterLowaccResults

Examples

```
# From the workflow:
## Not run:
    # Filter out satellite peaks:
    shot <- filterPeakSatellites(shot)
    shot_satellite_n <- setdiff(row.names(shot_full), row.names(shot))
    shot_satellite <- shot_full[shot_satellite_n,]
    # shot_satellite contains the peaks which were eliminated as satellites.

## End(Not run)
```

---

filterPeaksMultiplicity

Multiplicity filtering: Removes peaks which occur only once in a n-spectra set.

Description

For every compound, every peak (with annotated formula) is compared across all spectra. Peaks whose formula occurs only once for all collision energies / spectra types, are discarded. This eliminates "stochastic formula hits" of pure electronic noise peaks efficiently from the spectra. Note that in the author's experimental setup two spectra were recorded at every collision energy, and therefore every peak-formula should appear at least twice if it is real, even if it is by chance a fragment which appears on only one collision energy setting. The function was not tested in a different setup. Therefore, use with a bit of caution.

Usage

```
filterPeaksMultiplicity(w, recalcBest = TRUE)
```

Arguments

w	a ‘msmsWorkspace‘ object where formulas have been assigned to peaks
recalcBest	Whether the best formula for each peak should be re-determined. This is necessary for results from the ordinary analyzeMsMs analysis which allows multiple potential formulas per peak - the old best match could potentially have been dropped because of multiplicity filtering. For results from reanalyzeFailpeak this is not necessary, since only one potential formula is assigned in this case.

Value

The peak table is returned, enriched with columns:

- formulaMultiplicityThe # of occurrences of this formula in the spectra of its compounds.

Author(s)

Michael Stravs, EAWAG <michael.stravs@eawag.ch>

Examples

```
## Not run:
  peaksFiltered <- filterPeaksMultiplicity(peaksMatched(w),
   "formula", TRUE)
  peaksOK <- subset(peaksFiltered, formulaMultiplicity > 1)

## End(Not run)
```

---

findEIC *Extract EICs*

Description

Extract EICs from raw data for a determined mass window.

Usage

```
findEIC(
  msRaw,
  mz,
  limit = NULL,
  rtLimit = NA,
  headerCache = NULL,
  floatingRecalibration = NULL,
  peaksCache = NULL,
  polarity = NA,
  msLevel = 1,
  precursor = NULL
)
```

Arguments

msRaw	The mzR file handle
mz	The mass or mass range to extract the EIC for: either a single mass (with the range specified by limit below) or a mass range in the form of c(min, max).
limit	If a single mass was given for mz: the mass window to extract. A limit of 0.001 means that the EIC will be returned for [mz - 0.001, mz + 0.001].
rtLimit	If given, the retention time limits in form c(rtmin, rtmax) in seconds.
headerCache	If present, the complete mzR::header(msRaw). Passing this value is useful if spectra for multiple compounds should be extracted from the same mzML file, since it avoids getting the data freshly from msRaw for every compound.
floatingRecalibration	A fitting function that predict()s a mass shift based on the retention time. Can be used if a lockmass calibration is known (however you have to build the calibration yourself.)
peaksCache	If present, the complete output of mzR::peaks(msRaw). This speeds up the lookup if multiple compounds should be searched in the same file.
polarity	If a value is given, scans are filtered to this polarity before EIC extraction. Valid values are 1 for positive, 0 for negative (according to mzR), or a RMassBank 'mode' (e.g. 'pH, pM, mH...') from which the polarity can be derived.
msLevel	Which MS level to target for EIC extraction. By default this is 1; level 2 can be used to extract the EIC of fragments for a specific precursor or to extract EICs from DIA data.
precursor	Which precursor to target for EIC extraction. If 'NULL', the scans are not filtered by precursor. Use this only for 'msLevel != 1'. If a 'precursor' filter is set for 'msLevel == 1', all scans will be filtered out.

Value

A [rt, intensity, scan] matrix (scan being the scan number.)

Author(s)

Michael A. Stravs, Eawag <michael.stravs@eawag.ch>

See Also

findMsMsHR

findMass *Calculate exact mass*

Description

Retrieves the exact mass of the uncharged molecule. It works directly from the SMILES and therefore is used in the MassBank workflow (mbWorkflow) - there, all properties are calculated from the SMILES code retrieved from the database. (Alternatively, takes also the compound ID as parameter and looks it up.) Calculation relies on Rcdk.

Usage

```
findMass(cpdID_or_smiles, retrieval = "standard", mode = "pH")
```

Arguments

cpdID_or_smiles	SMILES code or compound ID of the molecule. (Numerics are treated as compound ID).
retrieval	A value that determines whether the files should be handled either as "standard", if the compoundlist is complete, "tentative", if at least a formula is present or "unknown" if the only know thing is the m/z
mode	"pH", "pNa", "pM", "pNH4", "mH", "mM", "mFA" for different ions ([M+H]+, [M+Na]+, [M]+, [M+NH4]+, [M-H]-, [M]-, [M+FA]-). Only needed for retrieval="unknown"

Value

Returns the exact mass of the uncharged molecule.

Author(s)

Michael Stravs

See Also

findMz

Examples

```
##
findMass("OC[C@H]1OC(O)[C@H](O)[C@@H](O)[C@@H]1O")
```

findMsMsHR *Extract MS/MS spectra for specified precursor*

Description

Extracts MS/MS spectra from LC-MS raw data for a specified precursor, specified either via the RMassBank compound list (see loadList) or via a mass.

Usage

```
findMsMsHR(
  fileName = NULL,
  msRaw = NULL,
  cpdID,
  mode = "pH",
  confirmMode = 0,
  useRtLimit = TRUE,
  ppmFine = getOption("RMassBank")$findMsMsRawSettings$ppmFine,
  mzCoarse = getOption("RMassBank")$findMsMsRawSettings$mzCoarse,
 fillPrecursorScan = getOption("RMassBank")$findMsMsRawSettings$fillPrecursorScan,
  rtMargin = getOption("RMassBank")$rtMargin,
  deprofile = getOption("RMassBank")$deprofile,
  headerCache = NULL,
  peaksCache = NULL,
  enforcePolarity = getOption("RMassBank")$enforcePolarity,
  diaWindows = getOption("RMassBank")$findMsMsRawSettings$diaWindows
)

findMsMsHR.mass(
  msRaw,
  mz,
  limit.coarse,
  limit.fine,
  rtLimits = NA,
  maxCount = NA,
  headerCache = NULL,
  fillPrecursorScan = FALSE,
  deprofile = getOption("RMassBank")$deprofile,
  peaksCache = NULL,
  cpdID = NA,
  polarity = NA,
  diaWindows = getOption("RMassBank")$findMsMsRawSettings$diaWindows
)
```

Arguments

fileName	The file to open and search the MS2 spectrum in.

msRaw	The opened raw file (mzR file handle) to search the MS2 spectrum in. Specify either this or fileName.
cpdID	The compound ID in the compound list (see loadList) to use for formula lookup. Note: In \codefindMsMsHR.mass, this is entirely optional and used only in case a warning must be displayed; compound lookup is done via mass only.
mode	The processing mode (determines which ion/adduct is searched): "pH", "pNa", "pM", "pNH4", "mH", "mM", "mFA" for different ions ([M+H]+, [M+Na]+, [M]+, [M+NH4]+, [M-H]-, [M]-, [M+FA]-).
confirmMode	Whether to use the highest-intensity precursor (=0), second- highest (=1), third-highest (=2)...
useRtLimit	Whether to respect retention time limits from the compound list.
ppmFine	The limit in ppm to use for fine limit (see below) calculation.
mzCoarse	The coarse limit to use for locating potential MS2 scans: this tolerance is used when finding scans with a suitable precursor ion value.
fillPrecursorScan	If TRUE, the precursor scan will be filled from MS1 data. To be used for data where the precursor scan is not stored in the raw data.
rtMargin	The retention time tolerance to use.
deprofile	Whether deprofiling should take place, and what method should be used (cf. deprofile)
headerCache	If present, the complete mzR::header(msRaw). Passing this value is useful if spectra for multiple compounds should be extracted from the same mzML file, since it avoids getting the data freshly from msRaw for every compound.
peaksCache	If present, the complete output of mzR::peaks(msRaw). This speeds up the lookup if multiple compounds should be searched in the same file.
enforcePolarity	If TRUE, scans are filtered for the given 'mode''s polarity when finding the target spectrum.
diaWindows	A data frame with columns precursorMz, mzMin, mzMax which specifies the precursor and window size of each window for DIA acquisition.
mz	The mass to use for spectrum search.
limit.coarse	Parameter in findMsMsHR.mass corresponding to mzCoarse. (The parameters are distinct to clearly conceptually distinguish findMsMsHR.mass (a standalone useful function) from the cpdID based functions (workflow functions).)
limit.fine	The fine limit to use for locating MS2 scans: this tolerance is used when locating an appropriate analyte peak in the MS1 precursor spectrum.
rtLimits	c(min, max): Minimum and maximum retention time to use when locating the MS2 scans.
maxCount	The maximal number of spectra groups to return. One spectra group consists of all data-dependent scans from the same precursor whose precursor mass matches the specified search mass.
polarity	If set (for ?findMsMsHR.mass), scans are filtered for the given 'mode''s polarity when finding the target spectrum.

Details

Different versions of the function get the data from different sources. Note that findMsMsHR and findMsMsHR.direct differ mainly in that findMsMsHR opens a file whereas findMsMs.direct uses an open file handle - both are intended to be used in a full process which involves compound lists etc. In contrast, findMsMsHR.mass is a low-level function which uses the mass directly for lookup and is intended for use as a standalone function in unrelated applications.

Value

An `RmbSpectraSet` (for `findMsMsHR`). Contains parent MS1 spectrum (`@parent`), a block of dependent MS2 spectra ((`@children`) and some metadata (`id`,`mz`,`name`,`mode` in which the spectrum was acquired.

For `findMsMsHR.mass`: a list of `RmbSpectraSets` as defined above, sorted by decreasing precursor intensity.

Functions

- `findMsMsHR.mass()`: A submethod of find MsMsHR that retrieves basic spectrum data

Note

`findMsMs.direct` is deactivated

Author(s)

Michael A. Stravs, Eawag <michael.stravs@eawag.ch>

See Also

findEIC

Examples

```
## Not run:
   loadList("mycompoundlist.csv")
   # if Atrazine has compound ID 1:
  msms_atrazine <- findMsMsHR(fileName = "Atrazine_0001_pos.mzML", cpdID = 1, mode = "pH")
   # Or alternatively:
   msRaw <- openMSfile("Atrazine_0001_pos.mzML")
   msms_atrazine <- findMsMsHR(msRaw=msRaw, cpdID = 1, mode = "pH")
   # Or directly by mass (this will return a list of spectra sets):
   mz <- findMz(1)$mzCenter
   msms_atrazine_all <- findMsMsHR.mass(msRaw, mz, 1, ppm(msRaw, 10, p=TRUE))
   msms_atrazine <- msms_atrazine_all[[1]]

## End(Not run)
```

findMsMsHR.direct *Discontinued: find MS/MS spectrum from open raw file*

Description

This interface has been discontinued. findMsMsHR now supports the same parameters (use named parameters).

Usage

```
findMsMsHR.direct(
  msRaw,
  cpdID,
  mode = "pH",
  confirmMode = 0,
  useRtLimit = TRUE,
  ppmFine = getOption("RMassBank")$findMsMsRawSettings$ppmFine,
  mzCoarse = getOption("RMassBank")$findMsMsRawSettings$mzCoarse,
 fillPrecursorScan = getOption("RMassBank")$findMsMsRawSettings$fillPrecursorScan,
  rtMargin = getOption("RMassBank")$rtMargin,
  deprofile = getOption("RMassBank")$deprofile,
  headerCache = NULL
)
```

Arguments

msRaw	x
cpdID	x
mode	x
confirmMode	x
useRtLimit	x
ppmFine	x
mzCoarse	x
fillPrecursorScan	x
rtMargin	x
deprofile	x
headerCache	x

Value

an error

Author(s)

stravsmi

`findMsMsHR.ticms2` *Extract an MS/MS spectrum from MS2 TIC*

Description

Extract an MS/MS spectrum or multiple MS/MS spectra based on the TIC of the MS2 and precursor mass, picking the most intense MS2 scan. Can be used, for example, to get a suitable MS2 from direct infusion data which was collected with purely targeted MS2 without MS1.

Usage

```
findMsMsHR.ticms2(
  msRaw,
  mz,
  limit.coarse,
  limit.fine,
  rtLimits = NA,
  maxCount = NA,
  headerCache = NULL,
  fillPrecursorScan = FALSE,
  deprofile = getOption("RMassBank")$deprofile,
  trace = "ms2tic"
)
```

Arguments

`msRaw`	The mzR raw file
`mz`	Mass to find
`limit.coarse`	Allowed mass deviation for scan precursor (in m/z values)
`limit.fine`	Unused here, but present for interface compatiblity with findMsMsHR
`rtLimits`	Unused here, but present for interface compatiblity with findMsMsHR
`maxCount`	Maximal number of spectra to return
`headerCache`	Cached results of header(msRaw), either to speed up the operations or to operate with preselected header() data
`fillPrecursorScan`	Unused here, but present for interface compatiblity with findMsMsHR
`deprofile`	Whether deprofiling should take place, and what method should be used (cf. `deprofile`)
`trace`	Either `"ms2tic"` or `"ms2basepeak"`: Which intensity trace to use - can be either the TIC of the MS2 or the basepeak intensity of the MS2.

Details

Note that this is not a precise function and only really makes sense in direct infusion and if the precursor is really known, because MS2 precursor data is only "roughly" accurate (to 2 dp). The regular `findMsMsHR` functions confirm the exact mass of the precursor in the MS1 scan.

Value

a list of "spectrum sets" as defined in findMsMsHR, sorted by decreasing precursor intensity.

Author(s)

stravsmi

`findMsMsHRperMsp` *Retrieve spectra from msp files*

Description

This function is currently used to read msp files containing data that were already processed in order to convert the results to MassBank records.

