Package: DEScan2 (via r-universe)

October 30, 2026

Type Package

Title Differential Enrichment Scan 2

Version 1.33.0

Date 2025-07-31

Maintainer Dario Righelli <dario.righelli@gmail.com>

Description Integrated peak and differential caller, specifically designed for broad epigenomic signals.

Encoding UTF-8

License Artistic-2.0

LazyData TRUE

biocViews ImmunoOncology, PeakDetection, Epigenetics, Software, Sequencing, Coverage

Depends R (>= 3.5), GenomicRanges

Imports BiocParallel, BiocGenerics, ChIPpeakAnno, data.table, DelayedArray, Seqinfo, GenomeInfoDb, GenomicAlignments, glue, IRanges, plyr, Rcpp (>= 0.12.13), rtracklayer, S4Vectors (>= 0.23.19), SummarizedExperiment, tools, utils

LinkingTo Rcpp, RcppArmadillo

RoxygenNote 7.3.2

Suggests BiocStyle, knitr, rmarkdown, testthat, edgeR, limma, EDASeq, RUVSeq, RColorBrewer, statmod

VignetteBuilder knitr

Config/pak/sysreqs make libbz2-dev libicu-dev liblzma-dev libpng-dev libxml2-dev libssl-dev xz-utils zlib1g-dev

Repository https://bioc.r-universe.dev

Date/Publication 2026-04-28 12:47:27 UTC

RemoteUrl https://github.com/bioc/DEScan2

RemoteRef HEAD

RemoteSha 47c99148d91e1565c14901476f1c592c8d15f830

Contents

binnedCoverage . . . 2
computeZ . . . 3
constructBedRanges . . . 4
countFinalRegions . . . 5
createGranges . . . 6
cutGRangesPerChromosome . . . 7
DEScan2 . . . 8
divideEachSampleByChromosomes . . . 8
finalRegions . . . 9
findOverlapsOverSamples . . . 10
findPeaks . . . 11
fromSamplesToChrsGRangesList . . . 13
keepRelevantChrs . . . 13
readBamAsBed . . . 14
readBedFile . . . 15
readFilesAsGRangesList . . . 15
RleListToRleMatrix . . . 16
saveGRangesAsBed . . . 17
saveGRangesAsTsv . . . 18
setGRGenomeInfo . . . 19

Index **20**

`binnedCoverage` *binnedCoverage*

Description

this function computes the coverage over a binned chromosome, starting from a per base computed coverage.

Usage

```
binnedCoverage(
  bins,
  numvar,
  mcolname,
  covMethod = c("max", "mean", "sum", "min"),
  roundingMethod = c("none", "floor", "ceiling", "round")
)
```

Arguments

bins	a GRanges object representing a chromosome binned.
numvar	an RleList representing the per base coverage over the chr.
mcolname	the name of column where the sum have to be stored.
covMethod	a method to apply for the computing of the coverate it can be one of "max", "mean", "sum", "min". ("max" is default)
roundingMethod	a method to apply to round the computations it can be one of "none", "floor", "ceiling", "round". It's useful only when using covMethod="mean". ("none" is default)

Value

the bins GRanges with the mcolname attached

Examples

```
## dividing one chromosome in bins of 50 bp each
seqinfo <- Seqinfo::Seqinfo(genome="mm9")
bins <- GenomicRanges::tileGenome(
            seqlengths=Seqinfo::seqlengths(seqinfo)[1],
            tilewidth=50,
            cut.last.tile.in.chrom=TRUE)
gr <- GenomicRanges::GRanges(seqnames = S4Vectors::Rle("chr1", 100),
            ranges=IRanges::IRanges(start = seq(from=10, to=1000, by=10),
            end=seq(from=20, to=1010, by = 10)))
cov <- GenomicRanges::coverage(x=gr)
(binnedMaxCovGR <- binnedCoverage(bins, cov, "binned_cov"))
(binnedMeanCovGR <- binnedCoverage(bins, cov, "binned_cov",
                                covMethod="mean", roundingMethod="floor"))
(binnedSumCovGR <- binnedCoverage(bins, cov, "binned_cov", covMethod="sum"))
```

---

computeZ *computeZ*

Description

Computes Z-Scores returning the z matrix.