Usage

```
findMsMsHRperMsp(fileName, cpdIDs, mode = "pH")

findMsMsHRperMsp.direct(fileName, cpdIDs, mode = "pH")
```

Arguments

`fileName`	vector of character-strings The msp files to be searched for spectra
`cpdIDs`	vector of integers The IDs of compounds in the compoundlist for which spectra should be retrieved
`mode`	character, default: "pH" The processing mode that was used to produce the spectrum. Should be one of "pH": ([M+H]+) "pNa": ([M+Na]+) "pM": ([M]+) "mH": ([M-H]-) or "mFA": ([M+FA]-) (see the `RMassBank` vignette)

Value

An `RmbSpectraSet` with integrated information from the msp files

Functions

- `findMsMsHRperMsp.direct()`: A submethod of findMsMsHrperxcms that retrieves basic spectrum data

`findMsMsHRperxcms` *Read in mz-files using XCMS*

Description

Picks peaks from mz-files and returns the pseudospectra that CAMERA creates with the help of XCMS

Usage

```
findMsMsHRperxcms(
  fileName,
  cpdID,
  mode = "pH",
  findPeaksArgs = NULL,
  plots = FALSE,
  MSe = FALSE
)

findMsMsHRperxcms.direct(
  fileName,
  cpdID,
  mode = "pH",
  findPeaksArgs = NULL,
  plots = FALSE,
  MSe = FALSE
)
```

Arguments

`fileName`	The path to the mz-file that should be read
`cpdID`	The compoundID(s) of the compound that has been used for the file
`mode`	The ionization mode that has been used for the spectrum represented by the peaklist
`findPeaksArgs`	A list of arguments that will be handed to the xcms-method findPeaks via do.call
`plots`	A parameter that determines whether the spectra should be plotted or not
`MSe`	A boolean value that determines whether the spectra were recorded using MSe or not

Value

The spectra generated from XCMS

Functions

- `findMsMsHRperxcms.direct()`: A submethod of findMsMsHrperxcms that retrieves basic spectrum data

Author(s)

Erik Mueller

See Also

msmsWorkflow toRMB

Examples

```
## Not run:
 fileList <- list.files(system.file("XCMSinput", package = "RMassBank"), "Glucolesquerellin", full.names=TRUE)[3
 loadList(system.file("XCMSinput/compoundList.csv",package="RMassBank"))
     psp <- findMsMsHRperxcms(fileList,2184)

## End(Not run)
```

---

findMz *Find compound information*

Description

Retrieves compound information from the loaded compound list or calculates it from the SMILES code in the list.

Usage

```
findMz(cpdID, mode = "pH", ppm = 10, deltaMz = 0, retrieval="standard",
unknownMass = getOption("RMassBank")$unknownMass )

findRt(cpdID)

findSmiles(cpdID)

findFormula(cpdID, retrieval="standard")

findCAS(cpdID)

findName(cpdID)

findLevel(cpdID, compact=FALSE)
```

Arguments

cpdID	The compound ID in the compound list.

mode	Specifies the species of the molecule: An empty string specifies uncharged monoisotopic mass, *pH* (positive H) specifies [M+H]+, *pNa* specifies [M+Na]+, *pM* specifies [M]+, *mH* and *mFA* specify [M-H]- and [M+FA]-, respectively. (I apologize for the naming of *pH* which has absolutely nothing to do with chemical *pH* values.)
ppm	Specifies ppm window (10 ppm will return the range of the molecular mass + and - 10 ppm).
deltaMz	Specifies additional m/z window to add to the range (deltaMz = 0.02 will return the range of the molecular mass +- 0.02 (and additionally +- the set ppm value).
retrieval	A value that determines whether the files should be handled either as "standard", if the compoundlist is complete, "tentative", if at least a formula is present or "unknown" if the only know thing is the m/z
unknownMass	'charged' or 'neutral' ('charged' assumed by default) specifies whether a mass of an unknown compound (level 5) refers to the charged or neutral mass (and correspondingly, whether it must be shifted or not to find the m/z value)
compact	Only for `findLevel`, returns the "retrieval" parameter used for many functions within RMassBank if TRUE

Value

`findMz` will return a `list(mzCenter=, mzMin=, mzMax=)` with the molecular weight of the given ion, as calculated from the SMILES code and Rcdk.

`findRt`, `findSmiles`,`findCAS`,`findName` will return the corresponding entry from the compound list. `findFormula` returns the molecular formula as determined from the SMILES code.

Author(s)

Michael Stravs

See Also

`findMass`, `loadList`, `findMz.formula`

Examples

```
## Not run: %
 findMz(123, "pH", 5)
 findFormula(123)

## End(Not run)
```

`findMz.formula` *Find the exact mass +/- a given margin for a given formula or its ions and adducts.*

Description

Find the exact mass +/- a given margin for a given formula or its ions and adducts.

Usage

```
findMz.formula(formula, mode = "pH", ppm = 10, deltaMz = 0)
```

Arguments

`formula`	The molecular formula in text or list format (see `formulastring.to.list`
`mode`	`"pH"`, `"pNa"`, `"pM"`, `"pNH4"`, `"mH"`, `"mM"`, `"mFA"` for different ions ([M+H]+, [M+Na]+, [M]+, [M+NH4]+, [M-H]-, [M]-, [M+FA]-). "" for the uncharged molecule.
`ppm`	The ppm margin to add/subtract
`deltaMz`	The absolute mass to add/subtract. Cumulative with `ppm`

Value

A `list(mzMin=, mzCenter=, mzMax=)` with the masses.

Author(s)

Michael A. Stravs, Eawag <michael.stravs@eawag.ch>

See Also

`findMz`

Examples

```
findMz.formula("C6H6")
```

findProgress *Determine processed steps*

Description

This function reads out the content of different slots of the workspace object and finds out which steps have already been processed on it.

Usage

```
findProgress(workspace)
```

Arguments

workspace	A msmsWorkspace object.

Value

An array containing all msmsWorkflow steps which have likely been processed.

Author(s)

Stravs MA, Eawag <michael.stravs@eawag.ch>

Examples

```
## Not run:
findProgress(w)

## End(Not run)
```

flatten *Flatten, or re-read, MassBank header blocks*

Description

flatten converts a list of MassBank compound information sets (as retrieved by gatherData) to a flat table, to be exported into an infolist. readMbdata reads a single record from an infolist flat table back into a MassBank (half-)entry.

Usage

```
flatten(mbdata)

readMbdata(row)
```

Arguments

mbdata	A list of MassBank compound information sets as returned from gatherData.
row	One row of MassBank compound information retrieved from an infolist.

Details

Neither the flattening system itself nor the implementation are particularly fantastic, but since hand-checking of records is a necessary evil, there is currently no alternative (short of coding a complete GUI for this and working directly on the records.)

Value

flatten returns a tibble (not a data frame or matrix) to be written to CSV.

readMbdata returns a list of type list(id= *compoundID* , . . . , 'ACCESSION' = '', 'RECORD_TITLE' = '',) etc.

Author(s)

Michael Stravs

References

MassBank record format: http://www.massbank.jp/manuals/MassBankRecord_en.pdf

See Also

gatherData,loadInfolist

Examples

```
## Not run:
 # Collect some data to flatten
 ids <- c(40,50,60,70)
 data <- lapply(ids, gatherData)
 # Flatten the data trees to a table
 flat.table <- flatten(data)
 # reimport the table into a tree
 data.reimported <- apply(flat.table, 1, readMbdata)

## End(Not run)
```

`formulastring.to.list` *Interconvert molecular formula representations*

Description

Converts molecular formulas from string to list representation or vice versa.

Usage

```
list.to.formula(flist)

formulastring.to.list(formula)
```

Arguments

`formula`	A molecular formula in string format, e.g. `"C6H12O6"`.
`flist`	A molecular formula in list format, e.g. `list( "C" = 6, "H" = 12, "O" = 6 )`.

Details

The function doesn't care about whether your formula makes sense. However, `"C3.5O4"` will give `list("C" = 3, "O" = 4)` because regular expressions are used for matching (however, `list("C" = 3.5, "O" = 4)` gives `"C3.5O4"`.) Duplicate elements cause problems; only "strict" molecular formulas ("CH4O", but not "CH3OH") work correctly.

Value

`list.to.formula` returns a string representation of the formula; `formulastring.to.list` returns the list representation.

Author(s)

Michael Stravs

See Also

`add.formula`, `order.formula`, `is.valid.formula`

Examples

```
#
 list.to.formula(list("C" = 4, "H" = 12))
 # This is also OK and useful to calculate e.g. adducts or losses.
 list.to.formula(list("C" = 4, "H" = -1))
 formulastring.to.list(list.to.formula(formulastring.to.list("CHIBr")))
```

gatherCCTE *Retrieve supplemental annotation data from US EPA*

Description

Retrieves annotation data for a compound from the internet service US EPA CCTE based on the inchikey generated by babel or Cactus

Usage

```
gatherCCTE(key, api_key)
```

Arguments

key	An Inchi-Key or other chemical identifier (e.g. Chemical name, DTXSID, CASRN, InChIKey, DTXCID)
api_key	An US EPA CCTE API key (personal or application)

Details

The data retrieved is the US EPA DTXSID, the US EPA chemical dashboard substance ID, the CAS-RN, the DTX preferred name, and the DTXCID (chemical ID).

Value

Returns a list with 5 slots: `dtxsid` The US EPA chemical dashboard substance id `dtxcid` The US EPA chemical dashboard chemical id `preferredName` The US EPA chemical dashboard preferred name `casrn` The latest CAS registration number `smiles` The SMILES annotation of the structure

Author(s)

Tobias Schulze

References

CCTE REST: https://api-ccte.epa.gov/docs/

See Also

mbWorkflow

Examples

```
# Gather data for compound ID 131
## Not run: gatherCCTE("QEIXBXXKTUNWDK-UHFFFAOYSA-N", api_key = NA)
```

gatherData *Retrieve annotation data*

Description

Retrieves annotation data for a compound from the internet services CTS, Pubchem, Chemspider and Cactvs, based on the SMILES code and name of the compounds stored in the compound list.

Usage

```
gatherData(id)
```

Arguments

id	The compound ID.

Details

Composes the "upper part" of a MassBank record filled with chemical data about the compound: name, exact mass, structure, CAS no., links to PubChem, KEGG, ChemSpider. The instrument type is also written into this block (even if not strictly part of the chemical information). Additionally, index fields are added at the start of the record, which will be removed later: `id`, `dbcas`, `dbname` from the compound list, `dataused` to indicate the used identifier for CTS search (`smiles` or `dbname`).

Additionally, the fields `ACCESSION` and `RECORD_TITLE` are inserted empty and will be filled later on.

Value

Returns a list of type `list(id=`*compoundID*`, ..., 'ACCESSION' = '', 'RECORD_TITLE' = '', )` etc.

Author(s)

Michael Stravs

References

Chemical Translation Service: http://uranus.fiehnlab.ucdavis.edu:8080/cts/homePage cactus Chemical Identifier Resolver: http://cactus.nci.nih.gov/chemical/structure MassBank record format: http://www.massbank.jp/manuals/MassBankRecord_en.pdf Pubchem REST: https://pubchem.ncbi.nlm.nih.gov/pug_rest/PUG_REST.html Chemspider InChI conversion: https://www.chemspider.com/InChI.asmx

See Also

mbWorkflow

Examples

```
# Gather data for compound ID 131
## Not run: gatherData(131)
```

`gatherDataBabel` *Retrieve annotation data*

Description

Retrieves annotation data for a compound by using babel, based on the SMILES code and name of the compounds stored in the compound list.

Usage

```
gatherDataBabel(id)
```

Arguments

`id`	The compound ID.

Details

Composes the "upper part" of a MassBank record filled with chemical data about the compound: name, exact mass, structure, CAS no.. The instrument type is also written into this block (even if not strictly part of the chemical information). Additionally, index fields are added at the start of the record, which will be removed later: `id`, `dbcas`, `dbname` from the compound list.

Additionally, the fields `ACCESSION` and `RECORD_TITLE` are inserted empty and will be filled later on.

This function is an alternative to gatherData, in case CTS is down or if information on one or more of the compounds in the compound list are sparse

Value

Returns a list of type `list(id=`*compoundID*`, ...,'ACCESSION' = '', 'RECORD_TITLE' = '', )` etc.

Author(s)

Michael Stravs, Erik Mueller

References

MassBank record format: http://www.massbank.jp/manuals/MassBankRecord_en.pdf

See Also

mbWorkflow

Examples

```
# Gather data for compound ID 131
## Not run: gatherDataBabel(131)
```

`gatherDataUnknown` *Retrieve annotation data*

Description

Retrieves annotation data for an unknown compound by using basic information present

Usage

```
gatherDataUnknown(id, mode, retrieval)
```

Arguments

`id`	The compound ID.
`mode`	`"pH"`, `"pNa"`, `"pM"`, `"pNH4"`, `"mH"`, `"mM"`, `"mFA"` for different ions ([M+H]+, [M+Na]+, [M]+, [M+NH4]+, [M-H]-, [M]-, [M+FA]-).
`retrieval`	A value that determines whether the files should be handled either as "standard", if the compoundlist is complete, "tentative", if at least a formula is present or "unknown" if the only know thing is the m/z

Details

Composes the "upper part" of a MassBank record filled with chemical data about the compound: name, exact mass, structure, CAS no.. The instrument type is also written into this block (even if not strictly part of the chemical information). Additionally, index fields are added at the start of the record, which will be removed later: `id, dbcas, dbname` from the compound list.

Additionally, the fields `ACCESSION` and `RECORD_TITLE` are inserted empty and will be filled later on.

This function is used to generate the data in case a substance is unknown, i.e. not enough information is present to derive anything about formulas or links

Value

Returns a list of type `list(id=` *compoundID* `, ...,'ACCESSION' = '', 'RECORD_TITLE' = '', )` etc.

Author(s)

Michael Stravs, Erik Mueller

References

MassBank record format: http://www.massbank.jp/manuals/MassBankRecord_en.pdf

See Also

mbWorkflow

Examples

```
# Gather data for compound ID 131
## Not run: gatherDataUnknown(131,"pH")
```

---

gatherPubChem *Retrieve supplemental annotation data from Pubchem*

Description

Retrieves annotation data for a compound from the internet service Pubchem based on the inchikey generated by babel or Cactus

Usage

```
gatherPubChem(key)
```

Arguments

key	An Inchi-Key

Details

The data retrieved is the Pubchem CID, a synonym from the Pubchem database, the IUPAC name (using the preferred if available) and a Chebi link

Value

Returns a list with 4 slots: PcID The Pubchem CID Synonym An arbitrary synonym for the compound name IUPAC A IUPAC-name (preferred if available) Chebi The identification number of the chebi database

Author(s)

Erik Mueller

References

Pubchem REST: https://pubchem.ncbi.nlm.nih.gov/pug_rest/PUG_REST.html Chebi: http://www.ebi.ac.uk/chebi

See Also

mbWorkflow

Examples

```
# Gather data for compound ID 131
## Not run: gatherPubChem("QEIXBXXKTUNWDK-UHFFFAOYSA-N")
```

`getAnalyticalInfo` *Get analytical info for MassBank record*

Description

Collects the info for 'ai, ac_lc, ac_ms' for general, LC and MS info respectively. The info comes from the settings except for the compound-specific part, which is omitted if there is no 'cpd' specified.

Usage

```
getAnalyticalInfo(cpd = NULL)
```

Arguments

cpd	A 'RmbSpectraSet' object

`getCactus` *Retrieve information from Cactus*

Description

Retrieves information from the Cactus Chemical Identifier Resolver (PubChem).

Usage

```
getCactus(identifier, representation)
```

Arguments

identifier	Any identifier interpreted by the resolver, e.g. an InChI key or a SMILES code.
representation	The desired representation, as required from the resolver. e.g. `stdinchikey`, `chemspider_id`, `formula`... Refer to the webpage for details.

Details

It is not necessary to specify in which format the `identifier` is. Somehow, cactus does this automatically.

Value

The result of the query, in plain text. Can be NA, or one or multiple lines (character array) of results.

Note

Note that the InChI key is retrieved with a prefix (`InChIkey=`), which must be removed for most database searches in other databases (e.g. CTS).

Author(s)

Michael Stravs

References

cactus Chemical Identifier Resolver: http://cactus.nci.nih.gov/chemical/structure

See Also

getCtsRecord, getPcId

Examples

```
# Benzene:
getCactus("C1=CC=CC=C1", "cas")
getCactus("C1=CC=CC=C1", "stdinchikey")
getCactus("C1=CC=CC=C1", "chemspider_id")
```

`getCASRN` *Search CCTE CAS registration number*

Description

Retrieves CCTE CASRN from US EPA for a search term.

Usage

```
getCASRN(key, api_key)
```

Arguments

`key`	ID to be converted
`api_key`	API key for CCTE

Details

Only the first result is returned currently. **The function should be regarded as experimental and has not thoroughly been tested.**

Value

The CCTE CAS RN (in string type)

Author(s)

Tobias Schulze

References

CCTE search: https://api-ccte.epa.gov/docs

CCTE REST: https://api-ccte.epa.gov/docs

Examples

```
## Not run:
getCASRN("MKXZASYAUGDDCJ-NJAFHUGGSA-N")

## End(Not run)
```

`getCSID` *Retrieve the Chemspider ID for a given compound*

Description

Given an InChIKey, this function queries the chemspider web API to retrieve the Chemspider ID of he compound with that InChIkey.

Usage

```
getCSID(query)
```

Arguments

`query`	The InChIKey of the compound

Value

Returns the chemspide

Author(s)

Michele Stravs, Eawag <stravsmi@eawag.ch>

Erik Mueller, UFZ <erik.mueller@ufz.de>

Examples

```
## Not run:
# Return all CAS registry numbers stored for benzene.
data <- getCtsRecord("UHOVQNZJYSORNB-UHFFFAOYSA-N")
cas <- CTS.externalIdSubset(data, "CAS")

## End(Not run)
```

`getCtsKey` *Convert a single ID to another using CTS.*

Description

Convert a single ID to another using CTS.

Usage

```
getCtsKey(query, from = "Chemical Name", to = "InChIKey")
```

Arguments

`query`	ID to be converted
`from`	Type of input ID
`to`	Desired output ID

Value

An unordered array with the resulting converted key(s).

Author(s)

Michele Stravs, Eawag <stravsmi@eawag.ch>

Examples

```
k <- getCtsKey("benzene", "Chemical Name", "InChIKey")
```

getCtsRecord *Retrieve information from CTS*

Description

Retrieves a complete CTS record from the InChI key.

Usage

```
getCtsRecord(key)
```

Arguments

key	The InChI key.

Value

Returns a list with all information from CTS: `inchikey`,`inchicode`, `formula`, `exactmass` contain single values. `synonyms` contains an unordered list of scored synonyms (`type`, `name`, `score`, where `type` indicates either a normal name or a specific IUPAC name, see below). `externalIds` contains an unordered list of identifiers of the compound in various databases (`name`, `value`, where `name` is the database name and `value` the identifier in that database.)

Note

Currently, the CTS results are still incomplete; the name scores are all 0, formula and exact mass return zero.

Author(s)

Michele Stravs, Eawag <stravsmi@eawag.ch>

References

Chemical Translation Service: https://cts.fiehnlab.ucdavis.edu

Examples

```
data <- getCtsRecord("UHOVQNZJYSORNB-UHFFFAOYSA-N")
# show all synonym "types"
types <- unique(unlist(lapply(data$synonyms, function(i) i$type)))
## Not run: print(types)
```

getData *Get data frame with all present peak data*

Description

Returns a data frame with columns for all non-empty slots in a RmbSpectrum2 object. Note that MSnbase::Spectrum has a method as.data.frame, however that one will return only mz, intensity. This function is kept separate to ensure downwards compatibility since it returns more columns than MSnbase as.data.frame.

Usage

```
## S4 method for signature 'RmbSpectrum2'
getData(s)
```

Arguments

s	The RmbSpectrum2 object to extract data from.

Value

A data frame with columns for every set slot.

Author(s)

stravsmi

getDTXCID *Search CCTE DTXCID*

Description

Retrieves CCTE DTXCID from US EPA for a search term.

Usage

```
getDTXCID(key, api_key)
```

Arguments

key	ID to be converted
api_key	API key for CCTE

Details

Only the first result is returned currently. **The function should be regarded as experimental and has not thoroughly been tested.**

Value

The DTXCID (in string type)

Author(s)

Tobias Schulze

References

CCTE search: https://api-ccte.epa.gov/docs

CCTE REST: https://api-ccte.epa.gov/docs

Examples

```
## Not run:
getDTXCID("MKXZASYAUGDDCJ-NJAFHUGGSA-N")

## End(Not run)
```

---

getDTXSID *Search CCTE DTXSID*

Description

Retrieves CCTE DTXSID from US EPA for a search term.

Usage

```
getDTXSID(key, api_key)
```

Arguments

key	ID to be converted
api_key	API key for CCTE

Details

Only the first result is returned currently. **The function should be regarded as experimental and has not thoroughly been tested.**

Value

The DTXSID (in string type)

Author(s)

Tobias Schulze

References

CCTE search: https://api-ccte.epa.gov/docs

CCTE REST: https://api-ccte.epa.gov/docs

Examples

```
## Not run:
getDTXSID("MKXZASYAUGDDCJ-NJAFHUGGSA-N")

## End(Not run)
```

`getDTXSMILES` *Search CCTE SMILES*

Description

Retrieves CCTE SMILES from US EPA for a search term.

Usage

```
getDTXSMILES(key, api_key)
```

Arguments

`key`	ID to be converted
`api_key`	API key for CCTE

Details

Only the first result is returned currently. **The function should be regarded as experimental and has not thoroughly been tested.**

Value

The SMILES (in string type)

Author(s)

Tobias Schulze

References

CCTE search: https://api-ccte.epa.gov/docs

CCTE REST: https://api-ccte.epa.gov/docs

Examples

```
## Not run:
getDTXSMILES("MKXZASYAUGDDCJ-NJAFHUGGSA-N")

## End(Not run)
```

---

getField *Get the content of a field in a JCAMP file*

Description

The content will always be returned as character-string

Usage

```
getField(parsedJDX, field_name)
```

Arguments

parsedJDX	list as created by readJDX A parsed, single-block JCAMP file
field_name	character The name of the field (e.g. 'CAS REGISTRY NO')

Value

The field's content

Author(s)

pstahlhofen

See Also

readJDX

Examples

```
## Not run:
   parsedJDX <- readJDX('my_singleblock_jcamp.dx')
   title <- getField(parsedJDX, "TITLE")

## End(Not run)
```

`getMolecule` *Create Rcdk molecule from SMILES*

Description

Generates a Rcdk molecule object from SMILES code, which is fully typed and usable (in contrast to the built-in `parse.smiles`).

Usage

```
getMolecule(smiles)
```

Arguments

`smiles`	The SMILES code of the compound.

Details

NOTE: As of today (2012-03-16), Rcdk discards stereochemistry when loading the SMILES code! Therefore, do not trust this function blindly, e.g. don't generate InChI keys from the result. It is, however, useful if you want to compute the mass (or something else) with Rcdk.

Value

A Rcdk `IAtomContainer` reference.

Author(s)

Michael Stravs

See Also

`parse.smiles`

Examples

```
# Lindane:
getMolecule("C1(C(C(C(C(C1Cl)Cl)Cl)Cl)Cl)Cl")
# Benzene:
getMolecule("C1=CC=CC=C1")
```

getPcId *Search Pubchem CID*

Description

Retrieves PubChem CIDs for a search term.

Usage

```
getPcId(query, from = "inchikey")
```

Arguments

query	ID to be converted
from	Type of input ID

Details

Only the first result is returned currently. **The function should be regarded as experimental and has not thoroughly been tested.**

Value

The PubChem CID (in string type).

Author(s)

Michael Stravs, Erik Mueller

References

PubChem search: http://pubchem.ncbi.nlm.nih.gov/

Pubchem REST: https://pubchem.ncbi.nlm.nih.gov/pug_rest/PUG_REST.html

See Also

getCtsRecord, getCactus

Examples

```
getPcId("MKXZASYAUGDDCJ-NJAFHUGGSA-N")
```

getPrefName *Search CCTE Preferred Name*

Description

Retrieves CCTE Preferred Name from US EPA for a search term.

Usage

```
getPrefName(key, api_key)
```

Arguments

key	ID to be converted
api_key	API key for CCTE

Details

Only the first result is returned currently. **The function should be regarded as experimental and has not thoroughly been tested.**

Value

The CCTE Preferred Name (in string type)

Author(s)

Tobias Schulze

References

CCTE search: https://api-ccte.epa.gov/docs

CCTE REST: https://api-ccte.epa.gov/docs

Examples

```
## Not run:
getPrefName("MKXZASYAUGDDCJ-NJAFHUGGSA-N")

## End(Not run)
```

`is.valid.formula` *Check validity of formula*

Description

Checks whether the formula is chemically valid, i.e. has no zero-count or negative-count elements.

Usage

```
is.valid.formula(formula)
```

Arguments

`formula`	A molecular formula in string or list representation (`"C6H6"` or `list(C=6,H=6)`).

Details

The check is only meant to identify formulas which have negative elements, which can arise from the subtraction of adducts. It is **not** a high-level formula "validity" check like e.g. the Rcdk function `isvalid.formula` which uses the nitrogen rule or a DBE rule.

Author(s)

Michael Stravs

See Also

`list.to.formula`, `add.formula`, `order.formula`

Examples

```
#
is.valid.formula(list(C=0,H=1,Br=2))
is.valid.formula("CH2Cl")
is.valid.formula("C0H2")
```

`loadInfolists` *Load MassBank compound information lists*

Description

Loads MassBank compound information lists (i.e. the lists which were created in the first two steps of the MassBank `mbWorkflow` and subsequently edited by hand.).

Usage

```
loadInfolists(mb, path)

 loadInfolist(mb, fileName)

 resetInfolists(mb)
```

Arguments

`mb`	The `mbWorkspace` to load/reset the lists in.
`path`	Directory in which the namelists reside. All CSV files in this directory will be loaded.
`fileName`	A single namelist to be loaded.

Details

`resetInfolists` clears the information lists, i.e. it creates a new empty list in `mbdata_archive`. `loadInfolist` loads a single CSV file, whereas `loadInfolists` loads a whole directory.

Value

The new workspace with loaded/reset lists.

Author(s)

Michael Stravs, Tobias Schulze

Examples

```
#
## Not run: mb <- resetInfolists(mb)
 mb <- loadInfolist(mb, "my_csv_infolist.csv")
## End(Not run)
```

loadList *Load compound list for RMassBank*

Description

Loads a CSV compound list with compound IDs

Usage

```
loadList(path, listEnv=NULL, check=TRUE)

resetList()
```

Arguments

path	Path to the CSV list.
listEnv	The environment to load the list into. By default, the namelist is loaded into an environment internally in RMassBank.
check	A parameter that specifies whether the SMILES-Codes in the list should be checked for readability by rcdk.

Details

The list is loaded into the variable *compoundList* in the environment `listEnv` (which defaults to the global environment) and used by the `findMz`, `findCAS`, ... functions. The CSV file is required to have at least the following columns, which are used for further processing and must be named correctly (but present in any order): *ID*, *Name*, *SMILES*, *RT*, *CAS*

resetList() clears a currently loaded list.

Value

No return value.

Author(s)

Michael Stravs

See Also

`findMz`

Examples

```
##
## Not run: loadList("mylist.csv")
```

makeMollist *Write list.tsv file*

Description

Makes a list.tsv file in the "moldata" folder.

Usage

```
makeMollist(compiled)
```

Arguments

compiled	list of RmbSpectraSet compiled spectra for multiple compounds (one RmbSpectraSet each).

Details

Generates the list.tsv file which is needed by MassBank to connect records with their respective molfiles. The first compound name is linked to a mol-file with the compound ID (e.g. 2334.mol for ID 2334).

Value

No return value.

Author(s)

Michael A. Stravs, Eawag <michael.stravs@eawag.ch>

makePeaksCache *Generate peaks cache*

Description

Generates a peak cache table for use with findMsMsHR functions.

Usage

```
makePeaksCache(msRaw, headerCache)
```

Arguments

msRaw	the input raw datafile (opened)
headerCache	the cached header, or subset thereof for which peaks should be extracted. Peak extraction goes by seqNum.

Value

A list of dataframes as from `mzR::peaks`.

Author(s)

stravsmi

`makeRecalibration` *Recalibrate MS/MS spectra*

Description

Recalibrates MS/MS spectra by building a recalibration curve of the assigned putative fragments of all spectra in `aggregatedSpecs` (measured mass vs. mass of putative associated fragment) and additionally the parent ion peaks.

Usage

```
makeRecalibration(w,
  recalibrateBy = getOption("RMassBank")$recalibrateBy,
  recalibrateMS1 = getOption("RMassBank")$recalibrateMS1,
  recalibrator = getOption("RMassBank")$recalibrator,
  recalibrateMS1Window = getOption("RMassBank")$recalibrateMS1Window
  )

 recalibrateSpectra(rawspec = NULL, rc = NULL, rc.ms1=NULL, w = NULL,
  recalibrateBy = getOption("RMassBank")$recalibrateBy,
  recalibrateMS1 = getOption("RMassBank")$recalibrateMS1)




 recalibrateSingleSpec(spectrum, rc,
  recalibrateBy = getOption("RMassBank")$recalibrateBy)
```

Arguments

`w`	For `makeRecalibration`: to perform the recalibration with. For `recalibrateSpectra`: the `msmsWorkspace` which contains the recalibration curves (alternatively to specifying `rc`, `rc.ms1`).
`recalibrateBy`	Whether recalibration should be done by ppm ("ppm") or by m/z ("mz").
`recalibrateMS1`	Whether MS1 spectra should be recalibrated separately ("separate"), together with MS2 ("common") or not at all ("none"). Usually taken from settings.
`recalibrator`	The recalibrator functions to be used. Refer to `recalibrate` for details. Usually taken from settings.