Usage

```
computeZ(
  lambdaChrRleList,
  runWinRleList,
  chrLength,
  minCount = 0.1,
  binSize = 50,
  verbose = FALSE
)
```

Arguments

lambdaChrRleList	an RleList of lambda values computed by computeLambdaOnChr function each element of the list is an Rle representing the lambda for the moving window in the list position.
runWinRleList	an RleList of coverage values computed. by computeCoverageMovingWindowOnChr function each element of the list is an Rle representing the coverage for the moving window in the list position.
chrLength	the length of the chr in analysis.
minCount	A small constant (usually no larger than one) to be added to the counts prior to the log transformation to avoid problems with log(0).
binSize	the size of the bin.
verbose	verbose output.

Value

z a matrix of z scores for each window (column) and bin (row). where the rownames represent the starting base of each bin.

---

constructBedRanges *constructBedRanges*

Description

Constructs a GRanges object from a bam/bed/bed.zip file in a consistent way.

Usage

```
constructBedRanges(
  filename,
  filetype = c("bam", "bed", "bed.zip", "narrow", "broad"),
  genomeName = NULL,
  onlyStdChrs = FALSE,
  arePeaks = FALSE,
  verbose = FALSE
)
```

Arguments

filename	the complete file path of a bam?bed file.
filetype	the file type bam/bed/bed.zip/narrow/broad.
genomeName	the name of the genome used to map the reads (i.e. "mm9"). N.B. if NOT NULL the GRanges Seqinfo will be forced to genomeName Seqinfo (needs Internet access, but strongly suggested!)
onlyStdChrs	flag to keep only standard chromosome.
arePeaks	flag indicating if the file contains peaks.
verbose	flag to obtain verbose output.

Value

a GRanges object.

Examples

```
files <- list.files(system.file("extdata/bam/", package="DEScan2"),
                    pattern="bam$", full.names=TRUE)
bgr <- constructBedRanges(files[1], filetype="bam", genomeName="mm9",
                          onlyStdChrs=TRUE)
bgr
```

`countFinalRegions` *countFinalRegions*

Description

count reads falling within the final regions.

Usage

```
countFinalRegions(
  regionsGRanges,
  readsFilePath = NULL,
  fileType = c("bam", "bed"),
  minCarriers = 2,
  genomeName = NULL,
  onlyStdChrs = FALSE,
  carrierscolname = "k-carriers",
  ignStrandSO = TRUE,
  modeSO = "Union",
  saveFlag = FALSE,
  savePath = "finalRegions",
  verbose = TRUE
)
```

Arguments

`regionsGRanges`	a GRanges objects representing the peaks to compute the coverage, with a "k-carriers" mcols. (tipically generated by finalRegions function).
`readsFilePath`	the filepath of bam or bed files necessary to compute the coverage.
`fileType`	the file type of the input files.
`minCarriers`	minimum number of carriers (samples).
`genomeName`	code name of the genome of reads files (i.e. "mm9").
`onlyStdChrs`	a flag indicating if to keep only the standard chromosomes

carrierscolname
: character describing the name of the column within the carriers number (default is "k-carriers").

ignStrandSO
: a flag indicating if to ignore the reads strand. (see GenomicAlignments::summarizeOverlaps).

modeSO
: the mode to use, default is "Union". (see GenomicAlignments::summarizeOverlaps).

saveFlag
: a flag indicating if to save the results.

savePath
: the path where to store the results.

verbose
: verbose output.

Value

A SummarizedExperiment object containing as assays the read counts matrix with regions as rows and samples as columns, and as rowRanges the GRanges object representing the peaks used as rows in the matrix.

Examples

```
filename <- system.file("extdata/regions/regions.rds", package="DEScan2")
regionsGR <- readRDS(file=filename)
reads.path <- system.file("extdata/bam", package="DEScan2")
finalRegionsSE <- countFinalRegions(regionsGRanges=regionsGR,
    readsFilePath=reads.path, fileType="bam", minCarriers=1,
    genomeName="mm9", onlyStdChrs=TRUE, ignStrandSO=TRUE, saveFlag=FALSE,
    verbose=TRUE)
library("SummarizedExperiment")
assay(finalRegionsSE) ## matrix of counts
rowRanges(finalRegionsSE) ## the GRanges of the input regions
```

---

createGranges *createGranges*

Description

a simplified wrapper function to create a GRanges object.

Usage

```
createGranges(chrSeqInfo, starts, widths, mcolname = NULL, mcolvalues = NULL)
```

Arguments

chrSeqInfo
: a seqinfo object.

starts
: the start ranges.

widths
: the width of each range.

mcolname
: the name for the mcol attribute.

mcolvalues
: the values for the mcol attribute.