`recalibrateMS1Window`
: Window width to look for MS1 peaks to recalibrate (in ppm).

`spectrum`
: For `recalibrateSingleSpec`: a `MSnbase` `Spectrum`-derived object, commonly a `RmbSpectrum2` for MS2 or `Spectrum1` for MS1.

`rawspec`
: For `recalibrateSpectra`:an `RmbSpectraSetList` of `RmbSpectraSet` objects , as the `w@spectra` slot from `msmsWorkspace` or any object returned by `findMsMsHR`. If empty, no spectra are recalibrated, but the recalibration curve is returned.

`rc`, `rc.ms1`
: The recalibration curves to be used in the recalibration.

Details

Note that the actually used recalibration functions are governed by the general MassBank settings (see `recalibrate`).

If a set of acquired LC-MS runs contains spectra for two different ion types (e.g. [M+H]+ and [M+Na]+) which should both be processed by RMassBank, it is necessary to do this in two separate runs. Since it is likely that one ion type will be the vast majority of spectra (e.g. most in [M+H]+ mode), and only few spectra will be present for other specific adducts (e.g. only few [M+Na]+ spectra), it is possible that too few spectra are present to build a good recalibration curve using only e.g. the [M+Na]+ ions. Therefore we recommend, for one set of LC/MS runs, to build the recalibration curve for one ion type (`msmsWorkflow(mode="pH", steps=c(1:8), newRecalibration=TRUE)`) and reuse the same curve for processing different ion types (`msmsWorkflow(mode="pNa", steps=c(1:8), newRecalibration=FALSE)`). This also ensures a consistent recalibration across all spectra of the same batch.

Value

`makeRecalibration`: a `list(rc, rc.ms1)` with recalibration curves for the MS2 and MS1 spectra.

`recalibrateSpectra`: if `rawspec` is not `NULL`, returns the recalibrated spectra as `RmbSpectraSetList`. All spectra have their mass recalibrated and evaluation data deleted.

`recalibrateSingleSpec`: the recalibrated `Spectrum` (same object, recalibrated masses, evaluation data like assigned formulae etc. deleted).

Author(s)

Michael Stravs, Eawag <michael.stravs@eawag.ch>

Examples

```
## Not run:
   rcCurve <- recalibrateSpectra(w, "pH")
   w@spectra <- recalibrateSpectra(mode="pH", rawspec=w@spectra, w=myWorkspace)
  w@spectra <- recalibrateSpectra(mode="pH", rawspec=w@spectra, rcCurve$rc, rcCurve$rc.ms1)

## End(Not run)
```

mbWorkflow *MassBank record creation workflow*

Description

Uses data generated by msmsWorkflow to create MassBank records.

Usage

```
mbWorkflow(
  mb,
  steps = c(1, 2, 3, 4, 5, 6, 7, 8),
  infolist_path = "./infolist.csv",
  gatherData = "online",
  filter = TRUE
)
```

Arguments

mb	The mbWorkspace to work in.
steps	Which steps in the workflow to perform.
infolist_path	A path where to store newly downloaded compound informations, which should then be manually inspected.
gatherData	A variable denoting whether to retrieve information using several online databases gatherData= "online" or to use the local babel installation gatherData= "babel". Note that babel is used either way, if a directory is given in the settings. This setting will be ignored if retrieval is set to "standard"
filter	If TRUE, the peaks will be filtered according to the standard processing workflow in RMassBank - only the best formula for a peak is retained, and only peaks passing multiplicity filtering are retained. If FALSE, it is assumed that the user has already done filtering, and all peaks in the spectrum should be printed in the record (with or without formula.)

Details

See the vignette vignette("RMassBank") for detailed informations about the usage.

Steps:

Step 1: Find which compounds don't have annotation information yet. For these compounds, pull information from several databases (using gatherData).

Step 2: If new compounds were found, then export the infolist.csv and stop the workflow. Otherwise, continue.

Step 3: Take the archive data (in table format) and reformat it to MassBank tree format.

Step 4: Compile the spectra. Using the skeletons from the archive data, create MassBank records per compound and fill them with peak data for each spectrum. Also, assign accession numbers based on scan mode and relative scan no.

Step 5: Convert the internal tree-like representation of the MassBank data into flat-text string arrays (basically, into text-file style, but still in memory)

Step 6: For all OK records, generate a corresponding molfile with the structure of the compound, based on the SMILES entry from the MassBank record. (This molfile is still in memory only, not yet a physical file)

Step 7: If necessary, generate the appropriate subdirectories, and actually write the files to disk.

Step 8: Create the list.tsv in the molfiles folder, which is required by MassBank to attribute substances to their corresponding structure molfiles.

Value

The processed `mbWorkspace`.

Author(s)

Michael A. Stravs, Eawag <michael.stravs@eawag.ch>

See Also

`mbWorkspace-class`

Examples

```
## Not run:
  mb <- newMbWorkspace(w) # w being a msmsWorkspace
  mb <- loadInfolists(mb, "D:/myInfolistPath")
  mb <- mbWorkflow(mb, steps=c(1:3), "newinfos.csv")


## End(Not run)
```

`mbWorkspace-class` *Workspace for* `mbWorkflow` *data*

Description

A workspace which stores input and output data for use with `mbWorkflow`.

Usage

```
## S4 method for signature 'mbWorkspace'
show(object)
```

Arguments

`object`	The `mbWorkspace` to display.

Details

Slots:

spectra, aggregated The corresponding input data from msmsWorkspace-class

additionalPeaks A list of additional peaks which can be loaded using addPeaks.

mbdata, mbdata_archive, mbdata_relisted Infolist data: Data for annotation of MassBank records, which can be loaded using loadInfolists.

compiled, compiled_ok Compiled tree-structured MassBank records. compiled_ok contains only the compounds with at least one valid spectrum.

mbfiles Compiled MassBank records in text representation.

molfile MOL files with the compound structures.

ok,problems Index lists for internal use which denote which compounds have valid spectra.

Methods:

show Shows a brief summary of the object. Currently only a stub.

Author(s)

Michael Stravs, Eawag <michael.stravs@eawag.ch>

See Also

mbWorkflow

---

mergePeaks *Merge peaks for spectra merging, FT shoulder elimination etc.*

Description

This procedure first sorts peaks by intensity (descending sort) and then starts iterating over the peaks, removing all entries that deviate "sufficiently far" from the currently selected peak. See the Details section for a full explanation and information on how to fine-tune peak removal.

Usage

```
mergePeaks(peaks, ...)

## S4 method for signature 'data.frame'
mergePeaks(peaks, ...)

## S4 method for signature 'matrix'
mergePeaks(peaks, ...)

## S4 method for signature 'RmbSpectrum2'
mergePeaks(peaks, ...)

## S4 method for signature 'Spectrum'
mergePeaks(peaks, ...)
```

Arguments

peaks	data.frame, matrix or RmbSpectrum2 The peak-table to be merged. In case of an RmbSpectrum2-object, peaks are retrieved and updated via getData and setData, respectively
...	3 numeric values These define cutoff limits (see details)

Details

Three parameters must be passed to mergePeaks for peak-removal control in this order: - cutoff_dppm_limit - cutoff_absolute_limit - cutoff_intensity_limit The method iterates through the peaks, beginning with the highest-intensity peak and in each step removes all other peaks that fulfill conditions 1 AND 2 relative to the selected peak 1. Their m/z value does not deviate too far from the one of the selected peak. i.e. if the selected peak is p and the checked peak is c, it holds that EITHER |p$mz - c$mz| <= cutoff_absolute_limit OR |p$mz - c$mz| <= ppm(p$mz, cutoff_dppm_limit, p=TRUE) (see ppm) 2. Their intensity is much smaller than the one of the selected peak, i.e. c$mz < cutoff_intensity_limit * p$mz for a suitable cutoff_intensity_limit between 0 and 1.

Value

object of the same class as peaks The result contains a reduced peak-table ordered by m/z

See Also

getData, setData, ppm

Examples

```
## Not run: mergePeaks(spectrum, 10, 0.5, 0.05)
```

---

mergeSpectra *Merge multiple spectra into one*

Description

This method takes a collection of RmbSpectrum2 objects and merges them into a single RmbSpectrum2 object

Usage

```
mergeSpectra(spectra, ...)

## S4 method for signature 'RmbSpectrum2List'
mergeSpectra(spectra, ...)
```

Arguments

spectra	RmbSpectrum2List A list of RmbSpectrum2 objects to be merged
...	NOTHING (This parameter is reserved for future implementations of the generic)

Details

Information from all spectra is retrieved via getData combined with rbind and placed into the new spectrum with setData

Value

A single RmbSpectrum2 object containing the merged information

See Also

getData, setData

---

msmsRead *Extracts and processes spectra from a specified file list, according to loaded options and given parameters.*

Description

The filenames of the raw LC-MS runs are read from the array files in the global enviroment. See the vignette vignette("RMassBank") for further details about the workflow.

Usage

```
msmsRead(
  w,
  filetable = NULL,
  files = NULL,
  cpdids = NULL,
  readMethod,
  mode = NULL,
  confirmMode = FALSE,
  useRtLimit = TRUE,
  Args = NULL,
  settings = getOption("RMassBank"),
  progressbar = "progressBarHook",
  MSe = FALSE,
  plots = FALSE
)
```

Arguments

w	A msmsWorkspace to work with.
filetable	The path to a .csv-file that contains the columns "Files" and "ID" supplying the relationships between files and compound IDs. Either this or the parameter "files" need to be specified.
files	A vector or list containing the filenames of the files that are to be read as spectra. For the IDs to be inferred from the filenames alone, there need to be exactly 2 underscores.
cpdids	A vector or list containing the compound IDs of the files that are to be read as spectra. The ordering of this and files implicitly assigns each ID to the corresponding file. If this is supplied, then the IDs implicitly named in the filenames are ignored.
readMethod	Several methods are available to get peak lists from the files. Currently supported are "mzR", "xcms", "MassBank" and "peaklist". The first two read MS/MS raw data, and differ in the strategy used to extract peaks. MassBank will read existing records, so that e.g. a recalibration can be performed, and "peaklist" just requires a CSV with two columns and the column header "mz", "int".
mode	"pH", "pNa", "pM", "pNH4", "mH", "mM", "mFA" for different ions ([M+H]+, [M+Na]+, [M]+, [M+NH4]+, [M-H]-, [M]-, [M+FA]-). For 'readMethod == "mzR"', a vector of 'mode' entries is supported. The user should check that they are either all positive or negative. If this isn't the case, the recalibration will be incorrect.
confirmMode	Defaults to false (use most intense precursor). Value 1 uses the 2nd-most intense precursor for a chosen ion (and its data-dependent scans) , etc.
useRtLimit	Whether to enforce the given retention time window.
Args	A list of arguments that will be handed to the xcms-method findPeaks via do.call
settings	Options to be used for processing. Defaults to the options loaded via loadRmbSettings et al. Refer to there for specific settings.
progressbar	The progress bar callback to use. Only needed for specialized applications. Cf. the documentation of progressBarHook for usage.
MSe	A boolean value that determines whether the spectra were recorded using MSe or not
plots	A boolean value that determines whether the pseudospectra in XCMS should be plotted

Value

The msmsWorkspace with msms-spectra read.

Author(s)

Michael Stravs, Eawag <michael.stravs@eawag.ch>

Erik Mueller, UFZ

See Also

msmsWorkspace-class, msmsWorkflow

---

msmsRead.RAW *Extracts and processes spectra from a list of xcms-Objects*

Description

The filenames of the raw LC-MS runs are read from the array `files` in the global enviroment. See the vignette `vignette("RMassBank")` for further details about the workflow.

Usage

```
msmsRead.RAW(
  w,
  xRAW = NULL,
  cpdids = NULL,
  mode,
  findPeaksArgs = NULL,
  settings = getOption("RMassBank"),
  progressbar = "progressBarHook",
  plots = FALSE
)
```

Arguments

w	A msmsWorkspace to work with.
xRAW	A list of xcmsRaw objects whose peaks should be detected and added to the workspace. The relevant data must be in the MS1 data of the xcmsRaw object. You can coerce the msn-data in a usable object with the msn2xcmsRaw function of xcms.
cpdids	A vector or list containing the compound IDs of the files that are to be read as spectra. The ordering of this and files implicitly assigns each ID to the corresponding file. If this is supplied, then the IDs implicitly named in the filenames are ignored.
mode	"pH", "pNa", "pM", "pNH4", "mH", "mM", "mFA" for different ions ([M+H]+, [M+Na]+, [M]+, [M+NH4]+, [M-H]-, [M]-, [M+FA]-).
findPeaksArgs	A list of arguments that will be handed to the xcms-method findPeaks via do.call
settings	Options to be used for processing. Defaults to the options loaded via loadRmbSettings et al. Refer to there for specific settings.
progressbar	The progress bar callback to use. Only needed for specialized applications. Cf. the documentation of progressBarHook for usage.
plots	A boolean value that determines whether the pseudospectra in XCMS should be plotted

Value

The msmsWorkspace with msms-spectra read.

Author(s)

Michael Stravs, Eawag <michael.stravs@eawag.ch>

Erik Mueller, UFZ

See Also

msmsWorkspace-class, msmsWorkflow

---

msmsWorkflow *RMassBank mass spectrometry pipeline*

Description

Extracts and processes spectra from a specified file list, according to loaded options and given parameters.