Value

a GRanges object.

Examples

```
chrSeqInfo <- Seqinfo::Seqinfo(genome="mm9")["chr1"]
starts=sample(seq_len(100), 10)
widths=starts+10;
mcolname <- "z-score";
mcolvalues <- sample(seq_len(100), 10)
chrGR <- createGranges(chrSeqInfo=chrSeqInfo, starts=starts, widths=widths,
              mcolname=mcolname, mcolvalues=mcolvalues)
```

---

cutGRangesPerChromosome *cutGRangesPerChromosome*

Description

takes in input a GRanges object, producing a LIST of GRanges, one for each chromosome.

Usage

```
cutGRangesPerChromosome(GRanges)
```

Arguments

GRanges	a GRanges object.

Value

a named list of GRanges, one for each chromosome.

Examples

```
library("GenomicRanges")
gr <- GRanges(
        seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
        ranges=IRanges(1:10, end=10),
        strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
        seqlengths=c(chr1=11, chr2=12, chr3=13))
(grchrlist <- cutGRangesPerChromosome(gr))
```

DEScan2 *DEScan2*

Description

integrated peak and differential caller, specifically designed for broad epigenomic signals.

Author(s)

Dario Righelli

---

divideEachSampleByChromosomes *divideEachSampleByChromosomes*

Description

taken in input a grangeslist of samples, generate a list of samples where each element has a GRanges-List each element of the GRangesList represents a single chromosome.

Usage

```
divideEachSampleByChromosomes(samplesGRangesList)
```

Arguments

samplesGRangesList	a GRangesList of samples.

Value

list of samples where each element is a list of chromosomes and each of these elements is a GRanges.

Examples

```
library("GenomicRanges")
gr1 <- GRanges(
            seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
            ranges=IRanges(1:10, end=10),
            strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
            seqlengths=c(chr1=11, chr2=12, chr3=13))
gr2 <- GRanges(
            seqnames=Rle(c("chr1", "chr4", "chr1", "chr3"), c(1, 3, 2, 4)),
            ranges=IRanges(1:10, end=10),
            strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
            seqlengths=c(chr1=11, chr4=12, chr3=13))
```

```
sgrl <- GRangesList(gr1, gr2)
names(sgrl) <- c("samp1", "samp2")
(sampChrGrl <- divideEachSampleByChromosomes(sgrl))
```

---

finalRegions *finalRegions*

Description

Align peaks to form common regions then filter regions for presence in multiple replicates taking in input a GRangesList where each element is a sample of called peaks.

Usage

```
finalRegions(
  peakSamplesGRangesList,
  zThreshold = 20,
  minCarriers = 2,
  saveFlag = TRUE,
  outputFolder = "overlappedPeaks",
  verbose = FALSE,
  scorecolname = "z-score",
  coverageFlag = FALSE,
  BPPARAM = BiocParallel::bpparam()
)
```

Arguments

peakSamplesGRangesList	named GRangesList where each element is a sample of called peaks. A score mcols values is needed for each GRanges. The scorecolname param can be used as reference name for the score. (tipically returned by findPeaks function).
zThreshold	a minimum threshold for the z score. All peaks lesser than this value will be ignored.
minCarriers	a threshold of minimum samples (carriers) for overlapped regions.
saveFlag	a flag for saving results in a tsv file.
outputFolder	the directory name to store the bed file.
verbose	verbose output.
scorecolname	character describing the name of the column within the peaks score.
coverageFlag	boolean indicating if to compute the scores in a coverage mode (sum of the reads of merged peak) or in a score mode (a normalized score across the merged peaks)
BPPARAM	object of class bpparamClass that specifies the back-end to be used for computations. See bpparam for details.

Value

a GRanges of selected overlapping peaks with z-score, n-peaks, k-carriers as mcols object.

Examples

```
peak.path <- system.file("extdata/peaks/RData/peaksGRL_all_files.rds",
                         package="DEScan2")
grl <- readRDS(peak.path)
grl

regionsGR <- finalRegions(peakSamplesGRangesList=grl, zThreshold=1,
                      minCarriers=3, saveFlag=FALSE, verbose=TRUE)
```

`findOverlapsOverSamples` *findOverlapsOverSamples*

Description

given in input a GRangeList where each element is a sample computes the coverage extending a both direction window of prefixed length.

Usage

```
findOverlapsOverSamples(
  samplePeaksGRangelist,
  extendRegions = 200,
  minOverlap = 0L,
  maxGap = -1L,
  zThresh = 10,
  verbose = FALSE,
  scorecolname = "z-score",
  coverageFlag = FALSE
)
```

Arguments

`samplePeaksGRangelist`	given a granges list of samples finds the overlapping regions between them.
`extendRegions`	the number of bases to extend each region at its start and end.
`minOverlap`	the minimum overlap each peak needs to have. (see ChipPeakAnno::findOverlapsOfPeaks)
`maxGap`	the maximum gap admissible between the peaks. (see ChipPeakAnno::findOverlapsOfPeaks)
`zThresh`	a threshold value on z-score/scorecolname
`verbose`	verbose flag
`scorecolname`	character describing the name of the column within the peaks score.
`coverageFlag`	boolean indicating if to compute the scores in a coverage mode (sum of the reads of merged peak) or in a score mode (a normalized score across the merged peaks)

Value

a GRanges of peaks overlapped and unique between samples.

Examples

```
(peaks.file <- system.file("extdata/peaks/RData/peaksGRL_all_files.rds",
                           package="DEScan2"))
peaksGRLFiles <- readRDS(peaks.file)
(overlPeaks <- findOverlapsOverSamples(peaksGRLFiles))
```

`findPeaks` *findPeaks*

Description

This function calls peaks from bed or bam inputs using a variable window scan with a poisson model using the surrounding maxCompWinWidth (10kb) as background.

Usage

```
findPeaks(
  files,
  filetype = c("bam", "bed"),
  genomeName = NULL,
  binSize = 50,
  minWin = 50,
  maxWin = 1000,
  zthresh = 10,
  minCount = 0.1,
  minCompWinWidth = 5000,
  maxCompWinWidth = 10000,
  outputFolder = "Peaks",
  save = TRUE,
  force = TRUE,
  verbose = FALSE,
  sigwin = 10,
  onlyStdChrs = TRUE,
  chr = NULL,
  BPPARAM = BiocParallel::bpparam()
)
```

Arguments

`files`	Character vector containing paths of files to be analyzed.
`filetype`	Character, either "bam" or "bed" indicating format of input file.
`genomeName`	the code of the genome to use as reference for the input files. (cfr. constructBedRanges function parameters)

binSize	Integer size in bases of the minimum window for scanning, 50 is the default.
minWin	Integer indicating the minimum window size in bases notation.
maxWin	Integer indicating the maximum window size in bases notation.
zthresh	Cuttoff value for z-scores. Only windows with greater z-scores will be kept, default is 10.
minCount	A small constant (usually no larger than one) to be added to the counts prior to the log transformation to avoid problems with log(0).
minCompWinWidth	minimum bases width of a comparing window for Z-score.
maxCompWinWidth	maximum bases width of a comparing window for Z-score.
outputFolder	A string, Name of the folder to save the Peaks (optional) if the directory doesn't exist, it will be created. (Default is "Peaks")
save	Boolean, if TRUE files will be saved in a "./Peaks/chr*" directory created (if not already present) in the current working directory.
force	a boolean flag indicating if to force output overwriting.
verbose	if to show additional messages
sigwin	an integer value used to compute the length of the signal of a peak (default value is 10).
onlyStdChrs	a flag to work only with standard chromosomes. (cfr. constructBedRanges function parameters).
chr	if not NULL, a character like "chr#" indicating the chromosomes to use.
BPPARAM	object of class bpparamClass that specifies the back-end to be used for computations. See bpparam for details.

Value

A GRangesList where each element is a sample. Each GRanges represents the founded peaks and attached the z-score of the peak as mcols.

Examples

```
bam.files <- list.files(system.file("extdata/bam", package = "DEScan2"),
                        full.names = TRUE)

peaks <- findPeaks(files=bam.files[1], filetype="bam",
                        genomeName="mm9",
                        binSize=50, minWin=50, maxWin=1000,
                        zthresh=5, minCount=0.1, sigwin=10,
                        minCompWinWidth=5000, maxCompWinWidth=10000,
                        save=FALSE,
                        onlyStdChrs=TRUE,
                        chr=NULL,
                        verbose=FALSE)
head(peaks)
```

fromSamplesToChrsGRangesList *fromSamplesToChrsGRangesList*

Description

converts a GRangesList orgnized per samples to a GRangesList organized per Chromosomes where each element is a GRangesList of samples.