Usage

```
msmsWorkflow(
  w,
  mode = "pH",
  steps = c(1:8),
  confirmMode = FALSE,
  newRecalibration = TRUE,
  useRtLimit = TRUE,
  archivename = NA,
  readMethod = "mzR",
  filetable = NULL,
  findPeaksArgs = NULL,
  plots = FALSE,
  precursorscan.cf = FALSE,
  settings = getOption("RMassBank"),
  analyzeMethod = "formula",
  progressbar = "progressBarHook",
  MSe = FALSE
)
```

Arguments

w	A msmsWorkspace to work with.
mode	"pH", "pNa", "pM", "mH", "mM", "mFA", "pNH4" for different ions ([M+H]+, [M+Na]+, [M]+, [M-H]-, [M]-, [M+FA]-, [M+NH4]+).

steps	Which steps of the workflow to process. See the vignette vignette("RMassBank") for details.
confirmMode	Defaults to false (use most intense precursor). Value 1 uses the 2nd-most intense precursor for a chosen ion (and its data-dependent scans) , etc.
newRecalibration	Whether to generate a new recalibration curve (TRUE, default) or to reuse the currently stored curve (FALSE, useful e.g. for adduct-processing runs.)
useRtLimit	Whether to enforce the given retention time window.
archivename	The prefix under which to store the analyzed result files.
readMethod	Several methods are available to get peak lists from the files. Currently supported are "mzR", "xcms", "MassBank" and "peaklist". The first two read MS/MS raw data, and differ in the strategy used to extract peaks. MassBank will read existing records, so that e.g. a recalibration can be performed, and "peaklist" just requires a CSV with two columns and the column header "mz", "int".
filetable	If including step 1 (data extraction), a 'filetable' argument to be passed to msmsRead.
findPeaksArgs	A list of arguments that will be handed to the xcms-method findPeaks via do.call
plots	A parameter that determines whether the spectra should be plotted or not (This parameter is only used for the xcms-method)
precursorscan.cf	Whether to fill precursor scans. To be used with files which for some reasons do not contain precursor scan IDs in the mzML, e.g. AB Sciex converted files.
settings	Options to be used for processing. Defaults to the options loaded via loadRmbSettings et al. Refer to there for specific settings.
analyzeMethod	The "method" parameter to pass to analyzeMsMs.
progressbar	The progress bar callback to use. Only needed for specialized applications. Cf. the documentation of progressBarHook for usage.
MSe	A boolean value that determines whether the spectra were recorded using MSe or not

Details

The filenames of the raw LC-MS runs are read from the array files in the global enviroment. See the vignette vignette("RMassBank") for further details about the workflow.

Value

The processed msmsWorkspace.

Author(s)

Michael Stravs, Eawag <michael.stravs@eawag.ch>

See Also

msmsWorkspace-class

`msmsWorkspace-class` *Workspace for* `msmsWorkflow` *data*

Description

A workspace which stores input and output data for `msmsWorkflow`.

Usage

```
## S4 method for signature 'msmsWorkspace'
show(object)
```

Arguments

`object`	The `msmsWorkspace` to display.

Details

Slots:

files The input file names

spectra The spectra per compound (`RmbSpectraSet`) extracted from the raw files

aggregated A data.frame with an aggregated peak table from all `spectra`. Further columns are added during processing.

rc, rc.ms1 The recalibration curves generated in workflow step 4.

parent For the workflow steps after 4: the parent workspace containing the state (spectra, aggregate) before recalibration, such that the workflow can be reprocessed from start.

archivename The base name of the files the archive is stored to during the workflow.

settings The RMassBank settings used during the workflow, if stored with the workspace.#'

Methods:

show Shows a brief summary of the object and processing progress.

Author(s)

Michael Stravs, Eawag <michael.stravs@eawag.ch>

See Also

`msmsWorkflow`

newMbWorkspace *Create new workspace for* mbWorkflow

Description

Creates a new workspace for use with mbWorkflow.

Usage

```
newMbWorkspace(w)
```

Arguments

w	The input msmsWorkspace to load input data from.

Details

The workspace input data will be loaded from the msmsWorkspace-class object provided by the parameter w.

Value

A new mbWorkflow object with the loaded input data.

Author(s)

Michael Stravs, Eawag <michael.stravs@eawag.ch>

See Also

mbWorkflow, msmsWorkspace-class

newMsmsWorkspace *Create new empty workspace or load saved data for* msmsWorkflow

Description

Creates an empty workspace or loads an existing workspace from disk.

Usage

```
newMsmsWorkspace(files = character(0))
```

Arguments

files	If given, the files list to initialize the workspace with.

Details

newMsmsWorkspace creates a new empty workspace for use with msmsWorkflow.

loadMsmsWorkspace loads a workspace saved using archiveResults. Note that it also successfully loads data saved with the old RMassBank data format into the new msmsWorkspace object.

Value

A new msmsWorkspace object

Author(s)

Michael Stravs, Eawag <michael.stravs@eawag.ch>

See Also

msmsWorkflow, msmsWorkspace-class

---

normalize,RmbSpectrum2-method

Scale spectrum to specified intensity range

Description

Scale spectrum to specified intensity range

Usage

```
## S4 method for signature 'RmbSpectrum2'
normalize(object, ..., scale = 999, precision = 0, slot = "intensity")
```

Arguments

object	the 'RmbSpectrum2' object to scale
...	arguments passed to 'selectPeaks' to choose peaks for normalization
scale	Maximum intensity in normalized spectrum
precision	Digits after comma for normalized intensity, typically 0
slot	Which property of the spectrum should be scaled

normalize,RmbSpectrum2List-method *Normalize spectra*

Description

Scale all spectra in a 'RmbSpectrum2List' to a specified intensity.

Usage

```
## S4 method for signature 'RmbSpectrum2List'
normalize(object, ...)
```

Arguments

object	the 'RmbSpectrum2List' with spectra to scale
...	Arguments passed to 'normalize,RmbSpectrum2'

order.formula *Order a chemical formula correctly*

Description

Orders a chemical formula in the commonly accepted order (CH followed by alphabetic ordering).

Usage

```
order.formula(formula, as.formula = TRUE, as.list = FALSE)
```

Arguments

formula	A molecular formula in string or list representation ("C6H6" or list(C=6,H=6)).
as.formula	If TRUE, the return value is returned as a string. This is the default.
as.list	If TRUE, the return value is returned in list representation.

Author(s)

Michele Stravs

See Also

list.to.formula, add.formula, is.valid.formula

Examples

```
#
order.formula("H4C9")
order.formula("C2N5HClBr")
```

`parseMassBank` *MassBank-record Parser*

Description

Can parse MassBank-records(only V2)

Usage

```
parseMassBank(Files)
```

Arguments

`Files`	array of character-strings Paths to the plaintext-records that should be read

Value

The `mbWorkspace` that the plaintext-record creates. All parsed information will be stored in the 'compiled_ok' slot.

Author(s)

Erik Mueller

See Also

`validate`

Examples

```
## Not run:
 paths <- c("filepath_to_records/RC000001.txt",
            "filepath_to_records/RC000002.txt")
  mb <- parseMassBank(paths)

## End(Not run)
```

parseMbRecord *MassBank-record Parser*

Description

Can parse MassBank-records(only V2)

Usage

```
parseMbRecord(filename, readAnnotation=TRUE)
```

Arguments

filename	character A path to the plaintext-record that should be read
readAnnotation	logical, Default: TRUE If TRUE, parse annotations from the record file and add columns for 'formula', 'formulaCount', 'mzCalc' and 'dppm' to the peak table

Value

An RmbSpectrum2 object created from the plaintext-record

Author(s)

Erik Mueller

See Also

validate

Examples

```
## Not run:
  parseMassBank("filepath_to_records/RC00001.txt")

## End(Not run)
```

peaksMatched *Select matching/unmatching peaks from aggregate table*

Description

Select matching/unmatching peaks from aggregate table

Usage

```
peaksMatched(o)

## S4 method for signature 'data.frame'
peaksMatched(o)

## S4 method for signature 'msmsWorkspace'
peaksMatched(o)
```

Arguments

o	Workspace or aggregate table from a workspace

Value

Selects the peaks from the aggregate table which matched within filter criteria (peaksMatched) or didn't match (peaksUnmatched).

Functions

- peaksMatched(data.frame): A method to retrieve the matched peaks from the "aggregated" slot (a data.frame object) in an msmsWorkSpace
- peaksMatched(msmsWorkspace): A method to retrieve the matched peaks from an msmsWorkSpace

Author(s)

stravsmi

peaksUnmatched *Select matching/unmatching peaks from aggregate table*

Description

Select matching/unmatching peaks from aggregate table

Usage

```
peaksUnmatched(o, cleaned = FALSE)

## S4 method for signature 'data.frame'
peaksUnmatched(o, cleaned = FALSE)

## S4 method for signature 'msmsWorkspace'
peaksUnmatched(o, cleaned = FALSE)
```

Arguments

o	Workspace or aggregate table from a workspace
cleaned	Return only peaks which pass electronic noise filtering if TRUE.

Value

Selects the peaks from the aggregate table which matched within filter criteria (peaksMatched) or didn't match (peaksUnmatched).

Methods (by class)

- peaksUnmatched(data.frame): A method to retrieve the unmatched peaks from the "aggregated" slot (a data.frame object) in an msmsWorkSpace
- peaksUnmatched(msmsWorkspace): A method to retrieve the unmatched peaks from an msmsWorkSpace

Author(s)

stravsmi

---

plotMbWorkspaces *Plots mbWorkspaces*

Description

Plots the peaks of one or two mbWorkspace to compare them.

Usage

```
plotMbWorkspaces(w1, w2 = NULL)
```

Arguments

w1	The mbWorkspace to be plotted
w2	Another optional mbWorkspace be plotted as a reference.

Details

This functions plots one or two mbWorkspaces in case the use has used different methods to acquire similar spectra. w1 must always be supplied, while w2 is optional. The wokspaces need to be fully processed for this function to work.

Value

A logical indicating whether the information was plotted or not

Author(s)

Erik Mueller

Examples

```
#
## Not run: plotMbWorkspaces(w1,w2)
```

---

plotRecalibration *Plot the recalibration graph.*

Description

Plot the recalibration graph.

Usage

```
plotRecalibration(w, recalibrateBy = getOption("RMassBank")$recalibrateBy)

  plotRecalibration.direct(rcdata, rc, rc.ms1, title, mzrange,
  recalibrateBy = getOption("RMassBank")$recalibrateBy)
```

Arguments

w	The workspace to plot the calibration graph from
recalibrateBy	Whether recalibration was done by ppm ("ppm") or by m/z ("mz"). Important only for graph labeling here.
rcdata	A data frame with columns recalfield and mzFound.
rc	Predictor for MS2 data
rc.ms1	Predictor for MS1 data
title	Prefix for the graph titles
mzrange	m/z value range for the graph

Author(s)

Michele Stravs, Eawag <michael.stravs@eawag.ch>

ppm *Calculate ppm values*

Description

Calculates ppm values for a given mass.

Usage

```
ppm(mass, dppm, l = FALSE, p = FALSE)
```

Arguments

mass	The "real" mass
dppm	The mass deviation to calculate
l	Boolean: return limits? Defaults to FALSE.
p	Boolean: return ppm error itself? Defaults to FALSE.

Details

This is a helper function used in RMassBank code.

Value

By default (`l=FALSE, p=FALSE`) the function returns the mass plus the ppm error (for 123.00000 and 10 ppm: 123.00123, or for 123 and -10 ppm: 122.99877).

For `l=TRUE`, the function returns the upper and lower limit (sic!) For `p=TRUE`, just the difference itself is returned (0.00123 for 123/10ppm).

Author(s)

Michael A. Stravs, Eawag <michael.stravs@eawag.ch>

Examples

```
ppm(100, 10)
```

problematicPeaks *Identify intense peaks (in a list of unmatched peaks)*

Description

Finds a list of peaks in spectra with a high relative intensity (>10 1e4, or >1 checked. Peaks orbiting around the parent peak mass (calculated from the compound ID), which are very likely co-isolated substances, are ignored.

Usage

```
problematicPeaks(sp)
```

Arguments

sp	a RmbSpectrum2 object to be checked for problematic peaks.

Value

The modified RmbSpectrum2 object with additional columns/properties ‘problematicPeaks‘ (logical ‘TRUE‘ if the peak is intense and unannotated), ‘aMax‘ (base peak intensity), ‘mzCenter‘ (the precursor m/z).

Note

TODO: there is hardcoded logic in this function that needs to be resolved eventually!

Author(s)

Michael Stravs

See Also

msmsWorkflow

Examples

```
# As used in the workflow:

sp <- new("RmbSpectrum2", mz = c(100,200,300,400,500), intensity = c(999999,888888,777777,666666,555555))
sp@ok <- TRUE
property(sp, "mzFound", addNew=TRUE) <- sp@mz
sp@good <- c(TRUE, TRUE, TRUE, FALSE, FALSE)
sp@precursorMz <- 600
sp_checked <- problematicPeaks(sp)
# stopifnot(sum(getData(sp_checked)$problematicPeak) == 2)
```

processProblematicPeaks *Generate list of problematic peaks*

Description

Generates a list of intense unmatched peaks for further review (the "failpeak list") and exports it if the archive name is given.

Usage

```
processProblematicPeaks(w, archivename = NA)
```

Arguments

w	msmsWorkspace to analyze.
archivename	Base name of the archive to write to (for "abc" the exported failpeaks list will be "abc_Failpeaks.csv"). if the compoundlist is complete, "tentative", if at least a formula is present or "unknown" if the only know thing is the m/z

Value

Returns the aggregate data.frame with added column "problematic" (logical) which marks peaks which match the problematic criteria

Author(s)

stravsmi

progressBarHook *Standard progress bar hook.*

Description

This function provides a standard implementation for the progress bar in RMassBank.

Usage

```
progressBarHook(object = NULL, value = 0, min = 0, max = 100, close = FALSE)
```

Arguments

object	An identifier representing an instance of a progress bar.
value	The new value to assign to the progress indicator
min	The minimal value of the progress indicator
max	The maximal value of the progress indicator
close	If TRUE, the progress bar is closed.

Details

RMassBank calls the progress bar function in the following three ways: `pb <- progressBarHook(object=NULL, value=0, min=0, max=LEN)` to create a new progress bar. `pb <- progressBarHook(object=pb, value= VAL)` to set the progress bar to a new value (between the set `min` and `max`) `progressBarHook(object=pb, close=TRUE)` to close the progress bar. (The actual calls are performed with `do.call`, e.g. `progressbar <- "progressBarHook" pb <- do.call(progressbar, list(object=pb, value= nProg))` . See the source code for details.)

To substitute the standard progress bar for an alternative implementation (e.g. for use in a GUI), the developer can write his own function which behaves in the same way as `progressBarHook`, i.e. takes the same parameters and can be called in the same way.

Value

Returns a progress bar instance identifier (i.e. an identifier which can be used as `object` in subsequent calls.)

Author(s)

Michele Stravs, Eawag <stravsmi@eawag.ch>

`property` *Get a property of an RmbSpectrum2 object*

Description

This searches the 'properties' slot of the object and returns a column with matching name (if found) or NULL otherwise.