Usage

```
fromSamplesToChrsGRangesList(samplesGRangesList)
```

Arguments

samplesGRangesList
: a GRangesList of samples. Tipically generaed by findPeaks function.

Value

A GRangesList of chromosomes where each element is a GRanges list of samples.

Examples

```
library("GenomicRanges")
gr1 <- GRanges(
            seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
            ranges=IRanges(1:10, end=10),
            strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
            seqlengths=c(chr1=11, chr2=12, chr3=13))
gr2 <- GRanges(
            seqnames=Rle(c("chr1", "chr4", "chr1", "chr3"), c(1, 3, 2, 4)),
            ranges=IRanges(1:10, end=10),
            strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
            seqlengths=c(chr1=11, chr4=12, chr3=13))
sgrl <- GRangesList(gr1, gr2)
names(sgrl) <- c("samp1", "samp2")
(chrGrlSampGr <- fromSamplesToChrsGRangesList(sgrl))
```

keepRelevantChrs *keepRelevantChrs*

Description

subselect a list of GRanges created with cutGRangesPerChromosome returning only the relevant chromosomes GRanges.

Usage

```
keepRelevantChrs(chrGRangesList, chr = NULL)
```

Arguments

chrGRangesList	where each element is a chromosome, tipically created with cutGRangesPerChromosome.
chr	a character vector of chromosomes names of the form "chr#".

Value

the input chrGRangesList with only the relevat chromosomes.

Examples

```
library("GenomicRanges")
gr1 <- GRanges(
            seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
            ranges=IRanges(1:10, end=10),
            strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
            seqlengths=c(chr1=11, chr2=12, chr3=13))
grlc <- cutGRangesPerChromosome(gr1)
(grlChr <- keepRelevantChrs(grlc, c("chr1", "chr3")))
```

---

readBamAsBed *readBamAsBed*

Description

read a bam file into a bed like format. forcing UCSC format for chromosomes names.

Usage

```
readBamAsBed(file)
```

Arguments

file	Character indicating path to bam file.

Value

GRanges object.

Examples

```
files <- list.files(system.file("extdata/bam", package="DEScan2"),
                    full.names=TRUE)
gr <- readBamAsBed(files[1])
```

readBedFile *readBedFile*

Description

read a bed file into a GenomicRanges like format. forcing UCSC format for chromosomes names.

Usage

```
readBedFile(filename, arePeaks = FALSE)
```

Arguments

filename	the bed filename.
arePeaks	a flag indicating if the the bed file represents peaks.

Value

GRanges object

Examples

```
bedFile <- list.files(system.file("extdata/bed",package="DEScan2"),
                        full.names=TRUE)
gr <- readBedFile(bedFile)
```

readFilesAsGRangesList *readFilesAsGRangesList*

Description

Takes in input the path of bam/bed files to process and stores them in a GRangesList object, named with filePath/filenames. (for lazy people)

Usage

```
readFilesAsGRangesList(
  filePath,
  fileType = c("bam", "bed", "bed.zip", "narrow", "broad"),
  genomeName = NULL,
  onlyStdChrs = TRUE,
  arePeaks = TRUE,
  verbose = TRUE
)
```

Arguments

filePath	the path of input files.
fileType	the type of the files (bam/bed/bed.zip/narrow/broad).
genomeName	the genome code to associate to the files. (recommended) (i.e. "mm9", "hg17")
onlyStdChrs	a flag to keep only standard chromosomes.
arePeaks	a flag indicating if the files contain peaks.
verbose	verbose output flag.

Value

a GRangesList object

Examples

```
files.path <- system.file("extdata/bam", package="DEScan2")
grl <- readFilesAsGRangesList(filePath=files.path, fileType="bam",
                                genomeName="mm9", onlyStdChrs=TRUE,
                                verbose=TRUE)
class(grl)
names(grl)
grl
```

RleListToRleMatrix *RleListToRleMatrix*

Description

a wrapper to create a RleMatrix from a RleList object.

Usage

```
RleListToRleMatrix(RleList, dimnames = NULL)
```

Arguments

RleList	an RleList object with all elements of the same length.
dimnames	the names for dimensions of RleMatrix (see DelayedArray pkg).

Value

a RleMatrix from DelayedArray package.