Usage

```
property(o, property)

## S4 method for signature 'RmbSpectrum2,character'
property(o, property)
```

Arguments

`o`	`RmbSpectrum2`
`property`	character The name of a property

Value

The corresponding column of `o@properties`

property<- *Replacement function to set properties of an RmbSpectrum2 object*

Description

Update the 'properties' slot of the given object. If the column you want to update does not exist yet and addNew = FALSE (default), this will cause a warning and the object will not be changed

Usage

```
property(o, property, addNew=FALSE, class="") <- value

## S4 replacement method for signature 'RmbSpectrum2,character,logical,character'
property(o, property, addNew = FALSE, class = "") <- value

## S4 replacement method for signature 'RmbSpectrum2,character,missing,character'
property(o, property, addNew = FALSE, class = "") <- value

## S4 replacement method for signature 'RmbSpectrum2,character,logical,missing'
property(o, property, addNew = FALSE, class = "") <- value

## S4 replacement method for signature 'RmbSpectrum2,character,missing,missing'
property(o, property, addNew = FALSE, class = "") <- value
```

Arguments

o	RmbSpectrum2 The object whos 'properties' slot should be updated
property	character The name of the column in the 'properties' data frame to be updated
addNew	logical, Default: FALSE Whether or not a new column should be added in case a column of the given name does not exist yet.
class	character or missing The class of the entries for the column to be added/updated
value	ANY The value(s) to be written into the column

Details

Please note that this is a replacement method, meaning that property(o, property) <- value can be used as a short-hand for the equivalent o <- 'property<-'(o, property, value)

Value

The RmbSpectrum2 object with an updated 'properties' slot

`reanalyzeFailpeaks` *Reanalyze unmatched peaks*

Description

Reanalysis of peaks with no matching molecular formula by allowing additional elements (e.g. "N2O").

Usage

```
reanalyzeFailpeaks(w, custom_additions, filterSettings =
    getOption("RMassBank")$filterSettings, progressbar = "progressBarHook")
reanalyzeFailpeak(mass, custom_additions, cpdID, mode,
    filterSettings = getOption("RMassBank")$filterSettings)
```

Arguments

`w`	A 'msmsWorkspace' with annotated peaks.
`custom_additions`	The allowed additions, e.g. "N2O".
`filterSettings`	Settings for filtering data. Refer to`analyzeMsMs` for settings.
`progressbar`	The progress bar callback to use. Only needed for specialized applications. Cf. the documentation of `progressBarHook` for usage.
`mass`	(Usually recalibrated) m/z value of the peak.
`cpdID`	Compound ID of this spectrum.
`mode`	for 'reanalyzeFailpeak', the 'mode' (adduct) of the analyzed spectrum.

Details

`reanalyzeFailpeaks` examines the `unmatchedPeaksC` table in `specs` and sends every peak through `reanalyzeFailpeak`.

Value

The aggregate data frame extended by the columns: #'

`reanalyzed.???`	If reanalysis (step 7) has already been processed: matching values from the reanalyzed peaks
`matchedReanalysis`	Whether reanalysis has matched (`TRUE`), not matched(`FALSE`) or has not been conducted for the peak(`NA`).

It would be good to merge the analysis functions of `analyzeMsMs` with the one used here, to simplify code changes.

Author(s)

Michael Stravs

See Also

analyzeMsMs, msmsWorkflow

Examples

```
## As used in the workflow:
## Not run:
 reanalyzedRcSpecs <- reanalyzeFailpeaks(w@aggregated, custom_additions="N2O", mode="pH")
# A single peak:
reanalyzeFailpeak("N2O", 105.0447, 1234, 1, 1)

## End(Not run)
```

---

recalibrate *Predefined recalibration functions.*

Description

Predefined fits to use for recalibration: Loess fit and GAM fit.

Usage

```
recalibrate.loess(rcdata)

  recalibrate.identity(rcdata)

  recalibrate.mean(rcdata)

  recalibrate.linear(rcdata)
```

Arguments

rcdata	A data frame with at least the columns recalfield and mzFound. recalfield will usually contain delta(ppm) or delta(mz) values and is the target parameter for the recalibration.

Details

`recalibrate.loess()` provides a Loess fit (`recalibrate.loess`) to a given recalibration parameter. If MS and MS/MS data should be fit together, recalibrate.loess provides good default settings for Orbitrap instruments.

`recalibrate.identity()` returns a non-recalibration, i.e. a predictor which predicts 0 for all input values. This can be used if the user wants to skip recalibration in the RMassBank workflow.

#' `recalibrate.mean()` and `recalibrate.linear()` are simple recalibrations which return a constant shift or a linear recalibration. They will be only useful in particular cases.

`recalibrate()` itself is only a dummy function and does not do anything.

Alternatively other functions can be defined. Which functions are used for recalibration is specified by the RMassBank options file. (Note: if `recalibrateMS1: common`, the `recalibrator: MS1` value is irrelevant, since for a common curve generated with the function specified in `recalibrator: MS2` will be used.)

Value

Returns a model for recalibration to be used with `predict` and the like.

Author(s)

Michael Stravs, EAWAG <michael.stravs@eawag.ch>

Examples

```
## Not run:
rcdata <- subset(spec$peaksMatched, formulaCount==1)
ms1data <- recalibrate.addMS1data(spec, mode, 15)
rcdata <- rbind(rcdata, ms1data)
rcdata$recalfield <- rcdata$dppm
rcCurve <- recalibrate.loess(rcdata)
# define a spectrum and recalibrate it
s <- matrix(c(100,150,200,88.8887,95.0005,222.2223), ncol=2)
colnames(s) <- c("mz", "int")
recalS <- recalibrateSingleSpec(s, rcCurve)

Alternative: define an custom recalibrator function with different parameters
recalibrate.MyOwnLoess <- function(rcdata)
{
 return(loess(recalfield ~ mzFound, data=rcdata, family=c("symmetric"),
     degree = 2, span=0.4))
}
# This can then be specified in the RMassBank settings file:
# recalibrateMS1: common
# recalibrator:
#    MS1: recalibrate.loess
#    MS2: recalibrate.MyOwnLoess")
# [...]

## End(Not run)
```

recalibrate.addMS1data

Return MS1 peaks to be used for recalibration

Description

Returns the precursor peaks for all MS1 spectra in the spec dataset with annotated formula to be used in recalibration.

For all spectra in spec$specFound, the precursor ion is extracted from the MS1 precursor spectrum. All found ions are returned in a data frame with a format matching spec$peaksMatched and therefore suitable for rbinding to the spec$peaksMatched table. However, only minimal information needed for recalibration is returned.

Usage

```
recalibrate.addMS1data(spec, recalibrateMS1Window =
    getOption("RMassBank")$recalibrateMS1Window)
```

Arguments

spec	A msmsWorkspace or RmbSpectraSetList containing spectra for which MS1 "peaks" should be "constructed".
recalibrateMS1Window	Window width to look for MS1 peaks to recalibrate (in ppm).

Value

A dataframe with columns mzFound, formula, mzCalc, dppm, dbe, int,dppmBest, formulaCount, good, cpdID, scan, parentScan, dppmRc. However, columns dbe, int, formulaCount, good, scan, parentScan do not contain real information and are provided only as fillers.

Author(s)

Michael Stravs, EAWAG <michael.stravs@eawag.ch>

Examples

```
## Not run:
# More or less as used in recalibrateSpectra:
  rcdata <- peaksMatched(w)
  rcdata <- rcdata[rcdata$formulaCount == 1, ,drop=FALSE]
  ms1data <- recalibrate.addMS1data(w, "pH", 15)
  rcdata <- rbind(rcdata, ms1data)
 # ... continue constructing recalibration curve with rcdata

## End(Not run)
```

rmb_log_debug — *Pass arguments to logger::log_debug using custom RMassBank-logging settings*

Description

The logging file to be used can be specified by the user in the `logging_file` field of `settings.ini`

Usage

```
rmb_log_debug(...)
```

Arguments

...	The log message, as for 'logger::log_...' functions

Author(s)

pstahlhofen

See Also

logger::log_debug

rmb_log_error — *Pass arguments to logger::log_error using custom RMassBank-logging settings*

Description

The logging file to be used can be specified by the user in the `logging_file` field of `settings.ini`

Usage

```
rmb_log_error(...)
```

Arguments

...	The log message, as for 'logger::log_...' functions

Author(s)

pstahlhofen

See Also

logger::log_error

`rmb_log_fatal` *Pass arguments to logger::log_fatal using custom RMassBank-logging settings*

Description

The logging file to be used can be specified by the user in the `logging_file` field of `settings.ini`

Usage

```
rmb_log_fatal(...)
```

Arguments

...	The log message, as for 'logger::log_...' functions

Author(s)

pstahlhofen

See Also

logger::log_fatal

`rmb_log_info` *Pass arguments to logger::log_info using custom RMassBank-logging settings*

Description

The logging file to be used can be specified by the user in the `logging_file` field of `settings.ini`

Usage

```
rmb_log_info(...)
```

Arguments

...	The log message, as for 'logger::log_...' functions

Author(s)

pstahlhofen

See Also

logger::log_info

`rmb_log_success` *Pass arguments to logger::log_success using custom RMassBank-logging settings*

Description

The logging file to be used can be specified by the user in the `logging_file` field of `settings.ini`

Usage

```
rmb_log_success(...)
```

Arguments

...	The log message, as for 'logger::log_...' functions

Author(s)

pstahlhofen

See Also

logger::log_success

`rmb_log_trace` *Pass arguments to logger::log_trace using custom RMassBank-logging settings*

Description

The logging file to be used can be specified by the user in the `logging_file` field of `settings.ini`

Usage

```
rmb_log_trace(...)
```

Arguments

...	The log message, as for 'logger::log_...' functions

Author(s)

pstahlhofen

See Also

logger::log_trace

rmb_log_warn *Pass arguments to logger::log_warn using custom RMassBank-logging settings*

Description

The logging file to be used can be specified by the user in the `logging_file` field of `settings.ini`

Usage

```
rmb_log_warn(...)
```

Arguments

`...`	The log message, as for 'logger::log_...' functions

Author(s)

pstahlhofen

See Also

logger::log_warn

RmbDefaultSettings *RMassBank settings*

Description

Load, set and reset settings for RMassBank.

Usage

```
loadRmbSettings(file_or_list)

loadRmbSettingsFromEnv(env = .GlobalEnv)

RmbDefaultSettings()

RmbSettingsTemplate(target)
```

Arguments

`file_or_list`	The file (YML or R format) or R `list` with the settings to load.
`target`	The path where the template setting file should be stored.
`env`	The environment to load the settings from.

Details

RmbSettingsTemplate creates a template file in which you can adjust the settings as you like. Before using RMassBank, you must then load the settings file using loadRmbSettings. RmbDefaultSettings loads the default settings. loadRmbSettingsFromEnv loads the settings stored in env$RmbSettings, which is useful when reloading archives with saved settings inside.

Note: no settings are loaded upon loading MassBank! This is intended, so that one never forgets to load the correct settings.

The settings are described in RmbSettings.

Value

None.

Note

The default settings will not work for you unless you have, by chance, installed OpenBabel into the same directory as I have!

Author(s)

Michael Stravs

See Also

RmbSettings

Examples

```
# Create a standard settings file and load it (unedited)
RmbSettingsTemplate("mysettings.ini")
loadRmbSettings("mysettings.ini")
unlink("mysettings.ini")
```

RmbSettings *RMassBank settings*

Description

Describes all settings for the RMassBank settings file.

Details

- deprofile Whether and how to deprofile input raw files. Leave the setting empty if your raw files are already in "centroid" mode. If your input files are in profile mode, you have the choice between algorithms deprofile.spline, deprofile.fwhm, deprofile.localMax; refer to the individual manpages for more information.
- rtMargin, rtShift The allowed retention time deviation relative to the values specified in your compound list (see loadList), and the systematic shift (due to the use of, e.g., pre-columns or other special equipment.
- babeldir Directory to OpenBabel. Required for creating molfiles for MassBank export. If no OpenBabel directory is given, RMassBank will attempt to use the CACTUS webservice for SDF generation. It is strongly advised to install OpenBabel; the CACTUS structures have explicit hydrogen atoms. The path should point to the directory where babel.exe (or the Linux "babel" equivalent) lies.
- use_version Which MassBank record format to use; version 2 is strongly advised, version 1 is considered outdated and should be used only if for some reason you are running old servers and an upgrade is not feasible.
- use_rean_peaks Whether to include peaks from reanalysis (see reanalyzeFailpeaks) in the MassBank records. Boolean, TRUE or FALSE.
- annotations A list of constant annotations to use in the MassBank records. The entries authors, copyright, license, instrument, instrument_type, compound_class correspond to the MassBank entries AUTHORS, COPYRIGHT, PUBLICATION, LICENSE, AC$INSTRUMENT,AC$INSTRUMENT_TYP CH$COMPOUND_CLASS. The entry confidence_comment is added as COMMENT: CONFIDENCE entry.

  The entry internal_id_fieldname is used to name the MassBank entry which will keep a reference to the internal compound ID used in the workflow: for internal_id_fieldname = MYID and e.g. compound 1234, an entry will be added to the MassBank record with COMMENT: MYID 1234. The internal fieldname should not be left empty!

  The entries lc_gradient, lc_flow, lc_solvent_a, lc_solvent_b, lc_column correspond to the MassBank entries AC$CHROMATOGRAPHY: FLOW_GRADIENT, FLOW_RATE,SOLVENT A, SOLVENT B, COLUMN_NAME.

  ms_type, ionization correspond to AC$MASS_SPECTROMETRY: MS_TYPE, IONIZATION.

  entry_prefix is the two-letter prefix used when building MassBank accession codes.

  Entries under ms_dataprocessing are added as MS$DATA_PROCESSING: entries, in addition to the default WHOLE: RMassBank.
- annotator For advanced users: option to select your own custom annotator. Check annotator.default and the source code for details.
- spectraList This setting describes the experimental annotations for the single data-dependent scans. For every data-dependent scan event, a spectraList entry with mode, ces, ce, res denoting collision mode, collision energy in short and verbose notation, and FT resolution.
- accessionNumberShifts This denotes the starting points for accession numbers for different ion types. For example, pH: 0, mH: 50 means that [M+H]+ spectra will start at XX123401 (XX being the entry_prefix and 1234 the compound id) and [M-H]- will start at XX123451.
- electronicNoise, electronicNoiseWidth Known electronic noise peaks and the window to be used by cleanElnoise

- `recalibrateBy` `dppm` or `dmz` to recalibrate either by delta ppm or by delta mz.
- `recalibrateMS1` `common` or `separate` to recalibrate MS1 data points together or separately from MS2 data points.
- `recalibrator: MS1, MS2` The functions to use for recalibration of MS1 and MS2 data points. Note that the `MS1` setting is only meaningful if `recalibrateMS1: separate`, otherwise the `MS2` setting is used for a common recalibration curve. See `recalibrate.loess` for details.
- `multiplicityFilter` Define the multiplicity filtering level. Default is 2, a value of 1 is off (no filtering) and >2 is harsher filtering.
- `titleFormat` The title of MassBank records is a mini-summary of the record, for example "Dinotefuran; LC-ESI-QFT; MS2; CE: 35 By default, the first compound name `CH$NAME`, instrument type `AC$INSTRUMENT_TYPE`, MS/MS type `AC$MASS_SPECTROMETRY: MS_TYPE`, collision energy `RECORD_TITLE_CE`, resolution `AC$MASS_SPECTROMETRY: RESOLUTION` and precursor `MS$FOCUSED_ION: PRECURSOR_TYPE` are used. If alternative information is relevant to differentiate acquired spectra, the title should be adjusted. For example, many TOFs do not have a resolution setting. See MassBank documentation for more.
- `filterSettings` A list of settings that affect the MS/MS processing. The entries `ppmHighMass`, `ppmLowMass`, `massRangeDivision` set values for pre-processing, prior to recalibration. `ppmHighMass` defines the ppm error for the high mass range (default 10 ppm for Orbitraps), `ppmLowMass` is the error for the low mass range (default 15 ppm for Orbitraps) and `massRangeDivision` is the m/z value defining the split between the high and low mass range (default m/z = 120).

  The entry `ppmFine` defines the ppm cut-off post recalibration. The default value of 5 ppm is recommended for Orbitraps. For other instruments this can be interpreted from the recalibration plot. All ppm limits are one-sided (e.g. this includes values to +5 ppm or -5 ppm deviation from the exact mass).

  The entries `prelimCut`, `prelimCutRatio` define the intensity cut-off and cut-off ratio (in the peak selection for the recalibration only. Careful: the default value 1e4 for Orbitrap LTQ positive mode could remove all peaks for TOF data and will remove too many peaks for Orbitrap LTQ negative mode spectra!

  The entry `specOKLimit` defines the intensity limit to include MS/MS spectra. MS/MS spectra must have at least one peak above this limit to proceed through the workflow.

  `dbeMinLimit` defines the minimum allowable ring and double bond equivalents (DBE) allowed for assigned formulas. This assumes maximum valuences for elements with multiple valence states. The default is -0.5 (accounting for fragments being ions).

  The entries `satelliteMzLimit`, `satelliteIntLimit` define the cut-off m/z and intensity values for satellite peak removal (an artefact of Fourier Transform processing). All peaks within the m/z limit (default 0.5) and intensity ratio (default 0.05 or 5 Fourier Transform instruments only (e.g. Orbitrap).
- `filterSettings` Parameters for adjusting the raw data retrieval. The entry `ppmFine` defines the ppm error to look for the precursor in the MS1 (parent) spectrum. Default is 10 ppm for Orbitrap.

  `mzCoarse` defines the error to search for the precursor specification in the MS2 spectrum. This is often only saved to 2 decimal places and thus can be quite inaccurate. The accuracy also depends on the isolation window used. The default settings (for e.g. Orbitrap) is 0.5 (Da, or Th for m/z).

  The entry `fillPrecursorScan` is largely untested. The default value (FALSE) assumes all necessary precursor information is available in the mzML file. A setting ot TRUE tries to

fill in the precursor data scan number if it is missing. Only tested on one case study so far - feedback welcome!

Author(s)

Michael Stravs, Emma Schymanski

See Also

loadRmbSettings

RmbSpectraSet-class *Set of spectra pertaining to one compound*

Description

Set of spectra pertaining to one compound

Slots

`parent` Spectrum1 The precursor spectrum

`children` RmbSpectrum2List List of 'RmbSpectrum2' objects for the fragmentation spectra, which are first extracted and later processed during 'msmsWorkflow'

`found` logical, denotes whether or not fragmentation spectra were found for this compound

`complete` logical, denotes whether or not *all* expected collision energies were found for this compound

`empty` logical, 'TRUE' if there are zero found spectra for this compound

`formula` character, the molecular formula of the neutral compound

`id` The ID of the compound in the RMassBank compound list (see loadList)

`mz` the m/z value of the precursor

`name` The name of the compound

`mode` The ion type of the precursor, e.g. 'pH, mH, mNa'

`smiles` the SMILES string for the compound structure

RmbSpectrum2-class *RMassBank Representation of an MSMS Spectrum*

Description

This extends the Spectrum2 class of the MSnbase package and introduces further slots that are used to store information during the RMassBank workflow.

Slots

satellite logical If TRUE, the corresponding peak was removed as satellite.

low logical If TRUE, the corresponding peak was removed because it failed the intensity cutoff.

rawOK logical If TRUE, the peak passed satellite and low-intensity cutoff removal.

good logical If TRUE, a formula could be found for the peak and the peak passed all filter criteria. (see the RMassBank vignette or the documentation of analyzeMsMs#' for details on filter settings)

mzCalc numeric The mz value calculated from the found formula for each peak (if any)

formula character The formula found for each peak. generate.formula is used for formula-fitting

dbe numeric The number of double bond equivalents. This is calculated from the found formula for each peak (if any)

formulaCount integer The number of different formulae found for each peak. Note: A peak for which multiple formulas were found will appear multiple times. Hence there may be multiple entries in the formula , dppm and mzCalc slot for the same mz value.

formulaSource character "analyze" or "reanalysis" Shows whether the current formula for the peak was determined by normal analysis ("analyze") or by reanalysis of a failpeak ("reanalysis")

dppm numeric The ppm deviation of the mz value from the found formula (if any).

dppmBest numeric The ppm deviation of the mz value from the best formula found.

ok logical one-element vector If this is TRUE, the spectrum was successfully processed with at least one resulting peak. Otherwise, one of the following cases applies:

- All peaks failed the intensity cutoff i.e. the whole spectrum contains low intensity peaks, only.
- All peaks were marked as satellites.
- All peaks in the spectrum have a lower intensity than the value given in the specOkLimit filter setting. (see the RMassBank vignette or the documentation of analyzeMsMs)
- The precursor ion formula is invalid (see is.valid.formula)
- The spectrum is empty.
- No molecular formula could be found for any of the peaks.
- All peaks failed the dbeMinLimit criterion. (see the RMassBank vignette or the documentation of analyzeMsMs)

`info` list Spectrum identifying information (collision energy, resolution, collision mode) from the `spectraList`

`properties` data.frame This is used as a flexible placeholder to store additional properties for each peak throughout the workflow. After the last step of the `mbWorkflow`, this will typically contain columns `mzRaw`, `noise`, `formulaMultiplicity`, `bestMultiplicity` and `filterOK`. However, new columns may be added on demand (see `property<-`)

See Also

`generate.formula`, `property<-` `analyzeMsMs`, `generate.formula`, `is.valid.formula`

`RmbSpectrum2List-class` *SimpleList specializations*

Description

Typed lists using SimpleList

`selectPeaks` *Select peaks from aggregate table*

Description

Selects peaks from aggregate table according to different criteria.

Usage

```
selectPeaks(o, ...)

## S4 method for signature 'RmbSpectrum2'
selectPeaks(o, filter, ..., enclos = parent.frame(2))

## S4 method for signature 'Spectrum'
selectPeaks(o, filter, ..., enclos = parent.frame(2))

## S4 method for signature 'RmbSpectrum2List'
selectPeaks(o, ..., enclos = parent.frame(2))

## S4 method for signature 'RmbSpectraSetList'
selectPeaks(o, ..., enclos = parent.frame(2))

## S4 method for signature 'RmbSpectraSet'
selectPeaks(o, ..., enclos = parent.frame(2))
```

```
## S4 method for signature 'data.frame'
selectPeaks(o, filter, ..., enclos = parent.frame(2))

## S4 method for signature 'msmsWorkspace'
selectPeaks(o, ..., enclos = parent.frame(2))
```

Arguments

o	msmsWorkspace or aggregate data.frame from a workspace.
...	no additional parameters
filter	The expression (to be evaluated in context of the 'getData()' result on the spectrum) to define the peaks to keep. For example, 'good & filterOK'
enclos	The context in which to evaluate the filter expression, by default set such that the spectrum 'getData()' is retrieved.

Value

Peak dataframe according to the specified criteria.

Methods (by class)

- selectPeaks(RmbSpectraSetList): A method to filter spectra to the specified peaks
- selectPeaks(RmbSpectraSet): A method to filter spectra to the specified peaks
- selectPeaks(data.frame): A method to retrieve the specified peaks from the "aggregated" slot (a data.frame object) in an msmsWorkSpace
- selectPeaks(msmsWorkspace): A method to retrieve the specified peaks from an msmsWorkSpace

Author(s)

stravsmi

---

selectSpectra *Select a subset of spectra matching properties*

Description

From a list of RmbSpectraSets, returns the spectra which match a criterion (found, complete, empty as in checkSpectra). This can be returned either as a TRUE/FALSE vector, as a vector of indices for matching elements, as a vector of RmbSpectraSet objects matching the conditions, or as a vector of RmbSpectraSet objects NOT matching the conditions (sic!).

Usage

```
selectSpectra(s, property, value = "logical")

## S4 method for signature 'RmbSpectraSetList,character'
selectSpectra(s, property, value = "logical")

## S4 method for signature 'msmsWorkspace,character'
selectSpectra(s, property, value = "logical")
```

Arguments

s	The RmbSpectraSetList or msmsWorkspace to select RmbSpectraSets from.
property	The property to check (found, complete or empty)
value	logical if a TRUE/FALSE list should be returned; index if a vector of matching indices should be returned, object if matching objects should be returned, mismatch if mismatching objects should be returned.

Value

As described above.

Methods (by class)

- selectSpectra(s = RmbSpectraSetList, property = character): A method for selecting spectra from a spectra set list
- selectSpectra(s = msmsWorkspace, property = character): A method for selecting spectra from an msmsWorkspace

Author(s)

stravsmi

---

setAccessionBuilder *Define a programmatic or gluey ACCESSION builder*

Description

Define a programmatic or gluey ACCESSION builder

Usage

```
setAccessionBuilder(accessionBuilder)
```

Arguments

accessionBuilder
: a function that takes parameters ‘cpd‘ (an instance of ‘RmbSpectraSet‘), ‘spectrum‘ (an instance of ‘RmbSpectrum2‘) and ‘subscan‘ (an integer denoting relative scan id) and returns a ‘character‘. Alternatively a glue string just like the one in the RMassBank settings.

---

setData *Set* RmbSpectrum2 *data from data.frame*

Description

Sets all slots which are present as columns in the given dataframe. Optionally cleans the object, i.e. empties slots not defined in the data frame.

Usage

```
## S4 method for signature 'RmbSpectrum2,data.frame'
setData(s, df, clean = TRUE)
```

Arguments

s	The RmbSpectrum2 object to modify
df	The data frame with new data
clean	TRUE if slots which aren't present as columns in the data frame should be cleared.

Value

The modified RmbSpectrum2.

Author(s)

stravsmi

---

smiles2mass *Calculate the mass from a SMILES-String*

Description

Uses a SMILES-String to calculate the mass using rcdk-integrated functions.

Usage

```
smiles2mass(SMILES)
```

Arguments

SMILES	A String-object representing a SMILES

Value

The calculated mass of the given SMILES-Formula

Author(s)

Erik Mueller

Examples

```
## Not run:
 smiles2mass("CC(=O)NC(C(O)1)C(O)C(OC(O2)C(O)C(OC(O3)C(O)C(O)C(O)C(CO)3)C(O)C(CO)2)C(CO)O1")

## End(Not run)
```

---

spectraCount *Count MS2 spectra per compound*

Description

Counts the number of acquired spectra for a compound or multiple compounds

Usage

```
spectraCount(s)

## S4 method for signature 'RmbSpectraSet'
spectraCount(s)

## S4 method for signature 'RmbSpectraSetList'
spectraCount(s)

## S4 method for signature 'msmsWorkspace'
spectraCount(s)
```

Arguments

s	The object (RmbSpectraSet, RmbSpectraSetList or msmsWorkspace) to count the spectra in.

Value

For RmbSpectraSet objects, a single number counting the spectra in that object. For RmbSpectraSetList or msmsWorkspace, a vector with spectra counts for all compounds (RmbSpectraSets) in the object.

Methods (by class)

- `spectraCount(RmbSpectraSet)`: Counts the number of acquired spectra for an RmbSpectraSet
- `spectraCount(RmbSpectraSetList)`: Counts the number of acquired spectra for an RmbSpectraSetList
- `spectraCount(msmsWorkspace)`: Counts the number of acquired spectra for an msmsWorkSpace

Author(s)

stravsmi

`to.limits.rcdk` *Convert formula to Rcdk limits*

Description

Converts a molecular formula e.g. C15H20 into an upper limit appropriate for use with Rcdk's `generate.formula` function's `element` argument.

Usage

```
to.limits.rcdk(formula)
```

Arguments

`formula`	A molecular formula in string or list representation (`"C6H6"` or `list(C=6,H=6)`).

Details

This helper function is used to make the upper limits for `generate.formula` when finding subformulas to match to a MS2 fragment peak.

Value

An array in the form `c( c("C", "0", "12"), c("H", "0", "12"))` (for input of "C12H12").

Author(s)

Michael Stravs

See Also

`generate.formula`, `add.formula`

Examples

```
#
to.limits.rcdk("C6H6")
to.limits.rcdk(add.formula("C6H12O6", "H"))
```

`toMassbank` *Write MassBank record into character array*

Description

Writes a MassBank record in list format to a text array.

Usage

```
toMassbank(o, ...)

## S4 method for signature 'RmbSpectraSet'
toMassbank(o, addAnnotation = getOption("RMassBank")$add_annotation)

## S4 method for signature 'RmbSpectrum2'
toMassbank(o, addAnnotation = getOption("RMassBank")$add_annotation)
```

Arguments

`o`	An object to convert to MassBank record format. This may be a single 'RmbSpectrum2', or a complete compound (an 'RmbSpectraSet'),
`...`	Parameters passed to the implementation, in particular 'addAnnotation'
`addAnnotation`	'logical', whether to add peak annotations (putative formulas) to the record.