Examples

```
library("DelayedArray")
lengths <-  c(3, 1, 2)
values <- c(15, 5, 20)
el1 <- S4Vectors::Rle(values=values, lengths=lengths)

el2 <- S4Vectors::Rle(values=sort(values), lengths=lengths)

rleList <- IRanges::RleList(el1, el2)
names(rleList) <- c("one", "two")
(rleMat <- RleListToRleMatrix(rleList))
```

---

saveGRangesAsBed *saveGRangesAsBed*

Description

save a GRanges object as bed file.

Usage

```
saveGRangesAsBed(
  GRanges,
  filepath = tempdir(),
  filename = tempfile(),
  force = FALSE,
  verbose = FALSE
)
```

Arguments

GRanges	the GRanges object.
filepath	the path to store the files.@
filename	the name to give to the files.
force	force overwriting.
verbose	verbose output flag.

Value

none

Examples

```
library("GenomicRanges")
gr <- GRanges(
        seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
        ranges=IRanges(1:10, end=10),
        strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
        seqlengths=c(chr1=11, chr2=12, chr3=13))

saveGRangesAsBed(GRanges=gr, filepath=tempdir(), filename=tempfile(),
                    verbose=TRUE)
```

---

saveGRangesAsTsv *saveGRangesAsTsv*

Description

save a GRanges object as tsv file.

Usage

```
saveGRangesAsTsv(
  GRanges,
  filepath = tempdir(),
  filename = tempfile(),
  col.names = NA,
  row.names = TRUE,
  sep = "\t",
  force = FALSE,
  verbose = FALSE
)
```

Arguments

GRanges	the GRanges object.
filepath	the path to store the files.
filename	the name to give to the files.
col.names	a logical value indicating whether the column names are to be written in the file, or a character vector indicating the column names, or NA for writing column names for writing a TAB for the column name of the row names, default is NA (see write.table).
row.names	a logical value indicating whether the row names are to be written in the file, or a character vector indicating the row names (see write.table).
sep	the column separator character (default is "\t").
force	force overwriting.
verbose	verbose output flag.

Value

none

Examples

```
gr <- GRanges(
        seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
        ranges=IRanges(1:10, end=10),
        strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
        seqlengths=c(chr1=11, chr2=12, chr3=13))
saveGRangesAsTsv(gr, verbose=TRUE)
```

`setGRGenomeInfo` *setGRGenomeInfo given a genome code (i.e. "mm9","mm10","hg19","hg38") retrieve the SeqInfo of that genome and assigns it to the input GRanges. Finally filters out those Infos not necessary to the GRanges.*

Description

setGRGenomeInfo given a genome code (i.e. "mm9","mm10","hg19","hg38") retrieve the SeqInfo of that genome and assigns it to the input GRanges. Finally filters out those Infos not necessary to the GRanges.

Usage

```
setGRGenomeInfo(GRanges, genomeName = NULL, verbose = FALSE)
```

Arguments

`GRanges`	a GRanges object.
`genomeName`	a genome code (i.e. "mm9")
`verbose`	verbose output

Value

a GRanges object with the seqinfo of the genome code

Examples

```
library("GenomicRanges")
gr <- GRanges(
        seqnames=Rle(c("chr1", "chr2", "chr1", "chr3"), c(1, 3, 2, 4)),
        ranges=IRanges(1:10, end=10),
        strand=Rle(strand(c("-", "+", "*", "+", "-")), c(1, 2, 2, 3, 2)),
        seqlengths=c(chr1=11, chr2=12, chr3=13))
mm9gr <- setGRGenomeInfo(GRanges=gr, genomeName="mm9", verbose=TRUE)
```

Index

∗ **internal.**
computeZ, 3

binnedCoverage, 2
bpparam, *9*, *12*

computeZ, 3
constructBedRanges, 4
countFinalRegions, 5
createGranges, 6
cutGRangesPerChromosome, 7

DEScan2, 8
divideEachSampleByChromosomes, 8

finalRegions, 9
findOverlapsOverSamples, 10
findPeaks, 11
fromSamplesToChrsGRangesList, 13

keepRelevantChrs, 13

readBamAsBed, 14
readBedFile, 15
readFilesAsGRangesList, 15
RleListToRleMatrix, 16

saveGRangesAsBed, 17
saveGRangesAsTsv, 18
setGRGenomeInfo, 19

write.table, *18*