Details

The function is a general conversion tool for the MassBank format; i.e. the field names are not fixed. `mbdata` must be a named list, and the entries can be as follows:

- A single text line:
  `'CH\$EXACT_MASS' = '329.1023'`
  is written as
  `CH\$EXACT_MASS: 329.1023`
- A character array:
  `'CH\$NAME' = c('2-Aminobenzimidazole', '1H-Benzimidazol-2-amine')`
  is written as
  `CH\$NAME: 2-Aminobenzimidazole`
  `CH\$NAME: 1H-Benzimidazol-2-amine`

- A named list of strings:

  `'CH\$LINK' = list('CHEBI' = "27822", "KEGG" = "C10901")`

  is written as

  `CH\$LINK: CHEBI 27822`

  `CH\$LINK: KEGG C10901`

- A data frame (e.g. the peak table) is written as specified in the MassBank record format (Section 2.6.3): the column names are used as headers for the first line, all data rows are printed space-separated.

Value

The result is a text array, which is ready to be written to the disk as a file.

Note

The function iterates over the list item names. **This means that duplicate entries in** `mbdata` **are (partially) discarded!** The correct way to add them is by making a character array (as specified above): Instead of `'CH\$NAME' = 'bla', 'CH\$NAME' = 'blub'` specify `'CH\$NAME' = c('bla','blub')`.

Author(s)

Michael Stravs

References

MassBank record format: http://www.massbank.jp/manuals/MassBankRecord_en.pdf

See Also

buildRecord, mbWorkflow

Examples

```
## Not run:
# Read just the compound info skeleton from the Internet for some compound ID
id <- 35
mbdata <- gatherData(id)
#' # Export the mbdata blocks to line arrays
# (there is no spectrum information, just the compound info...)
mbtext <- toMassbank(mbdata)

## End(Not run)
```

toRMB *Conversion of XCMS-pseudospectra into RMassBank-spectra*

Description

Converts a pseudospectrum extracted from XCMS using CAMERA into the msmsWorkspace(at)spectrum-format that RMassBank uses

Usage

```
toRMB(msmsXCMSspecs, cpdID, mode, MS1spec)
```

Arguments

msmsXCMSspecs	The compoundID of the compound that has been used for the peaklist
cpdID	The compound ID of the substance of the given spectrum
mode	The ionization mode that has been used for the spectrum
MS1spec	The MS1-spectrum from XCMS, which can be optionally supplied

Value

One list element of the (at)specs-entry from an msmsWorkspace

Author(s)

Erik Mueller

See Also

msmsWorkspace-class

Examples

```
## Not run:
  XCMSpspectra <- findmsmsHRperxcms.direct("Glucolesquerellin_2184_1.mzdata", 2184)
     wspecs <- toRMB(XCMSpspectra)

## End(Not run)
```

updateHeader *Add a header to a Multiblock JCAMP file*

Description

JCAMP files containing multiple blocks are usually structured by so-called link blocks. If no link block is present, the readJDX package is not able to parse the file. This method will add a link block at the top of the given file or print a message if an existing link block is found. The file is not changed in this case.

Usage

```
updateHeader(filename)
```

Arguments

filename	character The name of the file to which a link block should be added. The filename is also used as content for the TITLE field in the link block

Value

Nothing is returned

Author(s)

pstahlhofen

Examples

```
## Not run:
   updateHeader("my_multiblock_jcamp.jdx")

## End(Not run)
```

updateSettings *Update settings to current version*

Description

Checks if all necessary fields are present in the current settings and fills in default values from the RmbDefaultSettings if required.

Usage

```
updateSettings(settings, warn = TRUE)
```

Arguments

settings	The set of settings to check and update.
warn	Whether to update parameters quietly (FALSE) or to notify the user of the changed parameters (TRUE, default.) This serves to make the user aware that standard parameters are filled in!

Value

The updated set of settings.

Note

Important: There is a change in behaviour of RMassBank in certain cases when filterSettings is not present in the old settings! The default pre-recalibration cutoff from RmbDefaultSettings is 10000. Formerly the pre-recalibration cutoff was set to be 10000 for positive spectra but 0 for negative spectra.

Updating the settings files is preferred to using the updateSettings function.

Author(s)

Stravs MA, Eawag <michael.stravs@eawag.ch>

Examples

```
## Not run:
w@settings <- updateSettings(w@settings)

## End(Not run)
```

---

validate *Validate MassBank records with a set of Unit tests*

Description

Validates a plain text MassBank record, or recursively all records within a directory. The Unit Tests to be used are installed in RMassBank/inst/validationTests and currently include checks for NAs, peaks versus precursor, precursor mz, precursor type, SMILES vs exact mass, total intensities and title versus type. The validation report is saved as "report.html" in the working directory.

Usage

```
validate(path, simple = TRUE)
```

Arguments

path	The filepath to a single record, or a directory to search recursively
simple	If TRUE the function creates a simpler form of the RUnit .html report, better readable for humans. If FALSE it returns the unchanged RUnit report.

Examples

```
## Not run:
validate("/tmp/MassBank/OpenData/record/")

## End(Not run)
```

Index

+,RmbSpectraSet,ANY-method, 8
+,RmbSpectrum2List,ANY-method, 8
+,Spectrum,numeric-method, 9
-,RmbSpectraSet,ANY-method, 5
-,RmbSpectrum2List,ANY-method, 5
-,Spectrum,numeric-method, 6
.RmbSpectraSet *(RmbSpectraSet-class)*, 113
.RmbSpectraSetList *(RmbSpectrum2List-class)*, 115
.RmbSpectrum2 *(RmbSpectrum2-class)*, 114
.RmbSpectrum2List *(RmbSpectrum2List-class)*, 115
.msmsWorkspace *(msmsWorkspace-class)*, 88
.parseTitleString, 6
.updateObject.RmbSpectrum2.formulaSource, 7

add.formula, 9, *55*, *73*, *91*, *120*
addMB, 10
addPeaks, 11, *20*, *81*
addPeaksManually, *10*, 12
addProperty, 13
addProperty,data.frame,character,character-method *(addProperty)*, 13
addProperty,RmbSpectrum2,character,character-method *(addProperty)*, 13
aggregateSpectra, 14, *24*
analyzeMsMs, 15, *15*, *19*, *20*, *36*, *38–40*, *87*, *102*, *103*, *114*, *115*
annotator.default, 18, *111*
archiveResults, 19, *90*

buildRecord, *7*, 19, *122*
buildRecord,RmbSpectraSet-method *(buildRecord)*, 19
buildRecord,RmbSpectrum2-method *(buildRecord)*, 19

CAS2SMILES, 21
checkIsotopes, 21
checkSpectra, 23, *116*
checkSpectra,RmbSpectraSet,character-method *(checkSpectra)*, 23
cleanElnoise, 24, *111*
cleanElnoise,data.frame,numeric,numeric-method *(cleanElnoise)*, 24
cleanElnoise,RmbSpectraSet,numeric,numeric-method *(cleanElnoise)*, 24
cleanElnoise,RmbSpectrum2,numeric,numeric-method *(cleanElnoise)*, 24
cleanElnoise,RmbSpectrum2List,numeric,numeric-method *(cleanElnoise)*, 24
combineMultiplicities, 25
compoundlist2SDF, 26
createCompoundlist, 27
createMolfile, 28, *33*, *34*
CTS.externalIdSubset, 29
CTS.externalIdTypes, 30

dbe, 30
deprofile, 31, *44*, *47*, *111*

exportMassbank, 33

fillback, 34
fillback,msmsWorkspace,missing,missing-method *(fillback)*, 34
fillback,RmbSpectraSet,missing,data.frame-method *(fillback)*, 34
fillback,RmbSpectrum2,character,data.frame-method *(fillback)*, 34
filterCompoundlist, 35
filterLowaccResults, *18*, 36, *39*
filterMultiplicity, 37
filterPeakSatellites, *16–18*, *36*, 38
filterPeaksMultiplicity, *37*, *38*, 39
findCAS *(findMz)*, 50
findEIC, 40
findFormula *(findMz)*, 50

findLevel (findMz), 50
findMass, 42, *51*
findMsMsHR, *16*, 43, *46*, *48*, *76*, *78*
findMsMsHR.direct, 46
findMsMsHR.ticMS2 (findMsHR.ticms2), 47
findMsMsHR.ticms2, 47
findMsMsHRperMsp, 48
findMsMsHRperxcms, 49
findMz, *42*, 50, *52*, *75*
findMz.formula, *51*, 52
findName (findMz), 50
findProgress, 53
findRt (findMz), 50
findSmiles, *28*
findSmiles (findMz), 50
flatten, 53
formulastring.to.list, *10*, *52*, 55

gatherCCTE, 56
gatherData, *53, 54*, 57
gatherDataBabel, 58
gatherDataUnknown, 59
gatherPubChem, 60
generate.formula, *114, 115*, *120*
getAnalyticalInfo, 61
getCactus, 61, *71*
getCASRN, 62
getCSID, 63
getCtsKey, 64
getCtsRecord, *29, 30*, *62*, 65, *71*
getData, 66, *82, 83*
getData,RmbSpectrum2-method (getData), 66
getDTXCID, 66
getDTXSID, 67
getDTXSMILES, 68
getField, 69
getMolecule, 70
getPcId, *62*, 71
getPrefName, 72

infolist, *53*
is.valid.formula, *10*, *55*, 73, *91*, *114, 115*

list.to.formula, *73*, *91*
list.to.formula (formulastring.to.list), 55
loadInfolist, *19*, *54*
loadInfolist (loadInfolists), 74
loadInfolists, 74, *81*
loadList, *43, 44*, *51*, 75, *111*, *113*
loadMsmsWorkspace (newMsmsWorkspace), 89
loadRmbSettings, *16*, *22*, *84, 85*, *87*, *113*
loadRmbSettings (RmbDefaultSettings), 109
loadRmbSettingsFromEnv (RmbDefaultSettings), 109

makeMollist, 76
makePeaksCache, 76
makeRecalibration, 77
mbWorkflow, *11*, *20*, *34*, *37*, *42*, *56–58*, *60, 61*, *74*, 79, *81*, *89*, *122*
mbWorkspace-class, 80
mergePeaks, 81
mergePeaks,data.frame-method (mergePeaks), 81
mergePeaks,matrix-method (mergePeaks), 81
mergePeaks,RmbSpectrum2-method (mergePeaks), 81
mergePeaks,Spectrum-method (mergePeaks), 81
mergeSpectra, 82
mergeSpectra,RmbSpectrum2List-method (mergeSpectra), 82
msmsRead, 83, *87*
msmsRead.RAW, 85
msmsWorkflow, *12*, *15*, *18*, *25, 26*, *50*, *79*, *85*, 86, *86*, *88*, *90*, *98*, *103*
msmsWorkspace-class, 88
multiply.formula (add.formula), 9

newMbWorkspace, 89
newMsmsWorkspace, 89
normalize, *20*
normalize,RmbSpectrum2-method, 90
normalize,RmbSpectrum2List-method, 91

order.formula, *10*, *55*, *73*, 91

parse.smiles, *70*
parseMassBank, 92
parseMbRecord, 93
peaksMatched, 94
peaksMatched,data.frame-method (peaksMatched), 94

peaksMatched,msmsWorkspace-method
(peaksMatched), 94
peaksUnmatched, 94
peaksUnmatched,data.frame-method
(peaksUnmatched), 94
peaksUnmatched,msmsWorkspace-method
(peaksUnmatched), 94
plotMbWorkspaces, 95
plotRecalibration, 96
ppm, *82*, 97
problematicPeaks, *37, 38*, 98
processProblematicPeaks, 99
progressBarHook, *84, 85*, *87*, 99, *102*
property, 100
property,RmbSpectrum2,character-method
(property), 100
property<-, 101
property<-,RmbSpectrum2,character,logical,character-method
(property<-), 101
property<-,RmbSpectrum2,character,logical,missing-method
(property<-), 101
property<-,RmbSpectrum2,character,missing,character-method
(property<-), 101
property<-,RmbSpectrum2,character,missing,missing-method
(property<-), 101

readMbdata, *19*
readMbdata *(*flatten*)*, 53
reanalyzeFailpeak, *40*
reanalyzeFailpeak *(*reanalyzeFailpeaks*)*,
102
reanalyzeFailpeaks, *18*, 102, *111*
recalibrate, *77, 78*, 103
recalibrate.addMS1data, 105
recalibrate.loess, *112*
recalibrateSingleSpec
(makeRecalibration), 77
recalibrateSpectra *(*makeRecalibration*)*,
77
resetInfolists *(*loadInfolists*)*, 74
resetList *(*loadList*)*, 75
rmb_log_debug, 106
rmb_log_error, 106
rmb_log_fatal, 107
rmb_log_info, 107
rmb_log_success, 108
rmb_log_trace, 108
rmb_log_warn, 109
RmbDefaultSettings, 109, *124, 125*
RmbSettings, 110, *110*
RmbSettingsTemplate
(RmbDefaultSettings), 109
RmbSpectraSet-class, 113
RmbSpectraSetList
(RmbSpectrum2List-class), 115
RmbSpectraSetList-class
(RmbSpectrum2List-class), 115
RmbSpectrum2-class, 114
RmbSpectrum2List-class, 115

selectPeaks, *20*, 115
selectPeaks,data.frame-method
(selectPeaks), 115
selectPeaks,msmsWorkspace-method
(selectPeaks), 115
selectPeaks,RmbSpectraSet-method
(selectPeaks), 115
selectPeaks,RmbSpectraSetList-method
(selectPeaks), 115
selectPeaks,RmbSpectrum2-method
(selectPeaks), 115
selectPeaks,RmbSpectrum2List-method
(selectPeaks), 115
selectPeaks,Spectrum-method
(selectPeaks), 115
selectSpectra, 116
selectSpectra,msmsWorkspace,character-method
(selectSpectra), 116
selectSpectra,RmbSpectraSetList,character-method
(selectSpectra), 116
setAccessionBuilder, 117
setData, *82, 83*, 118
setData,RmbSpectrum2,data.frame-method
(setData), 118
show,mbWorkspace-method
(mbWorkspace-class), 80
show,msmsWorkspace-method
(msmsWorkspace-class), 88
smiles2mass, 118
spectraCount, 119
spectraCount,msmsWorkspace-method
(spectraCount), 119
spectraCount,RmbSpectraSet-method
(spectraCount), 119
spectraCount,RmbSpectraSetList-method
(spectraCount), 119

to.limits.rcdk, 120

toMassbank, *20*, *34*, 121
toMassbank,RmbSpectraSet-method
(toMassbank), 121
toMassbank,RmbSpectrum2-method
(toMassbank), 121
toRMB, *50*, 123

updateHeader, 124
updateSettings, 124

validate, *92*, *93*, 125